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(54) Title: SIGNAL TRANSDUCTION GENES AND METHODS OF USE (57) Abstract <p>The invention provides isolated signal transduction nucleic acids and their encoded proteins. The present invention provides methods and compositions relating to altering signal transduction gene expression levels in plants. The invention further provides recombinant expression cassettes, host cells, transgenic plants, and antibody compositions.</p>		

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SIGNAL TRANSDUCTION GENES AND METHODS OF USE

TECHNICAL FIELD

The present invention relates generally to plant molecular biology. More specifically, it relates to nucleic acids and methods for modulating their expression in plants.

BACKGROUND OF THE INVENTION

Cells are constantly bombarded by external signals that regulate their growth, differentiation, and stress level. To respond properly to these signals, eukaryotic cells assemble cascades of highly conserved protein kinases (mitogen-activated protein kinases, MAPKs, and their activator kinases), which form the central elements of signal transduction pathways that lead to and activate transcription factors in the nucleus and other effectors throughout the cell. (Elion, E.A. *Routing MAP Kinase Cascades*. *Science*, Vol. 281 (11 September 1998), pp. 1625-1626)

Plants' sessile nature requires strategies to adapt to changes in their local environment. Responses to signals such as pathogen attack, herbivore feeding, or changes in light, temperature, or nutrient availability often involve phytohormones such as ethylene. MAP kinase modules appear to be involved in ethylene signaling and auxin-induced cell proliferation. Upstream and downstream regulatory components suggest that MAP kinases may serve as a central point in a signal transduction network where different upstream pathways converge and may distribute the signals to different downstream targets. (Jonak, C., et al. *MAP kinases: universal multi-purpose signaling tools*. *Plant Molecular Biology* Vol. 24 (1994), pp. 407-416.)

DESCRIPTION OF THE INVENTION

Overview

Generally, it is the object of the present invention to provide nucleic acids and proteins relating to signal transduction genes. It is an object of the present invention to provide transgenic plants comprising the nucleic acids of the present invention, and methods for modulating, in a transgenic plant, expression of the nucleic acids of the present invention.

Therefore, in one aspect the present invention relates to an isolated nucleic acid comprising a member selected from the group consisting of (a) a polynucleotide having a

specified sequence identity to a polynucleotide encoding a polypeptide of the present invention; (b) a polynucleotide which is complementary to the polynucleotide of (a); and, (c) a polynucleotide comprising a specified number of contiguous nucleotides from a polynucleotide of (a) or (b). The isolated nucleic acid can be DNA.

5 In other aspects the present invention relates to: 1) recombinant expression cassettes, comprising a nucleic acid of the present invention operably linked to a promoter, 2) a host cell into which has been introduced the recombinant expression cassette, and 3) a transgenic plant comprising the recombinant expression cassette. The host cell and plant are optionally from maize, wheat, rice, or soybean.

10 Unless otherwise stated, the polynucleotide and polypeptide sequences identified in the accompanying Sequence Listing represent polynucleotides and polypeptides of the present invention. A nucleic acid of the present invention comprises a polynucleotide of the present invention. A protein of the present invention comprises a polypeptide of the present invention.

15 The present invention provides utility in such exemplary applications as regulating expression of phytohormones, including ethylene, auxins, cytokinins, and gibberellin, in order to effect developmental changes in plants and provide control of plant response to environmental stresses. Various preferred embodiments are disclosed throughout the specification.

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Definitions

Units, prefixes, and symbols may be denoted in their SI accepted form. Unless otherwise indicated, nucleic acids are written left to right in 5' to 3' orientation; amino acid sequences are written left to right in amino to carboxy orientation, respectively. Numeric
25 ranges recited within the specification are inclusive of the numbers defining the range and include each integer within the defined range. Amino acids may be referred to herein by either their commonly known three letter symbols or by the one-letter symbols recommended by the IUPAC-IUBMB Nomenclature Commission. Nucleotides, likewise, may be referred to by their commonly accepted single-letter codes. Unless otherwise
30 provided for, software, electrical, and electronics terms as used herein are as defined in The New IEEE Standard Dictionary of Electrical and Electronics Terms (5th edition, 1993). The terms defined below are more fully defined by reference to the specification as a whole. Section headings provided throughout the specification are not limitations to the various objects and embodiments of the present invention.

By "amplified" is meant the construction of multiple copies of a nucleic acid sequence or multiple copies complementary to the nucleic acid sequence using at least one of the nucleic acid sequences as a template. Amplification systems include the polymerase chain reaction (PCR) system, ligase chain reaction (LCR) system, nucleic acid sequence based amplification (NASBA, Cingene, Mississauga, Ontario), Q-Beta Replicase systems, transcription-based amplification system (TAS), and strand displacement amplification (SDA). See, e.g., *Diagnostic Molecular Microbiology: Principles and Applications*, D. H. Persing *et al.*, Ed., American Society for Microbiology, Washington, D.C. (1993). The product of amplification is termed an amplicon.

As used herein, "antisense orientation" includes reference to a duplex polynucleotide sequence that is operably linked to a promoter in an orientation where the antisense strand is transcribed. The antisense strand is sufficiently complementary to an endogenous transcription product such that translation of the endogenous transcription product is often inhibited.

By "encoding" or "encoded", with respect to a specified nucleic acid, is meant comprising the information for translation into the specified protein. A nucleic acid encoding a protein may comprise non-translated sequences (e.g., introns) within translated regions of the nucleic acid, or may lack such intervening non-translated sequences (e.g., as in cDNA). The information by which a protein is encoded is specified by the use of codons. Typically, the amino acid sequence is encoded by the nucleic acid using the "universal" genetic code. However, variants of the universal code, such as are present in some plant, animal, and fungal mitochondria, the bacterium *Mycoplasma capricolum*, or the ciliate *Macronucleus*, may be used when the nucleic acid is expressed therein.

When the nucleic acid is prepared or altered synthetically, advantage can be taken of known codon preferences of the intended host where the nucleic acid is to be expressed. For example, although nucleic acid sequences of the present invention may be expressed in both monocotyledonous and dicotyledonous plant species, sequences can be modified to account for the specific codon preferences and GC content preferences of monocotyledons or dicotyledons as these preferences have been shown to differ (Murray *et al. Nucl. Acids Res.* 17: 477-498 (1989)). Thus, the maize preferred codon for a particular amino acid may be derived from known gene sequences from maize. Maize codon usage for 28 genes from maize plants is listed in Table 4 of Murray *et al., supra*.

As used herein "full-length sequence" in reference to a specified polynucleotide or its encoded protein means having the entire amino acid sequence of, a native (non-

synthetic), endogenous, biologically (e.g., structurally or catalytically) active form of the specified protein. Methods to determine whether a sequence is full-length are well known in the art including such exemplary techniques as northern or western blots, primer extension, S1 protection, and ribonuclease protection. See, e.g., *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997). Comparison to known full-length homologous (orthologous and/or paralogous) sequences can also be used to identify full-length sequences of the present invention. Additionally, consensus sequences typically present at the 5' and 3' untranslated regions of mRNA aid in the identification of a polynucleotide as full-length. For example, the consensus sequence ANNNNAUGG, where the underlined codon represents the N-terminal methionine, aids in determining whether the polynucleotide has a complete 5' end. Consensus sequences at the 3' end, such as polyadenylation sequences, aid in determining whether the polynucleotide has a complete 3' end.

As used herein, "heterologous" in reference to a nucleic acid is a nucleic acid that originates from a foreign species, or, if from the same species, is substantially modified from its native form in composition and/or genomic locus by human intervention. For example, a promoter operably linked to a heterologous structural gene is from a species different from that from which the structural gene was derived, or, if from the same species, one or both are substantially modified from their original form. A heterologous protein may originate from a foreign species or, if from the same species, is substantially modified from its original form by human intervention.

By "host cell" is meant a cell which contains a vector and supports the replication and/or expression of the vector. Host cells may be prokaryotic cells such as *E. coli*, or eukaryotic cells such as yeast, insect, amphibian, or mammalian cells. Preferably, host cells are monocotyledonous or dicotyledonous plant cells. A particularly preferred monocotyledonous host cell is a maize host cell.

The term "introduced" includes reference to the incorporation of a nucleic acid into a eukaryotic or prokaryotic cell where the nucleic acid may be incorporated into the genome of the cell (e.g., chromosome, plasmid, plastid or mitochondrial DNA), converted into an autonomous replicon, or transiently expressed (e.g., transfected mRNA). The term includes such nucleic acid introduction means as "transfection", "transformation" and "transduction".

The term "isolated" refers to material, such as a nucleic acid or a protein, which is: (1) substantially or essentially free from components which normally accompany or

interact with it as found in its natural environment. The isolated material optionally comprises material not found with the material in its natural environment; or (2) if the material is in its natural environment, the material has been synthetically altered or synthetically produced by deliberate human intervention and/or placed at a different location within the cell. The synthetic alteration or creation of the material can be performed on the material within or apart from its natural state. For example, a naturally-occurring nucleic acid becomes an isolated nucleic acid if it is altered or produced by non-natural, synthetic methods, or if it is transcribed from DNA which has been altered or produced by non-natural, synthetic methods. The isolated nucleic acid may also be produced by the synthetic re-arrangement ("shuffling") of a part or parts of one or more allelic forms of the gene of interest. Likewise, a naturally-occurring nucleic acid (e.g., a promoter) becomes isolated if it is introduced to a different locus of the genome. Nucleic acids which are "isolated," as defined herein, are also referred to as "heterologous" nucleic acids. See, e.g., Compounds and Methods for Site Directed Mutagenesis in Eukaryotic Cells, Kmiec, U.S. Patent No. 5,565,350; *In Vivo* Homologous Sequence Targeting in Eukaryotic Cells; Zarling *et al.*, WO 93/22443; Methods for In Vitro Recombination, Stemmer, U.S. Patent No. 5,605,793.

As used herein, "nucleic acid" includes reference to a deoxyribonucleotide or ribonucleotide polymer, or chimeras thereof, in either single- or double-stranded form, and unless otherwise limited, encompasses known analogues having the essential nature of natural nucleotides in that they hybridize to single-stranded nucleic acids in a manner similar to naturally occurring nucleotides (e.g., peptide nucleic acids).

Unless otherwise stated, a "signal transduction nucleic acid" is a nucleic acid of the present invention and means a nucleic acid comprising a polynucleotide of the present invention (a "signal transduction polynucleotide") encoding a signal transduction polypeptide. A "signal transduction gene" is a gene of the present invention and refers to a full-length signal transduction polynucleotide.

By "nucleic acid library" is meant a collection of isolated DNA or RNA molecules which comprise and substantially represent the entire transcribed fraction of a genome of a specified organism, tissue, or of a cell type from that organism. Construction of exemplary nucleic acid libraries, such as genomic and cDNA libraries, is taught in standard molecular biology references such as Berger and Kimmel, *Guide to Molecular Cloning Techniques, Methods in Enzymology*, Vol. 152, Academic Press, Inc., San Diego, CA (Berger); Sambrook *et al.*, *Molecular Cloning - A Laboratory Manual*, 2nd ed., Vol. 1-3 (1989); and

Current Protocols in Molecular Biology, F.M. Ausubel *et al.*, Eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (1994).

As used herein "operably linked" includes reference to a functional linkage between a promoter and a second sequence, wherein the promoter sequence initiates and mediates transcription of the DNA sequence corresponding to the second sequence. Generally, operably linked means that the nucleic acid sequences being linked are contiguous and, where necessary to join two protein coding regions, contiguous and in the same reading frame.

As used herein, the term "plant" includes reference to whole plants, plant parts or organs (e.g., leaves, stems, roots, etc.), plant cells, seeds and progeny of same. Plant cell, as used herein, further includes, without limitation, cells obtained from or found in: seeds, suspension cultures, embryos, meristematic regions, callus tissue, leaves, roots, shoots, gametophytes, sporophytes, pollen, and microspores. Plant cells can also be understood to include modified cells, such as protoplasts, obtained from the aforementioned tissues. The class of plants which can be used in the methods of the invention is generally as broad as the class of higher plants amenable to transformation techniques, including both monocotyledonous and dicotyledonous plants. A particularly preferred plant is *Zea mays*.

As used herein, "polynucleotide" includes reference to a deoxyribopolynucleotide, ribopolynucleotide, or chimeras or analogs thereof that have the essential nature of a natural deoxy- or ribo- nucleotide in that they hybridize, under stringent hybridization conditions, to substantially the same nucleotide sequence as naturally occurring nucleotides and/or allow translation into the same amino acid(s) as the naturally occurring nucleotide(s). A polynucleotide can be full-length or a subsequence of a native or heterologous structural or regulatory gene. Unless otherwise indicated, the term includes reference to the specified sequence as well as the complementary sequence thereof. Thus, DNAs or RNAs with backbones modified for stability or for other reasons are "polynucleotides" as that term is intended herein. Moreover, DNAs or RNAs comprising unusual bases, such as inosine, or modified bases, such as tritylated bases, to name just two examples, are polynucleotides as the term is used herein. It will be appreciated that a great variety of modifications have been made to DNA and RNA that serve many useful purposes known to those of skill in the art. The term polynucleotide as it is employed herein embraces such chemically, enzymatically or metabolically modified forms of polynucleotides, as well

as the chemical forms of DNA and RNA characteristic of viruses and cells, including among other things, simple and complex cells.

The terms "polypeptide", "peptide" and "protein" are used interchangeably herein to refer to a polymer of amino acid residues. The terms apply to amino acid polymers in which one or more amino acid residue is an artificial chemical analogue of a corresponding naturally occurring amino acid, as well as to naturally occurring amino acid polymers. The essential nature of such analogues of naturally occurring amino acids is that, when incorporated into a protein, that protein is specifically reactive to antibodies elicited to the same protein but consisting entirely of naturally occurring amino acids. The terms "polypeptide", "peptide" and "protein" are also inclusive of modifications including, but not limited to, glycosylation, lipid attachment, sulfation, gamma-carboxylation of glutamic acid residues, hydroxylation and ADP-ribosylation. Further, this invention contemplates the use of both the methionine-containing and the methionine-less amino terminal variants of the protein of the invention.

As used herein "promoter" includes reference to a region of DNA upstream from the start of transcription and involved in recognition and binding of RNA polymerase and other proteins to initiate transcription. A "plant promoter" is a promoter capable of initiating transcription in plant cells whether or not its origin is a plant cell. Exemplary plant promoters include, but are not limited to, those that are obtained from plants, plant viruses, and bacteria which comprise genes expressed in plant cells such as *Agrobacterium* or *Rhizobium*. Examples of promoters under developmental control include promoters that preferentially initiate transcription in certain tissues, such as leaves, roots, or seeds. Such promoters are referred to as "tissue preferred". Promoters which initiate transcription only in certain tissue are referred to as "tissue specific". A "cell type" specific promoter primarily drives expression in certain cell types in one or more organs, for example, vascular cells in roots or leaves. An "inducible" or "repressible" promoter is a promoter which is under environmental control. Examples of environmental conditions that may effect transcription by inducible promoters include anaerobic conditions or the presence of light. Tissue specific, tissue preferred, cell type specific, and inducible promoters constitute the class of "non-constitutive" promoters. A "constitutive" promoter is a promoter which is active under most environmental conditions.

As used herein "recombinant" includes reference to a cell or vector, that has been modified by the introduction of a heterologous nucleic acid or that the cell is derived from a cell so modified. Thus, for example, recombinant cells express genes that are not found

in identical form within the native (non-recombinant) form of the cell or express native genes that are otherwise abnormally expressed, under-expressed or not expressed at all as a result of human intervention. The term "recombinant" as used herein does not encompass the alteration of the cell or vector by naturally occurring events (e.g., spontaneous
5 mutation, natural transformation/transduction/transposition) such as those occurring without human intervention.

As used herein, a "recombinant expression cassette" is a nucleic acid construct, generated recombinantly or synthetically, with a series of specified nucleic acid elements which permit transcription of a particular nucleic acid in a host cell. The recombinant
10 expression cassette can be incorporated into a plasmid, chromosome, mitochondrial DNA, plastid DNA, virus, or nucleic acid fragment. Typically, the recombinant expression cassette portion of an expression vector includes, among other sequences, a nucleic acid to be transcribed, and a promoter.

The term "residue" or "amino acid residue" or "amino acid" are used
15 interchangeably herein to refer to an amino acid that is incorporated into a protein, polypeptide, or peptide (collectively "protein"). The amino acid may be a naturally occurring amino acid and, unless otherwise limited, may encompass non-natural analogs of natural amino acids that can function in a similar manner as naturally occurring amino acids.

The term "selectively hybridizes" includes reference to hybridization, under
20 stringent hybridization conditions, of a nucleic acid sequence to a specified nucleic acid target sequence to a detectably greater degree (e.g., at least 2-fold over background) than its hybridization to non-target nucleic acid sequences and to the substantial exclusion of non-target nucleic acids. Selectively hybridizing sequences typically have about at least
25 80% sequence identity, preferably 90% sequence identity, and most preferably 100% sequence identity (i.e., complementary) with each other.

The term "stringent conditions" or "stringent hybridization conditions" includes reference to conditions under which a probe will selectively hybridize to its target sequence, to a detectably greater degree than to other sequences (e.g., at least 2-fold over
30 background). Stringent conditions are sequence-dependent and will be different in different circumstances. By controlling the stringency of the hybridization and/or washing conditions, target sequences can be identified which are 100% complementary to the probe (homologous probing). Alternatively, stringency conditions can be adjusted to allow some mismatching in sequences so that lower degrees of similarity are detected (heterologous

stringency of hybridization and/or wash solutions are inherently described. If the desired degree of mismatching results in a T_m of less than 45 °C (aqueous solution) or 32 °C (formamide solution) it is preferred to increase the SSC concentration so that a higher temperature can be used. Hybridization and/or wash conditions can be applied for at least 10, 30, 60, 90, 120, or 240 minutes. An extensive guide to the hybridization of nucleic acids is found in Tijssen, *Laboratory Techniques in Biochemistry and Molecular Biology--Hybridization with Nucleic Acid Probes*, Part I, Chapter 2 "Overview of principles of hybridization and the strategy of nucleic acid probe assays", Elsevier, New York (1993); and *Current Protocols in Molecular Biology*, Chapter 2, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995).

As used herein, "transgenic plant" includes reference to a plant which comprises within its genome a heterologous polynucleotide. Generally, the heterologous polynucleotide is stably integrated within the genome such that the polynucleotide is passed on to successive generations. The heterologous polynucleotide may be integrated into the genome alone or as part of a recombinant expression cassette. "Transgenic" is used herein to include any cell, cell line, callus, tissue, plant part or plant, the genotype of which has been altered by the presence of heterologous nucleic acid including those transgenics initially so altered as well as those created by sexual crosses or asexual propagation from the initial transgenic. The term "transgenic" as used herein does not encompass the alteration of the genome (chromosomal or extra-chromosomal) by conventional plant breeding methods or by naturally occurring events such as random cross-fertilization, non-recombinant viral infection, non-recombinant bacterial transformation, non-recombinant transposition, or spontaneous mutation.

As used herein, "vector" includes reference to a nucleic acid used in introduction of a polynucleotide of the present invention into a host cell. Vectors are often replicons. Expression vectors permit transcription of a nucleic acid inserted therein.

The following terms are used to describe the sequence relationships between a polynucleotide/polypeptide of the present invention with a reference polynucleotide/polypeptide: (a) "reference sequence", (b) "comparison window", (c) "sequence identity", and (d) "percentage of sequence identity".

(a) As used herein, "reference sequence" is a defined sequence used as a basis for sequence comparison with a polynucleotide/polypeptide of the present invention. A reference sequence may be a subset or the entirety of a specified sequence; for example, as

probing). Generally, a probe is less than about 1000 nucleotides in length, optionally less than 500 nucleotides in length.

Typically, stringent conditions will be those in which the salt concentration is less than about 1.5 M Na ion, typically about 0.01 to 1.0 M Na ion concentration (or other salts) at pH 7.0 to 8.3 and the temperature is at least about 30°C for short probes (*e.g.*, 10 to 50 nucleotides) and at least about 60°C for long probes (*e.g.*, greater than 50 nucleotides). Stringent conditions may also be achieved with the addition of destabilizing agents such as formamide. Exemplary low stringency conditions include hybridization with a buffer solution of 30 to 35% formamide, 1 M NaCl, 1% SDS (sodium dodecyl sulphate) at 37°C, and a wash in 1X to 2X SSC (20X SSC = 3.0 M NaCl/0.3 M trisodium citrate) at 50 to 55°C. Exemplary moderate stringency conditions include hybridization in 40 to 45% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.5X to 1X SSC at 55 to 60°C. Exemplary high stringency conditions include hybridization in 50% formamide, 1 M NaCl, 1% SDS at 37°C, and a wash in 0.1X SSC at 60 to 65°C.

Specificity is typically the function of post-hybridization washes, the critical factors being the ionic strength and temperature of the final wash solution. For DNA-DNA hybrids, the T_m can be approximated from the equation of Meinkoth and Wahl, *Anal. Biochem.*, 138:267-284 (1984): $T_m = 81.5^\circ\text{C} + 16.6 (\log M) + 0.41 (\%GC) - 0.61 (\% \text{ form}) - 500/L$; where M is the molarity of monovalent cations, %GC is the percentage of guanosine and cytosine nucleotides in the DNA, % form is the percentage of formamide in the hybridization solution, and L is the length of the hybrid in base pairs. The T_m is the temperature (under defined ionic strength and pH) at which 50% of a complementary target sequence hybridizes to a perfectly matched probe. T_m is reduced by about 1°C for each 1% of mismatching; thus, T_m , hybridization and/or wash conditions can be adjusted to hybridize to sequences of the desired identity. For example, if sequences with $\geq 90\%$ identity are sought, the T_m can be decreased 10°C. Generally, stringent conditions are selected to be about 5°C lower than the thermal melting point (T_m) for the specific sequence and its complement at a defined ionic strength and pH. However, severely stringent conditions can utilize a hybridization and/or wash at 1, 2, 3, or 4 °C lower than the thermal melting point (T_m); moderately stringent conditions can utilize a hybridization and/or wash at 6, 7, 8, 9, or 10 °C lower than the thermal melting point (T_m); low stringency conditions can utilize a hybridization and/or wash at 11, 12, 13, 14, 15, or 20 °C lower than the thermal melting point (T_m). Using the equation, hybridization and wash compositions, and desired T_m , those of ordinary skill will understand that variations in the

a segment of a full-length cDNA or gene sequence, or the complete cDNA or gene sequence.

(b) As used herein, "comparison window" includes reference to a contiguous and specified segment of a polynucleotide/polypeptide sequence, wherein the polynucleotide/polypeptide sequence may be compared to a reference sequence and wherein the portion of the polynucleotide/polypeptide sequence in the comparison window may comprise additions or deletions (i.e., gaps) compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. Generally, the comparison window is at least 20 contiguous nucleotides/amino acids residues in length, and optionally can be 30, 40, 50, 100, or longer. Those of skill in the art understand that to avoid a high similarity to a reference sequence due to inclusion of gaps in the polynucleotide/polypeptide sequence, a gap penalty is typically introduced and is subtracted from the number of matches.

Methods of alignment of sequences for comparison are well-known in the art. Optimal alignment of sequences for comparison may be conducted by the local homology algorithm of Smith and Waterman, *Adv. Appl. Math.* 2: 482 (1981); by the homology alignment algorithm of Needleman and Wunsch, *J. Mol. Biol.* 48: 443 (1970); by the search for similarity method of Pearson and Lipman, *Proc. Natl. Acad. Sci.* 85: 2444 (1988); by computerized implementations of these algorithms, including, but not limited to: CLUSTAL in the PC/Gene program by Intelligenetics, Mountain View, California; GAP, BESTFIT, BLAST, FASTA, and TFASTA in the Wisconsin Genetics Software Package, Genetics Computer Group (GCG), 575 Science Dr., Madison, Wisconsin, USA; the CLUSTAL program is well described by Higgins and Sharp, *Gene* 73: 237-244 (1988); Higgins and Sharp, *CABIOS* 5: 151-153 (1989); Corpet, *et al.*, *Nucleic Acids Research* 16: 10881-90 (1988); Huang, *et al.*, *Computer Applications in the Biosciences* 8: 155-65 (1992), and Pearson, *et al.*, *Methods in Molecular Biology* 24: 307-331 (1994).

The BLAST family of programs which can be used for database similarity searches includes: BLASTN for nucleotide query sequences against nucleotide database sequences; BLASTX for nucleotide query sequences against protein database sequences; BLASTP for protein query sequences against protein database sequences; TBLASTN for protein query sequences against nucleotide database sequences; and TBLASTX for nucleotide query sequences against nucleotide database sequences. See, *Current Protocols in Molecular Biology*, Chapter 19, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New

York (1995); Altschul *et al.*, *J. Mol. Biol.*, 215:403-410 (1990); and, Altschul *et al.*, *Nucleic Acids Res.* 25:3389-3402 (1997).

Software for performing BLAST analyses is publicly available, e.g., through the National Center for Biotechnology Information (<http://www.ncbi.nlm.nih.gov/>). This algorithm involves first identifying high scoring sequence pairs (HSPs) by identifying short words of length W in the query sequence, which either match or satisfy some positive-valued threshold score T when aligned with a word of the same length in a database sequence. T is referred to as the neighborhood word score threshold. These initial neighborhood word hits act as seeds for initiating searches to find longer HSPs containing them. The word hits are then extended in both directions along each sequence for as far as the cumulative alignment score can be increased. Cumulative scores are calculated using, for nucleotide sequences, the parameters M (reward score for a pair of matching residues; always > 0) and N (penalty score for mismatching residues; always < 0). For amino acid sequences, a scoring matrix is used to calculate the cumulative score. Extension of the word hits in each direction are halted when: the cumulative alignment score falls off by the quantity X from its maximum achieved value; the cumulative score goes to zero or below, due to the accumulation of one or more negative-scoring residue alignments; or the end of either sequence is reached. The BLAST algorithm parameters W, T, and X determine the sensitivity and speed of the alignment. The BLASTN program (for nucleotide sequences) uses as defaults a wordlength (W) of 11, an expectation (E) of 10, a cutoff of 100, M=5, N=-4, and a comparison of both strands. For amino acid sequences, the BLASTP program uses as defaults a wordlength (W) of 3, an expectation (E) of 10, and the BLOSUM62 scoring matrix (*see* Henikoff & Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

In addition to calculating percent sequence identity, the BLAST algorithm also performs a statistical analysis of the similarity between two sequences (*see, e.g.,* Karlin & Altschul, *Proc. Nat'l. Acad. Sci. USA* 90:5873-5877 (1993)). One measure of similarity provided by the BLAST algorithm is the smallest sum probability (P(N)), which provides an indication of the probability by which a match between two nucleotide or amino acid sequences would occur by chance.

BLAST searches assume that proteins can be modeled as random sequences. However, many real proteins comprise regions of nonrandom sequences which may be homopolymeric tracts, short-period repeats, or regions enriched in one or more amino acids. Such low-complexity regions may be aligned between unrelated proteins even

though other regions of the protein are entirely dissimilar. A number of low-complexity filter programs can be employed to reduce such low-complexity alignments. For example, the SEG (Wooten and Federhen, *Comput. Chem.*, 17:149-163 (1993)) and XNU (Claverie and States, *Comput. Chem.*, 17:191-201 (1993)). low-complexity filters can be employed alone or in combination.

Unless otherwise stated, nucleotide and protein identity/similarity values provided herein are calculated using GAP (GCG Version 10) under default values.

GAP (Global Alignment Program) can also be used to compare a polynucleotide or polypeptide of the present invention with a reference sequence. GAP uses the algorithm of Needleman and Wunsch (*J. Mol. Biol.* 48: 443-453, 1970) to find the alignment of two complete sequences that maximizes the number of matches and minimizes the number of gaps. GAP considers all possible alignments and gap positions and creates the alignment with the largest number of matched bases and the fewest gaps. It allows for the provision of a gap creation penalty and a gap extension penalty in units of matched bases. GAP must make a profit of gap creation penalty number of matches for each gap it inserts. If a gap extension penalty greater than zero is chosen, GAP must, in addition, make a profit for each gap inserted of the length of the gap times the gap extension penalty. Default gap creation penalty values and gap extension penalty values in Version 10 of the Wisconsin Genetics Software Package for protein sequences are 8 and 2, respectively. For nucleotide sequences the default gap creation penalty is 50 while the default gap extension penalty is 3. The gap creation and gap extension penalties can be expressed as an integer selected from the group of integers consisting of from 0 to 100. Thus, for example, the gap creation and gap extension penalties can each independently be: 0, 1, 2, 3, 4, 5, 6, 7, 8, 9, 10, 15, 20, 30, 40, 50, 60 or greater.

GAP presents one member of the family of best alignments. There may be many members of this family, but no other member has a better quality. GAP displays four figures of merit for alignments: Quality, Ratio, Identity, and Similarity. The Quality is the metric maximized in order to align the sequences. Ratio is the quality divided by the number of bases in the shorter segment. Percent Identity is the percent of the symbols that actually match. Percent Similarity is the percent of the symbols that are similar. Symbols that are across from gaps are ignored. A similarity is scored when the scoring matrix value for a pair of symbols is greater than or equal to 0.50, the similarity threshold. The scoring matrix used in Version 10 of the Wisconsin Genetics Software Package is BLOSUM62 (see Henikoff & Henikoff (1989) *Proc. Natl. Acad. Sci. USA* 89:10915).

Multiple alignment of the sequences can be performed using the CLUSTAL method of alignment (Higgins and Sharp (1989) *CABIOS*, 5:151-153) with the default parameters (GAP PENALTY=10, GAP LENGTH PENALTY=10). Default parameters for pairwise alignments using the CLUSTAL method are KTUPLE 1, GAP PENALTY=3, WINDOW=5 and DIAGONALS SAVED=5.

(c) As used herein, "sequence identity" or "identity" in the context of two nucleic acid or polypeptide sequences includes reference to the residues in the two sequences which are the same when aligned for maximum correspondence over a specified comparison window. When percentage of sequence identity is used in reference to proteins it is recognized that residue positions which are not identical often differ by conservative amino acid substitutions, where amino acid residues are substituted for other amino acid residues with similar chemical properties (e.g. charge or hydrophobicity) and therefore do not change the functional properties of the molecule. Where sequences differ in conservative substitutions, the percent sequence identity may be adjusted upwards to correct for the conservative nature of the substitution. Sequences which differ by such conservative substitutions are said to have "sequence similarity" or "similarity". Means for making this adjustment are well-known to those of skill in the art. Typically this involves scoring a conservative substitution as a partial rather than a full mismatch, thereby increasing the percentage sequence identity. Thus, for example, where an identical amino acid is given a score of 1 and a non-conservative substitution is given a score of zero, a conservative substitution is given a score between zero and 1. The scoring of conservative substitutions is calculated, e.g., according to the algorithm of Meyers and Miller, *Computer Applic. Biol. Sci.*, 4: 11-17 (1988) e.g., as implemented in the program PC/GENE (Intelligenetics, Mountain View, California, USA).

(d) As used herein, "percentage of sequence identity" means the value determined by comparing two optimally aligned sequences over a comparison window, wherein the portion of the polynucleotide sequence in the comparison window may comprise additions or deletions (i.e., gaps) as compared to the reference sequence (which does not comprise additions or deletions) for optimal alignment of the two sequences. The percentage is calculated by determining the number of positions at which the identical nucleic acid base or amino acid residue occurs in both sequences to yield the number of matched positions, dividing the number of matched positions by the total number of positions in the window of comparison and multiplying the result by 100 to yield the percentage of sequence identity.

Utilities

The present invention provides, among other things, compositions and methods for modulating (i.e., increasing or decreasing) the level of polynucleotides and polypeptides of the present invention in plants. In particular, the polynucleotides and polypeptides of the present invention can be expressed temporally or spatially, e.g., at developmental stages, in tissues, and/or in quantities, which are uncharacteristic of non-recombinantly engineered plants.

The present invention also provides isolated nucleic acids comprising polynucleotides of sufficient length and complementarity to a polynucleotide of the present invention to use as probes or amplification primers in the detection, quantitation, or isolation of gene transcripts. For example, isolated nucleic acids of the present invention can be used as probes in detecting deficiencies in the level of mRNA in screenings for desired transgenic plants, for detecting mutations in the gene (e.g., substitutions, deletions, or additions), for monitoring upregulation of expression or changes in enzyme activity in screening assays of compounds, for detection of any number of allelic variants (polymorphisms), orthologs, or paralogs of the gene, or for site directed mutagenesis in eukaryotic cells (see, e.g., U.S. Patent No. 5,565,350). The isolated nucleic acids of the present invention can also be used for recombinant expression of their encoded polypeptides, or for use as immunogens in the preparation and/or screening of antibodies. The isolated nucleic acids of the present invention can also be employed for use in sense or antisense suppression of one or more genes of the present invention in a host cell, tissue, or plant. Attachment of chemical agents which bind, intercalate, cleave and/or crosslink to the isolated nucleic acids of the present invention can also be used to modulate transcription or translation.

The present invention also provides isolated proteins comprising a polypeptide of the present invention (e.g., preproenzyme, proenzyme, or enzymes). The present invention also provides proteins comprising at least one epitope from a polypeptide of the present invention. The proteins of the present invention can be employed in assays for enzyme agonists or antagonists of enzyme function, or for use as immunogens or antigens to obtain antibodies specifically immunoreactive with a protein of the present invention. Such antibodies can be used in assays for expression levels, for identifying and/or isolating nucleic acids of the present invention from expression libraries, for identification of

homologous polypeptides from other species, or for purification of polypeptides of the present invention.

The isolated nucleic acids and polypeptides of the present invention can be used over a broad range of plant types, particularly monocots such as the species of the family
5 *Gramineae* including *Hordeum*, *Secale*, *Oryza*, *Triticum*, *Sorghum* (e.g., *S. bicolor*) and *Zea* (e.g., *Z. mays*), and dicots such as *Glycine*.

The isolated nucleic acid and proteins of the present invention can also be used in species from the genera: *Cucurbita*, *Rosa*, *Vitis*, *Juglans*, *Fragaria*, *Lotus*, *Medicago*,
Onobrychis, *Trifolium*, *Trigonella*, *Vigna*, *Citrus*, *Linum*, *Geranium*, *Manihot*, *Daucus*,
10 *Arabidopsis*, *Brassica*, *Raphanus*, *Sinapis*, *Atropa*, *Capsicum*, *Datura*, *Hyoscyamus*,
Lycopersicon, *Nicotiana*, *Solanum*, *Petunia*, *Digitalis*, *Majorana*, *Cichorium*, *Helianthus*,
Lactuca, *Bromus*, *Asparagus*, *Antirrhinum*, *Heterocallis*, *Nemesis*, *Pelargonium*, *Panicum*,
Pennisetum, *Ranunculus*, *Senecio*, *Salpiglossis*, *Cucumis*, *Browallia*, *Pisum*, *Phaseolus*,
Lolium, and *Avena*.

Nucleic Acids

The present invention provides, among other things, isolated nucleic acids of RNA, DNA, and analogs and/or chimeras thereof, comprising a polynucleotide of the present invention.

20 A polynucleotide of the present invention is inclusive of those in the accompanying Sequence Listing and:

(a) an isolated polynucleotide encoding a polypeptide of the present invention, such as those polypeptides included in the accompanying Sequence Listing, including exemplary polynucleotides of the present invention;

25 (b) an isolated polynucleotide which is the product of amplification from a plant nucleic acid library using primer pairs which selectively hybridize under stringent conditions to loci within a polynucleotide of the present invention;

(c) an isolated polynucleotide which selectively hybridizes to a polynucleotide of (a) or (b);

30 (d) an isolated polynucleotide having a specified sequence identity with polynucleotides of (a), (b), or (c);

(e) an isolated polynucleotide encoding a protein having a specified number of contiguous amino acids from a prototype polypeptide, wherein the protein is specifically recognized by antisera elicited by presentation of the protein and wherein the protein does

not detectably immunoreact to antisera which has been fully immunosorbed with the protein;

(f) complementary sequences of polynucleotides of (a), (b), (c), (d), or (e); and

(g) an isolated polynucleotide comprising at least a specific number of contiguous
5 nucleotides from a polynucleotide of (a), (b), (c), (d), (e), or (f);

(h) an isolated polynucleotide from a full-length enriched cDNA library having the physico-chemical property of selectively hybridizing to a polynucleotide of (a), (b), (c), (d), (e), (f), or (g);

(i) an isolated polynucleotide made by the process of: 1) providing a full-length
10 enriched nucleic acid library, 2) selectively hybridizing the polynucleotide to a polynucleotide of (a), (b), (c), (d), (e), (f), (g), or (h), thereby isolating the polynucleotide from the nucleic acid library.

A. Polynucleotides Encoding A Polypeptide of the Present Invention

15 As indicated in (a), above, the present invention provides isolated nucleic acids comprising a polynucleotide of the present invention, wherein the polynucleotide encodes a polypeptide of the present invention. Every nucleic acid sequence herein that encodes a polypeptide also, by reference to the genetic code, describes every possible silent variation of the nucleic acid. One of ordinary skill will recognize that each codon in a nucleic acid
20 (except AUG, which is ordinarily the only codon for methionine; and UGG, which is ordinarily the only codon for tryptophan) can be modified to yield a functionally identical molecule. Thus, each silent variation of a nucleic acid which encodes a polypeptide of the present invention is implicit in each described polypeptide sequence and is within the scope of the present invention. Accordingly, the present invention includes
25 polynucleotides of the present invention and polynucleotides encoding a polypeptide of the present invention.

B. Polynucleotides Amplified from a Plant Nucleic Acid Library

30 As indicated in (b), above, the present invention provides an isolated nucleic acid comprising a polynucleotide of the present invention, wherein the polynucleotides are amplified, under nucleic acid amplification conditions, from a plant nucleic acid library. Nucleic acid amplification conditions for each of the variety of amplification methods are well known to those of ordinary skill in the art. The plant nucleic acid library can be constructed from a monocot such as a cereal crop. Exemplary cereals include maize,

sorghum, alfalfa, canola, wheat, or rice. The plant nucleic acid library can also be constructed from a dicot such as soybean. *Zea mays* lines B73, PHRE1, A632, BMS-P2#10, W23, and Mo17 are known and publicly available. Other publicly known and available maize lines can be obtained from the Maize Genetics Cooperation (Urbana, IL).

5 Wheat lines are available from the Wheat Genetics Resource Center (Manhattan, KS).

The nucleic acid library may be a cDNA library, a genomic library, or a library generally constructed from nuclear transcripts at any stage of intron processing. cDNA libraries can be normalized to increase the representation of relatively rare cDNAs. In optional embodiments, the cDNA library is constructed using an enriched full-length
10 cDNA synthesis method. Examples of such methods include Oligo-Capping (Maruyama, K. and Sugano, S. *Gene* 138: 171-174, 1994), Biotinylated CAP Trapper (Carninci, *et al.* *Genomics* 37: 327-336, 1996), and CAP Retention Procedure (Edery, E., Chu, L.L., *et al.* *Molecular and Cellular Biology* 15: 3363-3371, 1995). Rapidly growing tissues or rapidly dividing cells are preferred for use as an mRNA source for construction of a cDNA library.
15 Growth stages of maize are described in "How a Corn Plant Develops," Special Report No. 48, Iowa State University of Science and Technology Cooperative Extension Service, Ames, Iowa, Reprinted February 1993.

A polynucleotide of this embodiment (or subsequences thereof) can be obtained, for example, by using amplification primers which are selectively hybridized and primer
20 extended, under nucleic acid amplification conditions, to at least two sites within a polynucleotide of the present invention, or to two sites within the nucleic acid which flank and comprise a polynucleotide of the present invention, or to a site within a polynucleotide of the present invention and a site within the nucleic acid which comprises it. Methods for obtaining 5' and/or 3' ends of a vector insert are well known in the art. See, e.g., RACE
25 (Rapid Amplification of Complementary Ends) as described in Frohman, M. A., in PCR Protocols: A Guide to Methods and Applications, M. A. Innis, D. H. Gelfand, J. J. Sninsky, T. J. White, Eds. (Academic Press, Inc., San Diego), pp. 28-38 (1990)); see also, U.S. Pat. No. 5,470,722, and *Current Protocols in Molecular Biology*, Unit 15.6, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995); Frohman and
30 Martin, *Techniques* 1:165 (1989).

Optionally, the primers are complementary to a subsequence of the target nucleic acid which they amplify but may have a sequence identity ranging from about 85% to 99% relative to the polynucleotide sequence which they are designed to anneal to. As those skilled in the art will appreciate, the sites to which the primer pairs will selectively

hybridize are chosen such that a single contiguous nucleic acid can be formed under the desired nucleic acid amplification conditions. The primer length in nucleotides is selected from the group of integers consisting of from at least 15 to 50. Thus, the primers can be at least 15, 18, 20, 25, 30, 40, or 50 nucleotides in length. Those of skill will recognize that a
5 lengthened primer sequence can be employed to increase specificity of binding (i.e., annealing) to a target sequence. A non-annealing sequence at the 5' end of a primer (a "tail") can be added, for example, to introduce a cloning site at the terminal ends of the amplicon.

The amplification products can be translated using expression systems well known
10 to those of skill in the art. The resulting translation products can be confirmed as polypeptides of the present invention by, for example, assaying for the appropriate catalytic activity (e.g., specific activity and/or substrate specificity), or verifying the presence of one or more epitopes which are specific to a polypeptide of the present invention. Methods for protein synthesis from PCR derived templates are known in the art
15 and available commercially. See, e.g., Amersham Life Sciences, Inc, Catalog '97, p.354.

C. Polynucleotides Which Selectively Hybridize to a Polynucleotide of (A) or (B)

As indicated in (c), above, the present invention provides isolated nucleic acids comprising polynucleotides of the present invention, wherein the polynucleotides
20 selectively hybridize, under selective hybridization conditions, to a polynucleotide of sections (A) or (B) as discussed above. Thus, the polynucleotides of this embodiment can be used for isolating, detecting, and/or quantifying nucleic acids comprising the polynucleotides of (A) or (B). For example, polynucleotides of the present invention can be used to identify, isolate, or amplify partial or full-length clones in a deposited library.
25 In some embodiments, the polynucleotides are genomic or cDNA sequences isolated or otherwise complementary to a cDNA from a dicot or monocot nucleic acid library. Exemplary species of monocots and dicots include, but are not limited to: maize, canola, soybean, cotton, wheat, sorghum, sunflower, alfalfa, oats, sugar cane, millet, barley, and rice. The cDNA library comprises at least 50% to 95% full-length sequences (for
30 example, at least 50%, 60%, 70%, 80%, 90%, or 95% full-length sequences). The cDNA libraries can be normalized to increase the representation of rare sequences. See, e.g., U.S. Patent No. 5,482,845. Low stringency hybridization conditions are typically, but not exclusively, employed with sequences having a reduced sequence identity relative to complementary sequences. Moderate and high stringency conditions can optionally be

employed for sequences of greater identity. Low stringency conditions allow selective hybridization of sequences having about 70% to 80% sequence identity and can be employed to identify orthologous or paralogous sequences.

5 *D. Polynucleotides Having a Specific Sequence Identity with the Polynucleotides of (A), (B) or (C)*

As indicated in (d), above, the present invention provides isolated nucleic acids comprising polynucleotides of the present invention, wherein the polynucleotides have a specified identity at the nucleotide level to a polynucleotide as disclosed above in sections
10 (A), (B), or (C), above. Identity can be calculated using, for example, the BLAST, CLUSTALW, or GAP algorithms under default conditions. The percentage of identity to a reference sequence is at least 50% and, rounded upwards to the nearest integer, can be expressed as an integer selected from the group of integers consisting of from 50 to 99. Thus, for example, the percentage of identity to a reference sequence can be at least 60%,
15 70%, 75%, 80%, 85%, 90%, or 95%.

Optionally, the polynucleotides of this embodiment will encode a polypeptide that will share an epitope with a polypeptide encoded by the polynucleotides of sections (A), (B), or (C). Thus, these polynucleotides encode a first polypeptide which elicits production of antisera comprising antibodies which are specifically reactive to a second
20 polypeptide encoded by a polynucleotide of (A), (B), or (C). However, the first polypeptide does not bind to antisera raised against itself when the antisera has been fully immunosorbed with the first polypeptide. Hence, the polynucleotides of this embodiment can be used to generate antibodies for use in, for example, the screening of expression libraries for nucleic acids comprising polynucleotides of (A), (B), or (C), or for
25 purification of, or in immunoassays for, polypeptides encoded by the polynucleotides of (A), (B), or (C). The polynucleotides of this embodiment comprise nucleic acid sequences which can be employed for selective hybridization to a polynucleotide encoding a polypeptide of the present invention.

Screening polypeptides for specific binding to antisera can be conveniently
30 achieved using peptide display libraries. This method involves the screening of large collections of peptides for individual members having the desired function or structure. Antibody screening of peptide display libraries is well known in the art. The displayed peptide sequences can be from 3 to 5000 or more amino acids in length, frequently from 5-100 amino acids long, and often from about 8 to 15 amino acids long. In addition to direct

chemical synthetic methods for generating peptide libraries, several recombinant DNA methods have been described. One type involves the display of a peptide sequence on the surface of a bacteriophage or cell. Each bacteriophage or cell contains the nucleotide sequence encoding the particular displayed peptide sequence. Such methods are described
5 in PCT patent publication Nos. 91/17271, 91/18980, 91/19818, and 93/08278. Other systems for generating libraries of peptides have aspects of both *in vitro* chemical synthesis and recombinant methods. See, PCT Patent publication Nos. 92/05258, 92/14843, and 97/20078. See also, U.S. Patent Nos. 5,658,754; and 5,643,768. Peptide display libraries, vectors, and screening kits are commercially available from such
10 suppliers as Invitrogen (Carlsbad, CA).

E. Polynucleotides Encoding a Protein Having a Subsequence from a Prototype Polypeptide and Cross-Reactive to the Prototype Polypeptide

As indicated in (e), above, the present invention provides isolated nucleic acids
15 comprising polynucleotides of the present invention, wherein the polynucleotides encode a protein having a subsequence of contiguous amino acids from a prototype polypeptide of the present invention such as are provided in (a), above. The length of contiguous amino acids from the prototype polypeptide is selected from the group of integers consisting of from at least 10 to the number of amino acids within the prototype sequence. Thus, for
20 example, the polynucleotide can encode a polypeptide having a subsequence having at least 10, 15, 20, 25, 30, 35, 40, 45, or 50, contiguous amino acids from the prototype polypeptide. Further, the number of such subsequences encoded by a polynucleotide of the instant embodiment can be any integer selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5. The subsequences can be separated by any integer of nucleotides
25 from 1 to the number of nucleotides in the sequence such as at least 5, 10, 15, 25, 50, 100, or 200 nucleotides.

The proteins encoded by polynucleotides of this embodiment, when presented as an immunogen, elicit the production of polyclonal antibodies which specifically bind to a
30 prototype polypeptide such as but not limited to, a polypeptide encoded by the polynucleotide of (a) or (b), above. Generally, however, a protein encoded by a polynucleotide of this embodiment does not bind to antisera raised against the prototype polypeptide when the antisera has been fully immunosorbed with the prototype polypeptide. Methods of making and assaying for antibody binding specificity/affinity are well known in the art. Exemplary immunoassay formats include ELISA, competitive

immunoassays, radioimmunoassays, Western blots, indirect immunofluorescent assays and the like.

In a preferred assay method, fully immunosorbed and pooled antisera which is elicited to the prototype polypeptide can be used in a competitive binding assay to test the protein. The concentration of the prototype polypeptide required to inhibit 50% of the binding of the antisera to the prototype polypeptide is determined. If the amount of the protein required to inhibit binding is less than twice the amount of the prototype protein, then the protein is said to specifically bind to the antisera elicited to the immunogen.

Accordingly, the proteins of the present invention embrace allelic variants, conservatively modified variants, and minor recombinant modifications to a prototype polypeptide.

A polynucleotide of the present invention optionally encodes a protein having a molecular weight as the non-glycosylated protein within 20% of the molecular weight of the full-length non-glycosylated polypeptides of the present invention. Molecular weight can be readily determined by SDS-PAGE under reducing conditions. Optionally, the molecular weight is within 15% of a full length polypeptide of the present invention, more preferably within 10% or 5%, and most preferably within 3%, 2%, or 1% of a full length polypeptide of the present invention.

Optionally, the polynucleotides of this embodiment will encode a protein having a specific enzymatic activity at least 50%, 60%, 80%, or 90% of a cellular extract

comprising the native, endogenous full-length polypeptide of the present invention.

Further, the proteins encoded by polynucleotides of this embodiment will optionally have a substantially similar affinity constant (K_m) and/or catalytic activity (i.e., the microscopic rate constant, k_{cat}) as the native endogenous, full-length protein. Those of skill in the art will recognize that k_{cat}/K_m value determines the specificity for competing substrates and is

often referred to as the specificity constant. Proteins of this embodiment can have a k_{cat}/K_m value at least 10% of a full-length polypeptide of the present invention as determined using the endogenous substrate of that polypeptide. Optionally, the k_{cat}/K_m value will be at least 20%, 30%, 40%, 50%, and most preferably at least 60%, 70%, 80%, 90%, or 95% the k_{cat}/K_m value of the full-length polypeptide of the present invention.

Determination of k_{cat} , K_m , and k_{cat}/K_m can be determined by any number of means well known to those of skill in the art. For example, the initial rates (i.e., the first 5% or less of the reaction) can be determined using rapid mixing and sampling techniques (e.g., continuous-flow, stopped-flow, or rapid quenching techniques), flash photolysis, or relaxation methods (e.g., temperature jumps) in conjunction with such exemplary methods

of measuring as spectrophotometry, spectrofluorimetry, nuclear magnetic resonance, or radioactive procedures. Kinetic values are conveniently obtained using a Lineweaver-Burk or Eadie-Hofstee plot.

5 *F. Polynucleotides Complementary to the Polynucleotides of (A)-(E)*

As indicated in (f), above, the present invention provides isolated nucleic acids comprising polynucleotides complementary to the polynucleotides of paragraphs A-E, above. As those of skill in the art will recognize, complementary sequences base-pair throughout the entirety of their length with the polynucleotides of sections (A)-(E) (i.e.,
10 have 100% sequence identity over their entire length). Complementary bases associate through hydrogen bonding in double stranded nucleic acids. For example, the following base pairs are complementary: guanine and cytosine; adenine and thymine; and adenine and uracil.

15 *G. Polynucleotides Which are Subsequences of the Polynucleotides of (A)-(F)*

As indicated in (g), above, the present invention provides isolated nucleic acids comprising polynucleotides which comprise at least 15 contiguous bases from the polynucleotides of sections (A) through (F) as discussed above. The length of the polynucleotide is given as an integer selected from the group consisting of from at least 15
20 to the length of the nucleic acid sequence from which the polynucleotide is a subsequence of. Thus, for example, polynucleotides of the present invention are inclusive of polynucleotides comprising at least 15, 20, 25, 30, 40, 50, 60, 75, or 100 contiguous nucleotides in length from the polynucleotides of (A)-(F). Optionally, the number of such subsequences encoded by a polynucleotide of the instant embodiment can be any integer
25 selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5. The subsequences can be separated by any integer of nucleotides from 1 to the number of nucleotides in the sequence such as at least 5, 10, 15, 25, 50, 100, or 200 nucleotides.

Subsequences can be made by *in vitro* synthetic, *in vitro* biosynthetic, or *in vivo* recombinant methods. In optional embodiments, subsequences can be made by nucleic
30 acid amplification. For example, nucleic acid primers will be constructed to selectively hybridize to a sequence (or its complement) within, or co-extensive with, the coding region.

The subsequences of the present invention can comprise structural characteristics of the sequence from which it is derived. Alternatively, the subsequences can lack certain

structural characteristics of the larger sequence from which it is derived such as a poly (A) tail. Optionally, a subsequence from a polynucleotide encoding a polypeptide having at least one epitope in common with a prototype polypeptide sequence as provided in (a), above, may encode an epitope in common with the prototype sequence. Alternatively, the subsequence may not encode an epitope in common with the prototype sequence but can be used to isolate the larger sequence by, for example, nucleic acid hybridization with the sequence from which it's derived. Subsequences can be used to modulate or detect gene expression by introducing into the subsequences compounds which bind, intercalate, cleave and/or crosslink to nucleic acids. Exemplary compounds include acridine, psoralen, phenanthroline, naphthoquinone, daunomycin or chloroethylaminoaryl conjugates.

H. Polynucleotides From a Full-length Enriched cDNA Library Having the Physico-Chemical Property of Selectively Hybridizing to a Polynucleotide of (A)-(G)

As indicated in (h), above, the present invention provides an isolated polynucleotide from a full-length enriched cDNA library having the physico-chemical property of selectively hybridizing to a polynucleotide of paragraphs (A), (B), (C), (D), (E), (F), or (G) as discussed above. Methods of constructing full-length enriched cDNA libraries are known in the art and discussed briefly below. The cDNA library comprises at least 50% to 95% full-length sequences (for example, at least 50%, 60%, 70%, 80%, 90%, or 95% full-length sequences). The cDNA library can be constructed from a variety of tissues from a monocot or dicot at a variety of developmental stages. Exemplary species include maize, wheat, rice, canola, soybean, cotton, sorghum, sunflower, alfalfa, oats, sugar cane, millet, barley, and rice. Methods of selectively hybridizing, under selective hybridization conditions, a polynucleotide from a full-length enriched library to a polynucleotide of the present invention are known to those of ordinary skill in the art. Any number of stringency conditions can be employed to allow for selective hybridization. In optional embodiments, the stringency allows for selective hybridization of sequences having at least 70%, 75%, 80%, 85%, 90%, 95%, or 98% sequence identity over the length of the hybridized region. Full-length enriched cDNA libraries can be normalized to increase the representation of rare sequences.

I. Polynucleotide Products Made by a cDNA Isolation Process

As indicated in (I), above, the present invention provides an isolated polynucleotide made by the process of: 1) providing a full-length enriched nucleic acid library, 2)

selectively hybridizing the polynucleotide to a polynucleotide of paragraphs (A), (B), (C), (D), (E), (F), (G, or (H) as discussed above, and thereby isolating the polynucleotide from the nucleic acid library. Full-length enriched nucleic acid libraries are constructed as discussed in paragraph (G) and below. Selective hybridization conditions are as discussed in paragraph (G). Nucleic acid purification procedures are well known in the art. Purification can be conveniently accomplished using solid-phase methods; such methods are well known to those of skill in the art and kits are available from commercial suppliers such as Advanced Biotechnologies (Surrey, UK). For example, a polynucleotide of paragraphs (A)-(H) can be immobilized to a solid support such as a membrane, bead, or particle. See, e.g., U.S. Patent No. 5,667,976. The polynucleotide product of the present process is selectively hybridized to an immobilized polynucleotide and the solid support is subsequently isolated from non-hybridized polynucleotides by methods including, but not limited to, centrifugation, magnetic separation, filtration, electrophoresis, and the like.

Construction of Nucleic Acids

The isolated nucleic acids of the present invention can be made using (a) standard recombinant methods, (b) synthetic techniques, or combinations thereof. In some embodiments, the polynucleotides of the present invention will be cloned, amplified, or otherwise constructed from a monocot such as maize, rice, or wheat, or a dicot such as soybean.

The nucleic acids may conveniently comprise sequences in addition to a polynucleotide of the present invention. For example, a multi-cloning site comprising one or more endonuclease restriction sites may be inserted into the nucleic acid to aid in isolation of the polynucleotide. Also, translatable sequences may be inserted to aid in the isolation of the translated polynucleotide of the present invention. For example, a hexahistidine marker sequence provides a convenient means to purify the proteins of the present invention. A polynucleotide of the present invention can be attached to a vector, adapter, or linker for cloning and/or expression of a polynucleotide of the present invention. Additional sequences may be added to such cloning and/or expression sequences to optimize their function in cloning and/or expression, to aid in isolation of the polynucleotide, or to improve the introduction of the polynucleotide into a cell. Typically, the length of a nucleic acid of the present invention less the length of its polynucleotide of the present invention is less than 20 kilobase pairs, often less than 15 kb, and frequently less than 10 kb. Use of cloning vectors, expression vectors, adapters, and linkers is well

known and extensively described in the art. For a description of various nucleic acids see, for example, Stratagene Cloning Systems, Catalogs 1999 (La Jolla, CA); and, Amersham Life Sciences, Inc, Catalog '99 (Arlington Heights, IL).

5 *A. Recombinant Methods for Constructing Nucleic Acids*

 The isolated nucleic acid compositions of this invention, such as RNA, cDNA, genomic DNA, or a hybrid thereof, can be obtained from plant biological sources using any number of cloning methodologies known to those of skill in the art. In some
10 embodiments, oligonucleotide probes which selectively hybridize, under stringent conditions, to the polynucleotides of the present invention are used to identify the desired sequence in a cDNA or genomic DNA library. Isolation of RNA, and construction of cDNA and genomic libraries is well known to those of ordinary skill in the art. See, e.g.,
15 *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997); and, *Current Protocols in Molecular Biology*, Ausubel, et al., Eds., Greene Publishing and Wiley-Interscience, New York (1995).

A1. Full-length Enriched cDNA Libraries

 A number of cDNA synthesis protocols have been described which provide enriched full-length cDNA libraries. Enriched full-length cDNA libraries are constructed
20 to comprise at least 600%, and more preferably at least 70%, 80%, 90% or 95% full-length inserts amongst clones containing inserts. The length of insert in such libraries can be at least 2,3, 4, 5, 6, 7, 8, 9, 10 or more kilobase pairs. Vectors to accommodate inserts of these sizes are known in the art and available commercially. See, e.g., Stratagene's lambda ZAP Express (cDNA cloning vector with 0 to 12 kb cloning capacity). An exemplary
25 method of constructing a greater than 95% pure full-length cDNA library is described by Carninci et al., *Genomics*, 37:327-336 (1996). Other methods for producing full-length libraries are known in the art. See, e.g., Edery et al., *Mol. Cell Biol.*, 15(6):3363-3371 (1995); and, PCT Application WO 96/34981.

30 *A2 Normalized or Subtracted cDNA Libraries*

 A non-normalized cDNA library represents the mRNA population of the tissue it was made from. Since unique clones are out-numbered by clones derived from highly expressed genes their isolation can be laborious. Normalization of a cDNA library is the process of creating a library in which each clone is more equally represented.

Construction of normalized libraries is described in Ko, *Nucl. Acids. Res.*, 18(19):5705-5711 (1990); Patanjali *et al.*, *Proc. Natl. Acad. U.S.A.*, 88:1943-1947 (1991); U.S. Patents 5,482,685, 5,482,845, and 5,637,685. In an exemplary method described by Soares *et al.*, normalization resulted in reduction of the abundance of clones from a range of four orders of magnitude to a narrow range of only 1 order of magnitude. *Proc. Natl. Acad. Sci. USA*, 91:9228-9232 (1994).

Subtracted cDNA libraries are another means to increase the proportion of less abundant cDNA species. In this procedure, cDNA prepared from one pool of mRNA is depleted of sequences present in a second pool of mRNA by hybridization. The cDNA:mRNA hybrids are removed and the remaining un-hybridized cDNA pool is enriched for sequences unique to that pool. See, Foote *et al.* in, *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997); Kho and Zarbl, *Technique*, 3(2):58-63 (1991); Sive and St. John, *Nucl. Acids Res.*, 16(22):10937 (1988); *Current Protocols in Molecular Biology*, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995); and, Swaroop *et al.*, *Nucl. Acids Res.*, 19(8):1954 (1991). cDNA subtraction kits are commercially available. See, e.g., PCR-Select (Clontech, Palo Alto, CA).

To construct genomic libraries, large segments of genomic DNA are generated by fragmentation, e.g. using restriction endonucleases, and are ligated with vector DNA to form concatemers that can be packaged into the appropriate vector. Methodologies to accomplish these ends, and sequencing methods to verify the sequence of nucleic acids are well known in the art. Examples of appropriate molecular biological techniques and instructions sufficient to direct persons of skill through many construction, cloning, and screening methodologies are found in Sambrook, *et al.*, *Molecular Cloning: A Laboratory Manual*, 2nd Ed., Cold Spring Harbor Laboratory Vols. 1-3 (1989), Methods in Enzymology, Vol. 152: *Guide to Molecular Cloning Techniques*, Berger and Kimmel, Eds., San Diego: Academic Press, Inc. (1987), *Current Protocols in Molecular Biology*, Ausubel, *et al.*, Eds., Greene Publishing and Wiley-Interscience, New York (1995); *Plant Molecular Biology: A Laboratory Manual*, Clark, Ed., Springer-Verlag, Berlin (1997). Kits for construction of genomic libraries are also commercially available.

The cDNA or genomic library can be screened using a probe based upon the sequence of a polynucleotide of the present invention such as those disclosed herein. Probes may be used to hybridize with genomic DNA or cDNA sequences to isolate homologous genes in the same or different plant species. Those of skill in the art will

appreciate that various degrees of stringency of hybridization can be employed in the assay; and either the hybridization or the wash medium can be stringent.

The nucleic acids of interest can also be amplified from nucleic acid samples using amplification techniques. For instance, polymerase chain reaction (PCR) technology can be used to amplify the sequences of polynucleotides of the present invention and related genes directly from genomic DNA or cDNA libraries. PCR and other *in vitro* amplification methods may also be useful, for example, to clone nucleic acid sequences that code for proteins to be expressed, to make nucleic acids to use as probes for detecting the presence of the desired mRNA in samples, for nucleic acid sequencing, or for other purposes. The T4 gene 32 protein (Boehringer Mannheim) can be used to improve yield of long PCR products.

PCR-based screening methods have been described. Wilfinger *et al.* describe a PCR-based method in which the longest cDNA is identified in the first step so that incomplete clones can be eliminated from study. *BioTechniques*, 22(3): 481-486 (1997). Such methods are particularly effective in combination with a full-length cDNA construction methodology, above.

B. Synthetic Methods for Constructing Nucleic Acids

The isolated nucleic acids of the present invention can also be prepared by direct chemical synthesis by methods such as the phosphotriester method of Narang *et al.*, *Meth. Enzymol.* 68: 90-99 (1979); the phosphodiester method of Brown *et al.*, *Meth. Enzymol.* 68: 109-151 (1979); the diethylphosphoramidite method of Beaucage *et al.*, *Tetra. Lett.* 22: 1859-1862 (1981); the solid phase phosphoramidite triester method described by Beaucage and Caruthers, *Tetra. Letts.* 22(20): 1859-1862 (1981), *e.g.*, using an automated synthesizer, *e.g.*, as described in Needham-VanDevanter *et al.*, *Nucleic Acids Res.*, 12: 6159-6168 (1984); and, the solid support method of U.S. Patent No. 4,458,066. Chemical synthesis generally produces a single stranded oligonucleotide. This may be converted into double stranded DNA by hybridization with a complementary sequence, or by polymerization with a DNA polymerase using the single strand as a template. One of skill will recognize that while chemical synthesis of DNA is best employed for sequences of about 100 bases or less, longer sequences may be obtained by the ligation of shorter sequences.

Recombinant Expression Cassettes

The present invention further provides recombinant expression cassettes comprising a nucleic acid of the present invention. A nucleic acid sequence coding for the desired polypeptide of the present invention, for example a cDNA or a genomic sequence encoding a full length polypeptide of the present invention, can be used to construct a recombinant expression cassette which can be introduced into the desired host cell. A recombinant expression cassette will typically comprise a polynucleotide of the present invention operably linked to transcriptional initiation regulatory sequences which will direct the transcription of the polynucleotide in the intended host cell, such as tissues of a transformed plant.

For example, plant expression vectors may include (1) a cloned plant gene under the transcriptional control of 5' and 3' regulatory sequences and (2) a dominant selectable marker. Such plant expression vectors may also contain, if desired, a promoter regulatory region (e.g., one conferring inducible or constitutive, environmentally- or developmentally-regulated, or cell- or tissue-specific/selective expression), a transcription initiation start site, a ribosome binding site, an RNA processing signal, a transcription termination site, and/or a polyadenylation signal.

A plant promoter fragment can be employed which will direct expression of a polynucleotide of the present invention in all tissues of a regenerated plant. Such promoters are referred to herein as "constitutive" promoters and are active under most environmental conditions and states of development or cell differentiation. Examples of constitutive promoters include the cauliflower mosaic virus (CaMV) 35S transcription initiation region, the 1'- or 2'- promoter derived from T-DNA of *Agrobacterium tumefaciens*, the ubiquitin 1 promoter, the Smas promoter, the cinnamyl alcohol dehydrogenase promoter (U.S. Patent No. 5,683,439), the *Nos* promoter, the pEmu promoter, the rubisco promoter, and the GRP1-8 promoter.

Alternatively, the plant promoter can direct expression of a polynucleotide of the present invention in a specific tissue or may be otherwise under more precise environmental or developmental control. Such promoters are referred to here as "inducible" promoters. Environmental conditions that may effect transcription by inducible promoters include pathogen attack, anaerobic conditions, or the presence of light. Examples of inducible promoters are the *Adh1* promoter which is inducible by hypoxia or cold stress, the *Hsp70* promoter which is inducible by heat stress, and the *PPDK* promoter which is inducible by light.

Examples of promoters under developmental control include promoters that initiate transcription only, or preferentially, in certain tissues, such as leaves, roots, fruit, seeds, or flowers. Exemplary promoters include the anther specific promoter 5126 (U.S. Patent Nos. 5,689,049 and 5,689,051), glob-1 promoter, and gamma-zein promoter. See also US
5 patent applications 60/163,114, filed November 2, 1999, and 60/155,859, filed September 24, 1999. The operation of a promoter may also vary depending on its location in the genome. Thus, an inducible promoter may become fully or partially constitutive in certain locations.

Both heterologous and non-heterologous (i.e., endogenous) promoters can be
10 employed to direct expression of the nucleic acids of the present invention. These promoters can also be used, for example, in recombinant expression cassettes to drive expression of antisense nucleic acids to reduce, increase, or alter concentration and/or composition of the proteins of the present invention in a desired tissue. Thus, in some
15 embodiments, the nucleic acid construct will comprise a promoter, functional in a plant cell, operably linked to a polynucleotide of the present invention. Promoters useful in these embodiments include the endogenous promoters driving expression of a polypeptide of the present invention.

In some embodiments, isolated nucleic acids which serve as promoter or enhancer elements can be introduced in the appropriate position (generally upstream) of a non-
20 heterologous form of a polynucleotide of the present invention so as to up or down regulate expression of a polynucleotide of the present invention. For example, endogenous promoters can be altered *in vivo* by mutation, deletion, and/or substitution (see, Kmiec, U.S. Patent 5,565,350; Zarling *et al.*, WO 93/22443), or isolated promoters can be introduced into a plant cell in the proper orientation and distance from a cognate gene of a
25 polynucleotide of the present invention so as to control the expression of the gene. Gene expression can be modulated under conditions suitable for plant growth so as to alter the total concentration and/or alter the composition of the polypeptides of the present invention in plant cell. Thus, the present invention provides compositions, and methods for making, heterologous promoters and/or enhancers operably linked to a native, endogenous
30 (i.e., non-heterologous) form of a polynucleotide of the present invention.

If polypeptide expression is desired, it is generally desirable to include a polyadenylation region at the 3'-end of a polynucleotide coding region. The polyadenylation region can be derived from the natural gene, from a variety of other plant genes, or from T-DNA. The 3' end sequence to be added can be derived from, for

example, the nopaline synthase or octopine synthase genes, or alternatively from another plant gene, or less preferably from any other eukaryotic gene.

An intron sequence can be added to the 5' untranslated region or the coding sequence of the partial coding sequence to increase the amount of the mature message that accumulates in the cytosol. Inclusion of a spliceable intron in the transcription unit in both plant and animal expression constructs has been shown to increase gene expression at both the mRNA and protein levels up to 1000-fold. Buchman and Berg, *Mol. Cell Biol.* 8: 4395-4405 (1988); Callis *et al.*, *Genes Dev.* 1: 1183-1200 (1987). Such intron enhancement of gene expression is typically greatest when placed near the 5' end of the transcription unit.

Use of maize introns Adh1-S intron 1, 2, and 6, the Bronze-1 intron are known in the art. See generally, *The Maize Handbook*, Chapter 116, Freeling and Walbot, Eds., Springer, New York (1994). The vector comprising the sequences from a polynucleotide of the present invention will typically comprise a marker gene which confers a selectable phenotype on plant cells. Typical vectors useful for expression of genes in higher plants are well known in the art and include vectors derived from the tumor-inducing (Ti) plasmid of *Agrobacterium tumefaciens* described by Rogers *et al.*, *Meth. in Enzymol.*, 153:253-277 (1987).

A polynucleotide of the present invention can be expressed in either sense or anti-sense orientation as desired. It will be appreciated that control of gene expression in either sense or anti-sense orientation can have a direct impact on the observable plant characteristics. Antisense technology can be conveniently used to inhibit gene expression in plants. To accomplish this, a nucleic acid segment from the desired gene is cloned and operably linked to a promoter such that the anti-sense strand of RNA will be transcribed. The construct is then transformed into plants and the antisense strand of RNA is produced.

In plant cells, it has been shown that antisense RNA inhibits gene expression by preventing the accumulation of mRNA which encodes the enzyme of interest, see, e.g., Sheehy *et al.*, *Proc. Nat'l. Acad. Sci. (USA)* 85: 8805-8809 (1988); and Hiatt *et al.*, U.S. Patent No. 4,801,340.

Another method of suppression is sense suppression (i.e., co-suppression).

Introduction of nucleic acid configured in the sense orientation has been shown to be an effective means by which to block the transcription of target genes. For an example of the use of this method to modulate expression of endogenous genes see, Napoli *et al.*, *The Plant Cell* 2: 279-289 (1990) and U.S. Patent No. 5,034,323.

Catalytic RNA molecules or ribozymes can also be used to inhibit expression of plant genes. It is possible to design ribozymes that specifically pair with virtually any target RNA and cleave the phosphodiester backbone at a specific location, thereby functionally inactivating the target RNA. In carrying out this cleavage, the ribozyme is not itself altered, and is thus capable of recycling and cleaving other molecules, making it a true enzyme. The inclusion of ribozyme sequences within antisense RNAs confers RNA-cleaving activity upon them, thereby increasing the activity of the constructs. The design and use of target RNA-specific ribozymes is described in Haseloff *et al.*, *Nature* 334: 585-591 (1988).

A variety of cross-linking agents, alkylating agents and radical generating species as pendant groups on polynucleotides of the present invention can be used to bind, label, detect, and/or cleave nucleic acids. For example, Vlassov, V. V., *et al.*, *Nucleic Acids Res* (1986) 14:4065-4076, describe covalent bonding of a single-stranded DNA fragment with alkylating derivatives of nucleotides complementary to target sequences. A report of similar work by the same group is that by Knorre, D. G., *et al.*, *Biochimie* (1985) 67:785-789. Iverson and Dervan also showed sequence-specific cleavage of single-stranded DNA mediated by incorporation of a modified nucleotide which was capable of activating cleavage (*J Am Chem Soc* (1987) 109:1241-1243). Meyer, R. B., *et al.*, *J Am Chem Soc* (1989) 111:8517-8519, effect covalent crosslinking to a target nucleotide using an alkylating agent complementary to the single-stranded target nucleotide sequence. A photoactivated crosslinking to single-stranded oligonucleotides mediated by psoralen was disclosed by Lee, B. L., *et al.*, *Biochemistry* (1988) 27:3197-3203. Use of crosslinking in triple-helix forming probes was also disclosed by Home, *et al.*, *J Am Chem Soc* (1990) 112:2435-2437. Use of N⁴, N⁴-ethanocytosine as an alkylating agent to crosslink to single-stranded oligonucleotides has also been described by Webb and Matteucci, *J Am Chem Soc* (1986) 108:2764-2765; *Nucleic Acids Res* (1986) 14:7661-7674; Feteritz *et al.*, *J. Am. Chem. Soc.* 113:4000 (1991). Various compounds to bind, detect, label, and/or cleave nucleic acids are known in the art. See, for example, U.S. Patent Nos. 5,543,507; 5,672,593; 5,484,908; 5,256,648; and, 5,681,941.

Proteins

The isolated proteins of the present invention comprise a polypeptide having at least 10 amino acids from a polypeptide of the present invention (or conservative variants thereof) such as those encoded by any one of the polynucleotides of the present invention

as discussed more fully above. The proteins of the present invention or variants thereof can comprise any number of contiguous amino acid residues from a polypeptide of the present invention, wherein that number is selected from the group of integers consisting of from 10 to the number of residues in a full-length polypeptide of the present invention.

- 5 Optionally, this subsequence of contiguous amino acids is at least 15, 20, 25, 30, 35, or 40 amino acids in length, often at least 50, 60, 70, 80, or 90 amino acids in length. Further, the number of such subsequences can be any integer selected from the group consisting of from 1 to 20, such as 2, 3, 4, or 5.

- 10 The present invention further provides a protein comprising a polypeptide having a specified sequence identity/similarity with a polypeptide of the present invention. The percentage of sequence identity/similarity is an integer selected from the group consisting of from 50 to 99. Exemplary sequence identity/similarity values include 55%, 60%, 65%, 70%, 75%, 80%, 85%, 90%, and 95%. Sequence identity can be determined using, for example, the GAP, CLUSTALW, or BLAST algorithms.

- 15 As those of skill will appreciate, the present invention includes, but is not limited to, catalytically active polypeptides of the present invention (i.e., enzymes). Catalytically active polypeptides have a specific activity of at least 20%, 30%, or 40%, and preferably at least 50%, 60%, or 70%, and most preferably at least 80%, 90%, or 95% that of the native (non-synthetic), endogenous polypeptide. Further, the substrate specificity (k_{cat}/K_m) is
20 optionally substantially similar to the native (non-synthetic), endogenous polypeptide. Typically, the K_m will be at least 30%, 40%, or 50%, that of the native (non-synthetic), endogenous polypeptide; and more preferably at least 60%, 70%, 80%, or 90%. Methods of assaying and quantifying measures of enzymatic activity and substrate specificity (k_{cat}/K_m), are well known to those of skill in the art.

- 25 Generally, the proteins of the present invention will, when presented as an immunogen, elicit production of an antibody specifically reactive to a polypeptide of the present invention. Further, the proteins of the present invention will not bind to antisera raised against a polypeptide of the present invention which has been fully immunosorbed with the same polypeptide. Immunoassays for determining binding are well known to
30 those of skill in the art. A preferred immunoassay is a competitive immunoassay. Thus, the proteins of the present invention can be employed as immunogens for constructing antibodies immunoreactive to a protein of the present invention for such exemplary utilities as immunoassays or protein purification techniques.

Expression of Proteins in Host Cells

Using the nucleic acids of the present invention, one may express a protein of the present invention in a recombinantly engineered cell such as bacteria, yeast, insect, mammalian, or preferably plant cells. The cells produce the protein in a non-natural
5 condition (e.g., in quantity, composition, location, and/or time), because they have been genetically altered through human intervention to do so.

It is expected that those of skill in the art are knowledgeable in the numerous expression systems available for expression of a nucleic acid encoding a protein of the present invention. No attempt to describe in detail the various methods known for the
10 expression of proteins in prokaryotes or eukaryotes will be made.

In brief summary, the expression of isolated nucleic acids encoding a protein of the present invention will typically be achieved by operably linking, for example, the DNA or cDNA to a promoter (which is either constitutive or regulatable), followed by
15 incorporation into an expression vector. The vectors can be suitable for replication and integration in either prokaryotes or eukaryotes. Typical expression vectors contain transcription and translation terminators, initiation sequences, and promoters useful for regulation of the expression of the DNA encoding a protein of the present invention. To obtain high level expression of a cloned gene, it is desirable to construct expression vectors which contain, at the minimum, a strong promoter to direct transcription, a ribosome
20 binding site for translational initiation, and a transcription/translation terminator. One of skill would recognize that modifications can be made to a protein of the present invention without diminishing its biological activity. Some modifications may be made to facilitate the cloning, expression, or incorporation of the targeting molecule into a fusion protein. Such modifications are well known to those of skill in the art and include, for example, a
25 methionine added at the amino terminus to provide an initiation site, or additional amino acids (e.g., poly His) placed on either terminus to create conveniently located purification sequences. Restriction sites or termination codons can also be introduced.

Synthesis of Proteins

30 The proteins of the present invention can be constructed using non-cellular synthetic methods. Solid phase synthesis of proteins of less than about 50 amino acids in length may be accomplished by attaching the C-terminal amino acid of the sequence to an insoluble support followed by sequential addition of the remaining amino acids in the sequence. Techniques for solid phase synthesis are described by Barany and Merrifield,

Solid-Phase Peptide Synthesis, pp. 3-284 in *The Peptides: Analysis, Synthesis, Biology*, Vol. 2: *Special Methods in Peptide Synthesis, Part A.*; Merrifield, et al., *J. Am. Chem. Soc.* 85: 2149-2156 (1963), and Stewart et al., *Solid Phase Peptide Synthesis*, 2nd ed., Pierce Chem. Co., Rockford, Ill. (1984). Proteins of greater length may be synthesized by
5 condensation of the amino and carboxy termini of shorter fragments. Methods of forming peptide bonds by activation of a carboxy terminal end (e.g., by the use of the coupling reagent N,N'-dicyclohexylcarbodiimide) are known to those of skill.

Purification of Proteins

10 The proteins of the present invention may be purified by standard techniques well known to those of skill in the art. Recombinantly produced proteins of the present invention can be directly expressed or expressed as a fusion protein. The recombinant protein is purified by a combination of cell lysis (e.g., sonication, French press) and affinity chromatography. For fusion products, subsequent digestion of the fusion protein
15 with an appropriate proteolytic enzyme releases the desired recombinant protein.

The proteins of this invention, recombinant or synthetic, may be purified to substantial purity by standard techniques well known in the art, including detergent solubilization, selective precipitation with such substances as ammonium sulfate, column chromatography, immunopurification methods, and others. See, for instance, R. Scopes,
20 *Protein Purification: Principles and Practice*, Springer-Verlag: New York (1982); Deutscher, *Guide to Protein Purification*, Academic Press (1990). For example, antibodies may be raised to the proteins as described herein. Purification from *E. coli* can be achieved following procedures described in U.S. Patent No. 4,511,503. The protein may then be isolated from cells expressing the protein and further purified by standard
25 protein chemistry techniques as described herein. Detection of the expressed protein is achieved by methods known in the art and include, for example, radioimmunoassays, Western blotting techniques or immunoprecipitation.

Introduction of Nucleic Acids Into Host Cells

30 The method of introducing a nucleic acid of the present invention into a host cell is not critical to the instant invention. Transformation or transfection methods are conveniently used. Accordingly, a wide variety of methods have been developed to insert a DNA sequence into the genome of a host cell to obtain the transcription and/or

translation of the sequence to effect phenotypic changes in the organism. Thus, any method which provides for effective introduction of a nucleic acid may be employed.

A. Plant Transformation

5 A nucleic acid comprising a polynucleotide of the present invention is optionally introduced into a plant. Generally, the polynucleotide will first be incorporated into a recombinant expression cassette or vector. Isolated nucleic acid acids of the present invention can be introduced into plants according to techniques known in the art. Techniques for transforming a wide variety of higher plant species are well known and described in the technical, scientific, and patent literature. See, for example, Weising *et al.*, *Ann. Rev. Genet.* 22: 421-477 (1988). For example, the DNA construct may be introduced directly into the genomic DNA of the plant cell using techniques such as electroporation, polyethylene glycol (PEG), poration, particle bombardment, silicon fiber delivery, or microinjection of plant cell protoplasts or embryogenic callus. See, e.g.,
10 Tomes, *et al.*, Direct DNA Transfer into Intact Plant Cells Via Microprojectile Bombardment. pp.197-213 in *Plant Cell, Tissue and Organ Culture, Fundamental Methods*. eds. O. L. Gamborg and G.C. Phillips. Springer-Verlag Berlin Heidelberg New York, 1995; see, U.S. Patent No. 5,990,387. The introduction of DNA constructs using PEG precipitation is described in Paszkowski *et al.*, *Embo J.* 3: 2717-2722 (1984).
15 Electroporation techniques are described in Fromm *et al.*, *Proc. Natl. Acad. Sci. (USA)* 82: 5824 (1985). Ballistic transformation techniques are described in Klein *et al.*, *Nature* 327: 70-73 (1987).

Agrobacterium tumefaciens-mediated transformation techniques are well described in the scientific literature. See, for example Horsch *et al.*, *Science* 233: 496-498 (1984);
25 Fraley *et al.*, *Proc. Natl. Acad. Sci. (USA)* 80: 4803 (1983); and, *Plant Molecular Biology: A Laboratory Manual*, Chapter 8, Clark, Ed., Springer-Verlag, Berlin (1997). The DNA constructs may be combined with suitable T-DNA flanking regions and introduced into a conventional *Agrobacterium tumefaciens* host vector. The virulence functions of the *Agrobacterium tumefaciens* host will direct the insertion of the construct and adjacent
30 marker into the plant cell DNA when the cell is infected by the bacteria. See, U.S. Patent No. 5,591,616. Although *Agrobacterium* is useful primarily in dicots, certain monocots can be transformed by *Agrobacterium*. For instance, *Agrobacterium* transformation of maize is described in U.S. Patent No. 5,550,318.

Other methods of transfection or transformation include (1) *Agrobacterium rhizogenes*-mediated transformation (see, e.g., Lichtenstein and Fuller In: Genetic Engineering, vol. 6, PWJ Rigby, Ed., London. Academic Press, 1987; and Lichtenstein, C. P., and Draper, J., In: DNA Cloning, Vol. II, D. M. Glover, Ed., Oxford, IRI Press, 1985),
5 Application PCT/US87/02512 (WO 88/02405 published Apr. 7, 1988) describes the use of *A. rhizogenes* strain A4 and its Ri plasmid along with *A. tumefaciens* vectors pARC8 or pARC16 (2) liposome-mediated DNA uptake (see, e.g., Freeman *et al.*, *Plant Cell Physiol.* 25: 1353 (1984)), (3) the vortexing method (see, e.g., Kindle, *Proc. Natl. Acad. Sci., (USA)* 87: 1228 (1990)).

10 DNA can also be introduced into plants by direct DNA transfer into pollen as described by Zhou *et al.*, *Methods in Enzymology*, 101:433 (1983); D. Hess, *Intern Rev. Cytol.*, 107:367 (1987); Luo *et al.*, *Plant Mol. Biol. Reporter*, 6:165 (1988). Expression of polypeptide coding genes can be obtained by injection of the DNA into reproductive organs of a plant as described by Pena *et al.*, *Nature*, 325:274 (1987).
15 DNA can also be injected directly into the cells of immature embryos and the rehydration of desiccated embryos as described by Neuhaus *et al.*, *Theor. Appl. Genet.*, 75:30 (1987); and Benbrook *et al.*, in *Proceedings Bio Expo 1986*, Butterworth, Stoneham, Mass., pp. 27-54 (1986). A variety of plant viruses that can be employed as vectors are known in the art and include cauliflower mosaic virus (CaMV), geminivirus, brome mosaic virus, and
20 tobacco mosaic virus.

B. Transfection of Prokaryotes, Lower Eukaryotes, and Animal Cells

Animal and lower eukaryotic (e.g., yeast) host cells are competent or rendered competent for transfection by various means. There are several well-known methods of
25 introducing DNA into animal cells. These include: calcium phosphate precipitation, fusion of the recipient cells with bacterial protoplasts containing the DNA, treatment of the recipient cells with liposomes containing the DNA, DEAE dextran, electroporation, biolistics, and micro-injection of the DNA directly into the cells. The transfected cells are cultured by means well known in the art. Kuchler, R.J., *Biochemical Methods in Cell*
30 *Culture and Virology*, Dowden, Hutchinson and Ross, Inc. (1977).

Transgenic Plant Regeneration

Plant cells which directly result or are derived from the nucleic acid introduction techniques can be cultured to regenerate a whole plant which possesses the introduced

genotype. Such regeneration techniques often rely on manipulation of certain phytohormones in a tissue culture growth medium. Plants cells can be regenerated, e.g., from single cells, callus tissue or leaf discs according to standard plant tissue culture techniques. It is well known in the art that various cells, tissues, and organs from almost any plant can be successfully cultured to regenerate an entire plant. Plant regeneration from cultured protoplasts is described in Evans *et al.*, *Protoplasts Isolation and Culture, Handbook of Plant Cell Culture*, Macmillan Publishing Company, New York, pp. 124-176 (1983); and Binding, *Regeneration of Plants. Plant Protoplasts*, CRC Press, Boca Raton, pp. 21-73 (1985).

The regeneration of plants from either single plant protoplasts or various explants is well known in the art. See, for example, *Methods for Plant Molecular Biology*, A. Weissbach and H. Weissbach, eds., Academic Press, Inc., San Diego, Calif. (1988). This regeneration and growth process includes the steps of selection of transformant cells and shoots, rooting the transformant shoots and growth of the plantlets in soil. For maize cell culture and regeneration see generally, *The Maize Handbook*, Freeling and Walbot, Eds., Springer, New York (1994); *Corn and Corn Improvement*, 3rd edition, Sprague and Dudley Eds., American Society of Agronomy, Madison, Wisconsin (1988). For transformation and regeneration of maize see, Gordon-Kamm *et al.*, *The Plant Cell*, 2:603-618 (1990).

The regeneration of plants containing the polynucleotide of the present invention and introduced by *Agrobacterium* from leaf explants can be achieved as described by Horsch *et al.*, *Science*, 227:1229-1231 (1985). In this procedure, transformants are grown in the presence of a selection agent and in a medium that induces the regeneration of shoots in the plant species being transformed as described by Fraley *et al.*, *Proc. Natl. Acad. Sci. (U.S.A.)*, 80:4803 (1983). This procedure typically produces shoots within two to four weeks and these transformant shoots are then transferred to an appropriate root-inducing medium containing the selective agent and an antibiotic to prevent bacterial growth. Transgenic plants of the present invention may be fertile or sterile.

One of skill will recognize that after the recombinant expression cassette is stably incorporated in transgenic plants and confirmed to be operable, it can be introduced into other plants by sexual crossing. Any of a number of standard breeding techniques can be used, depending upon the species to be crossed. In vegetatively propagated crops, mature transgenic plants can be propagated by the taking of cuttings or by tissue culture techniques to produce multiple identical plants. Selection of desirable transgenics is made and new varieties are obtained and propagated vegetatively for commercial use. In seed

propagated crops, mature transgenic plants can be self crossed to produce a homozygous inbred plant. The inbred plant produces seed containing the newly introduced heterologous nucleic acid. These seeds can be grown to produce plants that would produce the selected phenotype. Parts obtained from the regenerated plant, such as flowers, seeds, leaves, branches, fruit, and the like are included in the invention, provided that these parts comprise cells comprising the isolated nucleic acid of the present invention. Progeny and variants, and mutants of the regenerated plants are also included within the scope of the invention, provided that these parts comprise the introduced nucleic acid sequences.

Transgenic plants expressing a polynucleotide of the present invention can be screened for transmission of the nucleic acid of the present invention by, for example, standard immunoblot and DNA detection techniques. Expression at the RNA level can be determined initially to identify and quantitate expression-positive plants. Standard techniques for RNA analysis can be employed and include PCR amplification assays using oligonucleotide primers designed to amplify only the heterologous RNA templates and solution hybridization assays using heterologous nucleic acid-specific probes. The RNA-positive plants can then analyzed for protein expression by Western immunoblot analysis using the specifically reactive antibodies of the present invention. In addition, *in situ* hybridization and immunocytochemistry according to standard protocols can be done using heterologous nucleic acid specific polynucleotide probes and antibodies, respectively, to localize sites of expression within transgenic tissue. Generally, a number of transgenic lines are usually screened for the incorporated nucleic acid to identify and select plants with the most appropriate expression profiles.

A preferred embodiment is a transgenic plant that is homozygous for the added heterologous nucleic acid; i.e., a transgenic plant that contains two added nucleic acid sequences, one gene at the same locus on each chromosome of a chromosome pair. A homozygous transgenic plant can be obtained by sexually mating (selfing) a heterozygous transgenic plant that contains a single added heterologous nucleic acid, germinating some of the seed produced and analyzing the resulting plants produced for altered expression of a polynucleotide of the present invention relative to a control plant (i.e., native, non-transgenic). Back-crossing to a parental plant and out-crossing with a non-transgenic plant are also contemplated.

Modulating Polypeptide Levels and/or Composition

The present invention further provides a method for modulating (i.e., increasing or decreasing) the concentration or ratio of the polypeptides of the present invention in a plant or part thereof. Modulation can be effected by increasing or decreasing the concentration and/or the ratio of the polypeptides of the present invention in a plant. The method comprises introducing into a plant cell a recombinant expression cassette comprising a polynucleotide of the present invention as described above to obtain a transgenic plant cell, culturing the transgenic plant cell under transgenic plant cell growing conditions, and inducing or repressing expression of a polynucleotide of the present invention in the transgenic plant for a time sufficient to modulate concentration and/or the ratios of the polypeptides in the transgenic plant or plant part.

In some embodiments, the concentration and/or ratios of polypeptides of the present invention in a plant may be modulated by altering, *in vivo* or *in vitro*, the promoter of a gene to up- or down-regulate gene expression. In some embodiments, the coding regions of native genes of the present invention can be altered via substitution, addition, insertion, or deletion to decrease activity of the encoded enzyme. See, e.g., Kmiec, U.S. Patent 5,565,350; Zarling *et al.*, WO 93/22443; Briggs, *et al.*, U.S. Patent 5,962,764. And in some embodiments, an isolated nucleic acid (e.g., a vector) comprising a promoter sequence is transfected into a plant cell. Subsequently, a plant cell comprising the promoter operably linked to a polynucleotide of the present invention is selected for by means known to those of skill in the art such as, but not limited to, Southern blot, DNA sequencing, or PCR analysis using primers specific to the promoter and to the gene and detecting amplicons produced therefrom. A plant or plant part altered or modified by the foregoing embodiments is grown under plant forming conditions for a time sufficient to modulate the concentration and/or ratios of polypeptides of the present invention in the plant. Plant forming conditions are well known in the art and discussed briefly, *supra*.

In general, concentration or the ratios of the polypeptides is increased or decreased by at least 5%, 10%, 20%, 30%, 40%, 50%, 60%, 70%, 80%, or 90% relative to a native control plant, plant part, or cell lacking the aforementioned recombinant expression cassette. Modulation in the present invention may occur during and/or subsequent to growth of the plant to the desired stage of development. Modulating nucleic acid expression temporally and/or in particular tissues can be controlled by employing the appropriate promoter operably linked to a polynucleotide of the present invention in, for example, sense or antisense orientation as discussed in greater detail, *supra*. Induction of expression of a polynucleotide of the present invention can also be controlled by

exogenous administration of an effective amount of inducing compound. Inducible promoters and inducing compounds which activate expression from these promoters are well known in the art. In preferred embodiments, the polypeptides of the present invention are modulated in monocots, particularly maize.

5

UTRs and Codon Preference

In general, translational efficiency has been found to be regulated by specific sequence elements in the 5' non-coding or untranslated region (5' UTR) of the RNA. Positive sequence motifs include translational initiation consensus sequences (Kozak, *Nucleic Acids Res.* 15:8125 (1987)) and the 7-methylguanosine cap structure (Drummond *et al.*, *Nucleic Acids Res.* 13:7375 (1985)). Negative elements include stable intramolecular 5' UTR stem-loop structures (Muesing *et al.*, *Cell* 48:691 (1987)) and AUG sequences or short open reading frames preceded by an appropriate AUG in the 5' UTR (Kozak, *supra*, Rao *et al.*, *Mol. and Cell. Biol.* 8:284 (1988)). Accordingly, the present invention provides 5' and/or 3' untranslated regions for modulation of translation of heterologous coding sequences.

Further, the polypeptide-encoding segments of the polynucleotides of the present invention can be modified to alter codon usage. Altered codon usage can be employed to alter translational efficiency and/or to optimize the coding sequence for expression in a desired host such as to optimize the codon usage in a heterologous sequence for expression in maize. Codon usage in the coding regions of the polynucleotides of the present invention can be analyzed statistically using commercially available software packages such as "Codon Preference" available from the University of Wisconsin Genetics Computer Group (see Devereaux *et al.*, *Nucleic Acids Res.* 12: 387-395 (1984)) or MacVector 4.1 (Eastman Kodak Co., New Haven, Conn.). Thus, the present invention provides a codon usage frequency characteristic of the coding region of at least one of the polynucleotides of the present invention. The number of polynucleotides that can be used to determine a codon usage frequency can be any integer from 1 to the number of polynucleotides of the present invention as provided herein. Optionally, the polynucleotides will be full-length sequences. An exemplary number of sequences for statistical analysis can be at least 1, 5, 10, 20, 50, or 100.

Sequence Shuffling

The present invention provides methods for sequence shuffling using polynucleotides of the present invention, and compositions resulting therefrom. Sequence shuffling is described in PCT publication No. WO 97/20078. See also, Zhang, J.- H., *et al. Proc. Natl. Acad. Sci. USA* 94:4504-4509 (1997). Generally, sequence shuffling provides a means for generating libraries of polynucleotides having a desired characteristic which can be selected or screened for. Libraries of recombinant polynucleotides are generated from a population of related sequence polynucleotides which comprise sequence regions which have substantial sequence identity and can be homologously recombined *in vitro* or *in vivo*. The population of sequence-recombined polynucleotides comprises a subpopulation of polynucleotides which possess desired or advantageous characteristics and which can be selected by a suitable selection or screening method. The characteristics can be any property or attribute capable of being selected for or detected in a screening system, and may include properties of: an encoded protein, a transcriptional element, a sequence controlling transcription, RNA processing, RNA stability, chromatin conformation, translation, or other expression property of a gene or transgene, a replicative element, a protein-binding element, or the like, such as any feature which confers a selectable or detectable property. In some embodiments, the selected characteristic will be a decreased K_m and/or increased K_{cat} over the wild-type protein as provided herein. In other embodiments, a protein or polynucleotide generated from sequence shuffling will have a ligand binding affinity greater than the non-shuffled wild-type polynucleotide. The increase in such properties can be at least 110%, 120%, 130%, 140% or at least 150% of the wild-type value.

Generic and Consensus Sequences

Polynucleotides and polypeptides of the present invention further include those having: (a) a generic sequence of at least two homologous polynucleotides or polypeptides, respectively, of the present invention; and, (b) a consensus sequence of at least three homologous polynucleotides or polypeptides, respectively, of the present invention. The generic sequence of the present invention comprises each species of polypeptide or polynucleotide embraced by the generic polypeptide or polynucleotide sequence, respectively. The individual species encompassed by a polynucleotide having an amino acid or nucleic acid consensus sequence can be used to generate antibodies or produce nucleic acid probes or primers to screen for homologs in other species, genera, families, orders, classes, phyla, or kingdoms. For example, a polynucleotide having a consensus

sequence from a gene family of *Zea mays* can be used to generate antibody or nucleic acid probes or primers to other *Gramineae* species such as wheat, rice, or sorghum.

Alternatively, a polynucleotide having a consensus sequence generated from orthologous genes can be used to identify or isolate orthologs of other taxa. Typically, a

5 polynucleotide having a consensus sequence will be at least 9, 10, 15, 20, 25, 30, or 40 amino acids in length, or 20, 30, 40, 50, 100, or 150 nucleotides in length. As those of skill in the art are aware, a conservative amino acid substitution can be used for amino acids which differ amongst aligned sequence but are from the same conservative substitution group as discussed above. Optionally, no more than 1 or 2 conservative amino
10 acids are substituted for each 10 amino acid length of consensus sequence.

Similar sequences used for generation of a consensus or generic sequence include any number and combination of allelic variants of the same gene, orthologous, or paralogous sequences as provided herein. Optionally, similar sequences used in generating a consensus or generic sequence are identified using the BLAST algorithm's smallest sum
15 probability (P(N)). Various suppliers of sequence-analysis software are listed in chapter 7 of *Current Protocols in Molecular Biology*, F.M. Ausubel *et al.*, Eds., Current Protocols, a joint venture between Greene Publishing Associates, Inc. and John Wiley & Sons, Inc. (Supplement 30). A polynucleotide sequence is considered similar to a reference sequence if the smallest sum probability in a comparison of the test nucleic acid to the reference
20 nucleic acid is less than about 0.1, more preferably less than about 0.01, or 0.001, and most preferably less than about 0.0001, or 0.00001. Similar polynucleotides can be aligned and a consensus or generic sequence generated using multiple sequence alignment software available from a number of commercial suppliers such as the Genetics Computer Group's (Madison, WI) PILEUP software, Vector NTI's (North Bethesda, MD) ALIGNX, or
25 Genecode's (Ann Arbor, MI) SEQUENCHER. Conveniently, default parameters of such software can be used to generate consensus or generic sequences.

Machine Applications

The present invention provides machines, data structures, and processes for
30 modeling or analyzing the polynucleotides and polypeptides of the present invention.

A. Machines: Data, Data Structures, Processes, and Functions

The present invention provides a machine having a memory comprising: 1) data representing a sequence of a polynucleotide or polypeptide of the present invention, 2) a

data structure which reflects the underlying organization and structure of the data and facilitates program access to data elements corresponding to logical sub-components of the sequence, 3) processes for effecting the use, analysis, or modeling of the sequence, and 4) optionally, a function or utility for the polynucleotide or polypeptide. Thus, the present invention provides a memory for storing data that can be accessed by a computer programmed to implement a process for effecting the use, analyses, or modeling of a sequence of a polynucleotide, with the memory comprising data representing the sequence of a polynucleotide of the present invention.

The machine of the present invention is typically a digital computer. The term "computer" includes one or several desktop or portable computers, computer workstations, servers (including intranet or internet servers), mainframes, and any integrated system comprising any of the above irrespective of whether the processing, memory, input, or output of the computer is remote or local, as well as any networking interconnecting the modules of the computer. The term "computer" is exclusive of computers of the United States Patent and Trademark Office or the European Patent Office when data representing the sequence of polypeptides or polynucleotides of the present invention is used for patentability searches.

The present invention contemplates providing as data a sequence of a polynucleotide of the present invention embodied in a computer readable medium. As those of skill in the art will be aware, the form of memory of a machine of the present invention, or the particular embodiment of the computer readable medium, are not critical elements of the invention and can take a variety of forms. The memory of such a machine includes, but is not limited to, ROM, or RAM, or computer readable media such as, but not limited to, magnetic media such as computer disks or hard drives, or media such as CD-ROMs, DVDs, and the like.

The present invention further contemplates providing a data structure that is also contained in memory. The data structure may be defined by the computer programs that define the processes (see below) or it may be defined by the programming of separate data storage and retrieval programs subroutines, or systems. Thus, the present invention provides a memory for storing a data structure that can be accessed by a computer programmed to implement a process for effecting the use, analysis, or modeling of a sequence of a polynucleotide. The memory comprises data representing a polynucleotide having the sequence of a polynucleotide of the present invention. The data is stored within memory. Further, a data structure, stored within memory, is associated with the data

reflecting the underlying organization and structure of the data to facilitate program access to data elements corresponding to logical sub-components of the sequence. The data structure enables the polynucleotide to be identified and manipulated by such programs.

In a further embodiment, the present invention provides a data structure that
5 contains data representing a sequence of a polynucleotide of the present invention stored within a computer readable medium. The data structure is organized to reflect the logical structuring of the sequence, so that the sequence is easily analyzed by software programs capable of accessing the data structure. In particular, the data structures of the present invention organize the reference sequences of the present invention in a manner which
10 allows software tools to perform a wide variety of analyses using logical elements and sub-elements of each sequence.

An example of such a data structure resembles a layered hash table, where in one dimension the base content of the sequence is represented by a string of elements A, T, C, G and N. The direction from the 5' end to the 3' end is reflected by the order from the
15 position 0 to the position of the length of the string minus one. Such a string, corresponding to a nucleotide sequence of interest, has a certain number of substrings, each of which is delimited by the string position of its 5' end and the string position of its 3' end within the parent string. In a second dimension, each substring is associated with or pointed to one or multiple attribute fields. Such attribute fields contain annotations to the
20 region on the nucleotide sequence represented by the substring.

For example, a sequence under investigation is 520 bases long and represented by a string named SeqTarget. There is a minor groove in the 5' upstream non-coding region from position 12 to 38, which is identified as a binding site for an enhancer protein HM-A, which in turn will increase the transcription of the gene represented by SeqTarget. Here,
25 the substring is represented as (12, 38) and has the following attributes: [upstream uncoded], [minor groove], [HM-A binding] and [increase transcription upon binding by HM-A]. Similarly, other types of information can be stored and structured in this manner, such as information related to the whole sequence, *e.g.*, whether the sequence is a full length viral gene, a mammalian house keeping gene or an EST from clone X, information
30 related to the 3' down stream non-coding region, *e.g.*, hair pin structure, and information related to various domains of the coding region, *e.g.*, Zinc finger.

This data structure is an open structure and is robust enough to accommodate newly generated data and acquired knowledge. Such a structure is also a flexible structure. It can be trimmed down to a 1-D string to facilitate data mining and analysis steps, such as

clustering, repeat-masking, and HMM analysis. Meanwhile, such a data structure also can extend the associated attributes into multiple dimensions. Pointers can be established among the dimensioned attributes when needed to facilitate data management and processing in a comprehensive genomics knowledgebase. Furthermore, such a data structure is object-oriented. Polymorphism can be represented by a family or class of sequence objects, each of which has an internal structure as discussed above. The common traits are abstracted and assigned to the parent object, whereas each child object represents a specific variant of the family or class. Such a data structure allows data to be efficiently retrieved, updated and integrated by the software applications associated with the sequence database and/or knowledgebase.

The present invention contemplates providing processes for effecting analysis and modeling, which are described in the following section.

Optionally, the present invention further contemplates that the machine of the present invention will embody in some manner a utility or function for the polynucleotide or polypeptide of the present invention. The function or utility of the polynucleotide or polypeptide can be a function or utility for the sequence data, *per se*, or of the tangible material. Exemplary function or utilities include the name (per International Union of Biochemistry and Molecular Biology rules of nomenclature) or function of the enzyme or protein represented by the polynucleotide or polypeptide of the present invention; the metabolic pathway of the protein represented by the polynucleotide or polypeptide of the present invention; the substrate or product or structural role of the protein represented by the polynucleotide or polypeptide of the present invention; or, the phenotype (e.g., an agronomic or pharmacological trait) affected by modulating expression or activity of the protein represented by the polynucleotide or polypeptide of the present invention.

B. Computer Analysis and Modeling

The present invention provides a process of modeling and analyzing data representative of a polynucleotide or polypeptide sequence of the present invention. The process comprises entering sequence data of a polynucleotide or polypeptide of the present invention into a machine having a hardware or software sequence modeling and analysis system, developing data structures to facilitate access to the sequence data, manipulating the data to model or analyze the structure or activity of the polynucleotide or polypeptide, and displaying the results of the modeling or analysis. Thus, the present invention provides a process for effecting the use, analysis, or modeling of a polynucleotide

sequence or its derived peptide sequence through use of a computer having a memory. The process comprises 1) placing into the memory data representing a polynucleotide having the sequence of a polynucleotide of the present invention, developing within the memory a data structure associated with the data and reflecting the underlying organization and structure of the data to facilitate program access to data elements corresponding to logical sub-components of the sequence, 2) programming the computer with a program containing instructions sufficient to implement the process for effecting the use, analysis, or modeling of the polynucleotide sequence or the peptide sequence, and, 3) executing the program on the computer while granting the program access to the data and to the data structure within the memory.

A variety of modeling and analytic tools are well known in the art and available commercially. Included amongst the modeling/analysis tools are methods to: 1) recognize overlapping sequences (e.g., from a sequencing project) with a polynucleotide of the present invention and create an alignment called a "contig"; 2) identify restriction enzyme sites of a polynucleotide of the present invention; 3) identify the products of a T1 ribonuclease digestion of a polynucleotide of the present invention; 4) identify PCR primers with minimal self-complementarity; 5) compute pairwise distances between sequences in an alignment, reconstruct phylogentic trees using distance methods, and calculate the degree of divergence of two protein coding regions; 6) identify patterns such as coding regions, terminators, repeats, and other consensus patterns in polynucleotides of the present invention; 7) identify RNA secondary structure; 8) identify sequence motifs, isoelectric point, secondary structure, hydrophobicity, and antigenicity in polypeptides of the present invention; 9) translate polynucleotides of the present invention and backtranslate polypeptides of the present invention; and 10) compare two protein or nucleic acid sequences and identifying points of similarity or dissimilarity between them.

The processes for effecting analysis and modeling can be produced independently or obtained from commercial suppliers. Exemplary analysis and modeling tools are provided in products such as InforMax's (Bethesda, MD) Vector NTI Suite (Version 5.5), Intelligenetics' (Mountain View, CA) PC/Gene program, and Genetics Computer Group's (Madison, WI) Wisconsin Package (Version 10.0); these tools, and the functions they perform, (as provided and disclosed by the programs and accompanying literature) are incorporated herein by reference and are described in more detail in section C which follows.

Thus, in a further embodiment, the present invention provides a machine-readable medium containing a computer program and data, comprising a program stored on the medium containing instructions sufficient to implement a process for effecting the use, analysis, or modeling of a representation of a polynucleotide or peptide sequence. The data stored on the medium represents a sequence of a polynucleotide having the sequence of a polynucleotide of the present invention. The medium also includes a data structure reflecting the underlying organization and structure of the data to facilitate program access to data elements corresponding to logical sub-components of the sequence, the data structure being inherent in the program and in the way in which the program organizes and accesses the data.

C. Homology Searches

As an example of such a comparative analysis, the present invention provides a process of identifying a candidate homologue (i.e., an ortholog or paralog) of a polynucleotide or polypeptide of the present invention. The process comprises entering sequence data of a polynucleotide or polypeptide of the present invention into a machine having a hardware or software sequence analysis system, developing data structures to facilitate access to the sequence data, manipulating the data to analyze the structure the polynucleotide or polypeptide, and displaying the results of the analysis. A candidate homologue has statistically significant probability of having the same biological function (e.g., catalyzes the same reaction, binds to homologous proteins/nucleic acids, has a similar structural role) as the reference sequence to which it is compared. Accordingly, the polynucleotides and polypeptides of the present invention have utility in identifying homologs in animals or other plant species, particularly those in the family *Gramineae* such as, but not limited to, sorghum, wheat, or rice.

The process of the present invention comprises obtaining data representing a polynucleotide or polypeptide test sequence. Test sequences can be obtained from a nucleic acid of an animal or plant. Test sequences can be obtained directly or indirectly from sequence databases including, but not limited to, those such as: GenBank, EMBL, GenSeq, SWISS-PROT, or those available on-line via the UK Human Genome Mapping Project (HGMP) GenomeWeb. In some embodiments the test sequence is obtained from a plant species other than maize whose function is uncertain but will be compared to the test sequence to determine sequence similarity or sequence identity. The test sequence data is entered into a machine, such as a computer, containing: i) data representing a reference

sequence and, ii) a hardware or software sequence comparison system to compare the reference and test sequence for sequence similarity or identity.

Exemplary sequence comparison systems are provided for in sequence analysis software such as those provided by the Genetics Computer Group (Madison, WI) or InforMax (Bethesda, MD), or Intelligenetics (Mountain View, CA). Optionally, sequence comparison is established using the BLAST or GAP suite of programs. Generally, a smallest sum probability value ($P(N)$) of less than 0.1, or alternatively, less than 0.01, 0.001, 0.0001, or 0.00001 using the BLAST 2.0 suite of algorithms under default parameters identifies the test sequence as a candidate homologue (i.e., an allele, ortholog, or paralog) of the reference sequence. Those of skill in the art will recognize that a candidate homologue has an increased statistical probability of having the same or similar function as the gene/protein represented by the test sequence.

The reference sequence can be the sequence of a polypeptide or a polynucleotide of the present invention. The reference or test sequence is each optionally at least 25 amino acids or at least 100 nucleotides in length. The length of the reference or test sequences can be the length of the polynucleotide or polypeptide described, respectively, above in the sections entitled "Nucleic Acids" (particularly section (g)), and "Proteins". As those of skill in the art are aware, the greater the sequence identity/similarity between a reference sequence of known function and a test sequence, the greater the probability that the test sequence will have the same or similar function as the reference sequence. The results of the comparison between the test and reference sequences are outputted (e.g., displayed, printed, recorded) via any one of a number of output devices and/or media (e.g., computer monitor, hard copy, or computer readable medium).

25 Detection of Nucleic Acids

The present invention further provides methods for detecting a polynucleotide of the present invention in a nucleic acid sample suspected of containing a polynucleotide of the present invention, such as a plant cell lysate, particularly a lysate of maize. In some embodiments, a cognate gene of a polynucleotide of the present invention or portion thereof can be amplified prior to the step of contacting the nucleic acid sample with a polynucleotide of the present invention. The nucleic acid sample is contacted with the polynucleotide to form a hybridization complex. The polynucleotide hybridizes under stringent conditions to a gene encoding a polypeptide of the present invention. Formation of the hybridization complex is used to detect a gene encoding a polypeptide of the present

invention in the nucleic acid sample. Those of skill will appreciate that an isolated nucleic acid comprising a polynucleotide of the present invention should lack cross-hybridizing sequences in common with non-target genes that would yield a false positive result.

Detection of the hybridization complex can be achieved using any number of well known methods. For example, the nucleic acid sample, or a portion thereof, may be assayed by hybridization formats including but not limited to, solution phase, solid phase, mixed phase, or *in situ* hybridization assays.

Detectable labels suitable for use in the present invention include any composition detectable by spectroscopic, radioisotopic, photochemical, biochemical, immunochemical, electrical, optical or chemical means. Useful labels in the present invention include biotin for staining with labeled streptavidin conjugate, magnetic beads, fluorescent dyes, radiolabels, enzymes, and colorimetric labels. Other labels include ligands which bind to antibodies labeled with fluorophores, chemiluminescent agents, and enzymes. Labeling the nucleic acids of the present invention is readily achieved such as by the use of labeled PCR primers.

Although the present invention has been described in some detail by way of illustration and example for purposes of clarity of understanding, it will be obvious that certain changes and modifications may be practiced within the scope of the appended claims.

Example 1

This example describes the construction of a cDNA library.

Total RNA can be isolated from maize tissues with TRIzol Reagent (Life Technology Inc. Gaithersburg, MD) using a modification of the guanidine isothiocyanate/acid-phenol procedure described by Chomczynski and Sacchi (Chomczynski, P., and Sacchi, N. *Anal. Biochem.* 162, 156 (1987)). In brief, plant tissue samples are pulverized in liquid nitrogen before the addition of the TRIzol Reagent, and then further homogenized with a mortar and pestle. Addition of chloroform followed by centrifugation is conducted for separation of an aqueous phase and an organic phase. The total RNA is recovered by precipitation with isopropyl alcohol from the aqueous phase.

The selection of poly(A)⁺ RNA from total RNA can be performed using PolyATact system (Promega Corporation, Madison, WI). Biotinylated oligo(dT) primers are used to hybridize to the 3' poly(A) tails on mRNA. The hybrids are captured using streptavidin coupled to paramagnetic particles and a magnetic separation stand. The

mRNA is then washed at high stringency conditions and eluted by RNase-free deionized water.

cDNA synthesis and construction of unidirectional cDNA libraries can be accomplished using the SuperScript Plasmid System (Life Technology Inc. Gaithersburg, MD). The first strand of cDNA is synthesized by priming an oligo(dT) primer containing a Not I site. The reaction is catalyzed by SuperScript Reverse Transcriptase II at 45°C. The second strand of cDNA is labeled with alpha-³²P-dCTP and a portion of the reaction analyzed by agarose gel electrophoresis to determine cDNA sizes. cDNA molecules smaller than 500 base pairs and unligated adapters are removed by Sephacryl-S400 chromatography. The selected cDNA molecules are ligated into pSPORT1 vector in between of Not I and Sal I sites.

Alternatively, cDNA libraries can be prepared by any one of many methods available. For example, the cDNAs may be introduced into plasmid vectors by first preparing the cDNA libraries in Uni-ZAP™ XR vectors according to the manufacturer's protocol (Stratagene Cloning Systems, La Jolla, CA). The Uni-ZAP™ XR libraries are converted into plasmid libraries according to the protocol provided by Stratagene. Upon conversion, cDNA inserts will be contained in the plasmid vector pBluescript. In addition, the cDNAs may be introduced directly into precut Bluescript II SK(+) vectors (Stratagene) using T4 DNA ligase (New England Biolabs), followed by transfection into DH10B cells according to the manufacturer's protocol (GIBCO BRL Products). Once the cDNA inserts are in plasmid vectors, plasmid DNAs are prepared from randomly picked bacterial colonies containing recombinant pBluescript plasmids, or the insert cDNA sequences are amplified via polymerase chain reaction using primers specific for vector sequences flanking the inserted cDNA sequences. Amplified insert DNAs or plasmid DNAs are sequenced in dye-primer sequencing reactions to generate partial cDNA sequences (expressed sequence tags or "ESTs"; see Adams *et al.*, (1991) *Science* 252:1651-1656). The resulting ESTs are analyzed using a Perkin Elmer Model 377 fluorescent sequencer.

Example 2

This method describes construction of a full-length enriched cDNA library.

An enriched full-length cDNA library can be constructed using one of two variations of the method of Carninci *et al.* *Genomics* 37: 327-336, 1996. These variations are based on chemical introduction of a biotin group into the diol residue of the 5' cap structure of eukaryotic mRNA to select full-length first strand cDNA. The selection

occurs by trapping the biotin residue at the cap sites using streptavidin-coated magnetic beads followed by RNase I treatment to eliminate incompletely synthesized cDNAs.

Second strand cDNA is synthesized using established procedures such as those provided in Life Technologies' (Rockville, MD) "SuperScript Plasmid System for cDNA Synthesis and Plasmid Cloning" kit. Libraries made by this method have been shown to contain
5 50% to 70% full-length cDNAs.

The first strand synthesis methods are detailed below. An asterisk denotes that the reagent was obtained from Life Technologies, Inc.

10 *A. First strand cDNA synthesis method 1 (with trehalose)*

	mRNA (10ug)	25μl
	*Not I primer (5ug)	10μl
	*5x 1 st strand buffer	43μl
	*0.1m DTT	20μl
15	*dNTP mix 10mm	10μl
	BSA 10ug/μl	1μl
	Trehalose (saturated)	59.2μl
	RNase inhibitor (Promega)	1.8μl
	*Superscript II RT 200u/μl	20μl
20	100 % glycerol	18μl
	Water	7μl

The mRNA and Not I primer are mixed and denatured at 65°C for 10 min. They are then chilled on ice and other components added to the tube. Incubation is at 45°C for 2
25 min. Twenty microliters of RT (reverse transcriptase) is added to the reaction and start program on the thermocycler (MJ Research, Waltham, MA):

	Step 1	45°C	10min
	Step 2	45°C	-0.3°C/cycle , 2 seconds/cycle
	Step 3	go to 2 for 33 cycles	
30	Step 4	35°C	5min
	Step 5	45°C	5min
	Step 6	45°C	0.2°C/cycle, 1 sec/cycle
	Step 7	go to 7 for 49 cycles	

- | | |
|-----------|--------------------------------|
| Step 8 | 55°C 0.1°C/cycle, 12 sec/cycle |
| Step 9 | go to 8 for 49 cycles |
| Step 10 | 55°C 2min |
| Step11 | 60°C 2min |
| 5 Step 12 | go to 11 for 9 times |
| Step 13 | 4°C forever |
| Step14 | end |

B. First strand cDNA synthesis method 2

- | | | |
|----|--|------|
| 10 | mRNA (10µg) | 25µl |
| | water | 30µl |
| | *Not I adapter primer (5µg) | 10µl |
| | 65°C for 10min, chill on ice, then add following reagents, | |
| | *5x first buffer | 20µl |
| 15 | *0.1M DTT | 10µl |
| | *10mM dNTP mix | 5µl |

Incubate at 45°C for 2min, then add 10µl of *Superscript II RT (200u/µl), start the following program:

- | | | |
|----|--------|----------------------------------|
| 20 | Step 1 | 45°C for 6 sec, -0.1°C/cycle |
| | Step 2 | go to 1 for 99 additional cycles |
| | Step 3 | 35°C for 5min |
| | Step 4 | 45°C for 60 min |
| | Step 5 | 50°C for 10 min |
| 25 | Step 6 | 4°C forever |
| | Step 7 | end |

After the 1st strand cDNA synthesis, the DNA is extracted by phenol according to standard procedures, and then precipitated in NaOAc and ethanol, and stored in -20°C.

C. Oxidization of the diol group of mRNA for biotin labeling

First strand cDNA is spun down and washed once with 70% EtOH. The pellet resuspended in 23.2 μ l of DEPC treated water and put on ice. Prepare 100 mM of NaIO4 freshly, and then add the following reagents:

mRNA:1 st cDNA (start with 20µg mRNA)	46.4µl
100mM NaIO ₄ (freshly made)	2.5µl
NaOAc 3M pH4.5	1.1µl

- 5 To make 100 mM NaIO₄, use 21.39µg of NaIO₄ for 1µl of water.

Wrap the tube in a foil and incubate on ice for 45min.

After the incubation, the reaction is then precipitated in:

5M NaCl	10µl
20%SDS	0.5µl

- 10 isopropanol 61µl

Incubate on ice for at least 30 min, then spin it down at max speed at 4°C for 30 min and wash once with 70% ethanol and then 80% EtOH.

D. Biotinylation of the mRNA diol group

- 15 Resuspend the DNA in 110µl DEPC treated water, then add the following reagents:

20% SDS	5 µl
2 M NaOAc pH 6.1	5 µl
10mm biotin hydrazide (freshly made)	300 µl

Wrap in a foil and incubate at room temperature overnight.

20

E. RNase I treatment

Precipitate DNA in:

5M NaCl	10µl
2M NaOAc pH 6.1	75µl
25 biotinylated mRNA:cDNA	420µl
100% EtOH (2.5Vol)	1262.5µl

(Perform this precipitation in two tubes and split the 420 µl of DNA into 210 µl each, add 5µl of 5M NaCl, 37.5µl of 2M NaOAc pH 6.1, and 631.25 µl of 100% EtOH).

- 30 Store at -20°C for at least 30 min. Spin the DNA down at 4°C at maximal speed for 30 min. and wash with 80% EtOH twice, then dissolve DNA in 70µl RNase free water. Pool two tubes and end up with 140 µl.

Add the following reagents:

- 55 -

RNase One 10U/ μ l	40 μ l
1 st cDNA:RNA	140 μ l
10X buffer	20 μ l

Incubate at 37°C for 15min.

- 5 Add 5 μ l of 40 μ g/ μ l yeast tRNA to each sample for capturing.

F. Full length 1st cDNA capturing

Blocking the beads with yeast tRNA:

Beads	1ml
10 Yeast tRNA 40 μ g/ μ l	5 μ l

Incubate on ice for 30min with mixing, wash 3 times with 1ml of 2M NaCl , 50mmEDTA, pH 8.0.

Resuspend the beads in 800 μ l of 2M NaCl , 50mm EDTA, pH 8.0, add RNase I treated sample 200 μ l, and incubate the reaction for 30min at room temperature.

- 15 Capture the beads using the magnetic stand, save the supernatant, and start following washes:

2 washes with 2M NaCl , 50mm EDTA, pH 8.0, 1 ml each time,

1 wash with 0.4% SDS, 50 μ g/ml tRNA,

1 wash with 10mm Tris-Cl pH 7.5, 0.2mm EDTA, 10mm NaCl, 20% glycerol,

- 20 1 wash with 50 μ g/ml tRNA,

1 wash with 1st cDNA buffer

G. Second strand cDNA synthesis

Resuspend the beads in:

25	*5X first buffer	8 μ l
	*0.1mM DTT	4 μ l
	*10mm dNTP mix	8 μ l
	*5X 2nd buffer	60 μ l
	*E.coli Ligase 10U/ μ l	2 μ l
30	*E.coli DNA polymerase 10U/ μ l	8 μ l
	*E. coli RNaseH 2U/ μ l	2 μ l
	P32 dCTP 10 μ ci/ μ l	2 μ l
	Or water up to 300 μ l	208 μ l

Incubate at 16°C for 2hr with mixing the reaction in every 30 min.

Add 4µl of T4 DNA polymerase and incubate for additional 5 min at 16°C.

Elute 2nd cDNA from the beads.

- 5 Use a magnetic stand to separate the 2nd cDNA from the beads, then resuspend the beads in 200µl of water, and then separate again, pool the samples (about 500µl).

Add 200 µl of water to the beads, then 200µl of phenol:chloroform, vortex, and spin to separate the sample with phenol.

- 10 Pool the DNA together (about 700µl) and use phenol to clean the DNA again, DNA is then precipitated in 2µg of glycogen and 0.5 vol of 7.5M NH₄OAc and 2 vol of 100% EtOH. Precipitate overnight. Spin down the pellet and wash with 70% EtOH, air-dry the pellet.

	DNA	250µl	DNA	200µl
15	7.5M NH ₄ OAc	125µl	7.5M NH ₄ OAc	100µl
	100% EtOH	750µl	100% EtOH	600µl
	glycogen 1µg/µl	2µl	glycogen 1µg/µl	2µl

H. *Sal I* adapter ligation

- 20 Resuspend the pellet in 26 µl of water and use 1µl for TAE gel.

Set up reaction as following:

	2 nd strand cDNA	25µl
	*5X T4 DNA ligase buffer	10µl
25	*Sal I adapters	10µl
	*T4 DNA ligase	5µl

Mix gently, incubate the reaction at 16°C overnight.

Add 2µl of ligase second day and incubate at room temperature for 2 hrs (optional).

- 30 Add 50µl water to the reaction and use 100µl of phenol to clean the DNA, 90µl of the upper phase is transferred into a new tube and precipitate in:

	Glycogen 1µg/µl	2µl
	Upper phase DNA	90µl

7.5M NH₄OAc 50μl

100% EtOH 300μl

precipitate at -20°C overnight

Spin down the pellet at 4°C and wash in 70% EtOH, dry the pellet.

5

I. Not I digestion

2nd cDNA 41μl

*Reaction 3 buffer 5μl

*Not I 15u/μl 4μl

10 Mix gently and incubate the reaction at 37°C for 2hr.

Add 50 μl of water and 100μl of phenol, vortex , and take 90μl of the upper phase to a new tube, then add 50μl of NH₄OAc and 300 μl of EtOH. Precipitate overnight at -20°C.

15 Cloning, ligation, and transformation are performed per the Superscript cDNA synthesis kit.

Example 3

This example describes cDNA sequencing and library subtraction.

20 Individual colonies can be picked and DNA prepared either by PCR with M13 forward primers and M13 reverse primers, or by plasmid isolation. cDNA clones can be sequenced using M13 reverse primers.

25 cDNA libraries are plated out on 22 x 22 cm² agar plate at density of about 3,000 colonies per plate. The plates are incubated in a 37°C incubator for 12-24 hours. Colonies are picked into 384-well plates by a robot colony picker, Q-bot (GENETIX Limited). These plates are incubated overnight at 37°C. Once sufficient colonies are picked, they are pinned onto 22 x 22 cm² nylon membranes using Q-bot. Each membrane holds 9,216 or 36,864 colonies. These membranes are placed onto an agar plate with an appropriate antibiotic. The plates are incubated at 37°C overnight.

30 After colonies are recovered on the second day, these filters are placed on filter paper prewetted with denaturing solution for four minutes, then incubated on top of a boiling water bath for an additional four minutes. The filters are then placed on filter paper prewetted with neutralizing solution for four minutes. After excess solution is removed by placing the filters on dry filter papers for one minute, the colony side of the

filters is placed into Proteinase K solution, incubated at 37°C for 40-50 minutes. The filters are placed on dry filter papers to dry overnight. DNA is then cross-linked to nylon membrane by UV light treatment.

Colony hybridization is conducted as described by Sambrook, J., Fritsch, E.F. and Maniatis, T., (in *Molecular Cloning: A laboratory Manual*, 2nd Edition). The following probes can be used in colony hybridization:

1. First strand cDNA from the same tissue as the library was made from to remove the most redundant clones.

2. 48-192 most redundant cDNA clones from the same library based on previous sequencing data.

3. 192 most redundant cDNA clones in the entire maize sequence database.

4. A Sal-A20 oligo nucleotide: TCG ACC CAC GCG TCC GAA AAA AAA AAA AAA AAA AAA, removes clones containing a poly A tail but no cDNA.

5. cDNA clones derived from rRNA.

The image of the autoradiography is scanned into computer and the signal intensity and cold colony addresses of each colony is analyzed. Re-arraying of cold-colonies from 384 well plates to 96 well plates is conducted using Q-bot.

Example 4

This example describes identification of the gene from a computer homology search.

Gene identities can be determined by conducting BLAST (Basic Local Alignment Search Tool; Altschul, S. F., et al., (1993) *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/) searches under default parameters for similarity to sequences contained in the BLAST "nr" database (comprising all non-redundant GenBank CDS translations, sequences derived from the 3-dimensional structure Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences are analyzed for similarity to all publicly available DNA sequences contained in the "nr" database using the BLASTN algorithm.

The DNA sequences are translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the "nr" database using the BLASTX algorithm (Gish, W. and States, D. J. *Nature Genetics* 3:266-272 (1993)) provided by the NCBI. In some cases, the sequencing data from two or more clones containing overlapping segments of DNA are used to construct contiguous DNA sequences.

Sequence alignments and percent identity calculations can be performed using the Megalign program of the LASERGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Multiple alignment of the sequences can be performed using the Clustal method of alignment (Higgins and Sharp (1989) *CABIOS*. 5:151-153) with the default parameters (GAP PENALTY=10, GAP LENGTH PENALTY=10). Default parameters for pairwise alignments using the Clustal method are KTUPLE 1, GAP PENALTY=3, WINDOW=5 and DIAGONALS SAVED=5.

Example 5

10 This example describes expression of transgenes in monocot cells.

A transgene comprising a cDNA encoding the instant polypeptides in sense orientation with respect to the maize 27 kD zein promoter that is located 5' to the cDNA fragment, and the 10 kD zein 3' end that is located 3' to the cDNA fragment, can be constructed. The cDNA fragment of this gene may be generated by polymerase chain reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites (NcoI or SmaI) can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the digested vector pML103 as described below. Amplification is then performed in a standard PCR. The amplified DNA is then digested with restriction enzymes NcoI and SmaI and fractionated on an agarose gel. The appropriate band can be isolated from the gel and combined with a 4.9 kb NcoI-SmaI fragment of the plasmid pML103. Plasmid pML103 has been deposited under the terms of the Budapest Treaty at ATCC (American Type Culture Collection, 10801 University Blvd., Manassas, VA 20110-2209), and bears accession number ATCC 97366. The DNA segment from pML103 contains a 1.05 kb SalI-NcoI promoter fragment of the maize 27 kD zein gene and a 0.96 kb SmaI-SalI fragment from the 3' end of the maize 10 kD zein gene in the vector pGem9Zf(+) (Promega). Vector and insert DNA can be ligated at 15°C overnight, essentially as described (Maniatis). The ligated DNA may then be used to transform *E. coli* XL1-Blue (Epicurian Coli XL-1 Blue; Stratagene). Bacterial transformants can be screened by restriction enzyme digestion of plasmid DNA and limited nucleotide sequence analysis using the dideoxy chain termination method (Sequenase DNA Sequencing Kit; U. S. Biochemical). The resulting plasmid construct would comprise a transgene encoding, in the 5' to 3' direction, the maize 27 kD zein promoter, a cDNA fragment encoding the instant polypeptides, and the 10 kD zein 3' region.

The transgene described above can then be introduced into maize cells by the following procedure. Immature maize embryos can be dissected from developing caryopses derived from crosses of the inbred maize lines H99 and LH132. The embryos are isolated 10 to 11 days after pollination when they are 1.0 to 1.5 mm long. The embryos are then placed with the axis-side facing down and in contact with agarose-solidified N6 medium (Chu *et al.* (1975) *Sci. Sin. Peking* 18:659-668). The embryos are kept in the dark at 27°C. Friable embryogenic callus consisting of undifferentiated masses of cells with somatic proembryoids and embryoids borne on suspensor structures proliferates from the scutellum of these immature embryos. The embryogenic callus isolated from the primary explant can be cultured on N6 medium and sub-cultured on this medium every 2 to 3 weeks.

The plasmid, p35S/Ac (Hoechst Ag, Frankfurt, Germany) or equivalent may be used in transformation experiments in order to provide for a selectable marker. This plasmid contains the *Pat* gene (see European Patent Publication 0 242 236) which encodes phosphinothricin acetyl transferase (PAT). The enzyme PAT confers resistance to herbicidal glutamine synthetase inhibitors such as phosphinothricin. The *pat* gene in p35S/Ac is under the control of the 35S promoter from Cauliflower Mosaic Virus (Odell *et al.* (1985) *Nature* 313:810-812) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*.

The particle bombardment method (Klein *et al.* (1987) *Nature* 327:70-73) may be used to transfer genes to the callus culture cells. According to this method, gold particles (1 μm in diameter) are coated with DNA using the following technique. Ten μg of plasmid DNAs are added to 50 μL of a suspension of gold particles (60 mg per mL). Calcium chloride (50 μL of a 2.5 M solution) and spermidine free base (20 μL of a 1.0 M solution) are added to the particles. The suspension is vortexed during the addition of these solutions. After 10 minutes, the tubes are briefly centrifuged (5 sec at 15,000 rpm) and the supernatant removed. The particles are resuspended in 200 μL of absolute ethanol, centrifuged again and the supernatant removed. The ethanol rinse is performed again and the particles resuspended in a final volume of 30 μL of ethanol. An aliquot (5 μL) of the DNA-coated gold particles can be placed in the center of a Kapton flying disc (Bio-Rad Labs). The particles are then accelerated into the maize tissue with a Biolistic PDS-1000/He (Bio-Rad Instruments, Hercules CA), using a helium pressure of 1000 psi, a gap distance of 0.5 cm and a flying distance of 1.0 cm.

For bombardment, the embryogenic tissue is placed on filter paper over agarose-solidified N6 medium. The tissue is arranged as a thin lawn and covers a circular area of about 5 cm in diameter. The petri dish containing the tissue can be placed in the chamber of the PDS-1000/He approximately 8 cm from the stopping screen. The air in the chamber is then evacuated to a vacuum of 28 inches of Hg. The macrocarrier is accelerated with a helium shock wave using a rupture membrane that bursts when the He pressure in the shock tube reaches 1000 psi.

Seven days after bombardment the tissue can be transferred to N6 medium that contains glufosinate (2 mg per liter) and lacks casein or proline. The tissue continues to grow slowly on this medium. After an additional 2 weeks the tissue can be transferred to fresh N6 medium containing glufosinate. After 6 weeks, areas of about 1 cm in diameter of actively growing callus can be identified on some of the plates containing the glufosinate-supplemented medium. These calli may continue to grow when sub-cultured on the selective medium.

Plants can be regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the tissue can be transferred to regeneration medium (Fromm *et al.* (1990) *Bio/Technology* 8:833-839).

Example 6

This example describes expression of transgenes in dicot cells.

A seed-specific expression cassette composed of the promoter and transcription terminator from the gene encoding the β subunit of the seed storage protein phaseolin from the bean *Phaseolus vulgaris* (Doyle *et al.* (1986) *J. Biol. Chem.* 261:9228-9238) can be used for expression of the instant polypeptides in transformed soybean. The phaseolin cassette includes about 500 nucleotides upstream (5') from the translation initiation codon and about 1650 nucleotides downstream (3') from the translation stop codon of phaseolin. Between the 5' and 3' regions are the unique restriction endonuclease sites Nco I (which includes the ATG translation initiation codon), SmaI, KpnI and XbaI. The entire cassette is flanked by Hind III sites.

The cDNA fragment of this gene may be generated by polymerase chain reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the expression vector. Amplification is then performed as described

above, and the isolated fragment is inserted into a pUC18 vector carrying the seed expression cassette.

Soybean embryos may then be transformed with the expression vector comprising sequences encoding the instant polypeptides. To induce somatic embryos, cotyledons, 3-5 mm in length dissected from surface sterilized, immature seeds of the soybean cultivar A2872, can be cultured in the light or dark at 26°C on an appropriate agar medium for 6-10 weeks. Somatic embryos which produce secondary embryos are then excised and placed into a suitable liquid medium. After repeated selection for clusters of somatic embryos which multiplied as early, globular staged embryos, the suspensions are maintained as described below.

Soybean embryogenic suspension cultures can be maintained in 35 mL liquid media on a rotary shaker, 150 rpm, at 26°C with florescent lights on a 16:8 hour day/night schedule. Cultures are subcultured every two weeks by inoculating approximately 35 mg of tissue into 35 mL of liquid medium.

Soybean embryogenic suspension cultures may then be transformed by the method of particle gun bombardment (Klein *et al.* (1987) *Nature* (London) 327:70-73, U.S. Patent No. 4,945,050). A Du Pont Biolistic PDS1000/HE instrument (helium retrofit) can be used for these transformations.

A selectable marker gene which can be used to facilitate soybean transformation is a transgene composed of the 35S promoter from Cauliflower Mosaic Virus (Odell *et al.* (1985) *Nature* 313:810-812), the hygromycin phosphotransferase gene from plasmid pJR225 (from *E. coli*; Gritz *et al.* (1983) *Gene* 25:179-188) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*. The seed expression cassette comprising the phaseolin 5' region, the fragment encoding the instant polypeptides and the phaseolin 3' region can be isolated as a restriction fragment. This fragment can then be inserted into a unique restriction site of the vector carrying the marker gene.

To 50 µL of a 60 mg/mL 1 µm gold particle suspension is added (in order): 5 µL DNA (1 µg/µL), 20 µl spermidine (0.1 M), and 50 µL CaCl₂ (2.5 M). The particle preparation is then agitated for three minutes, spun in a microfuge for 10 seconds and the supernatant removed. The DNA-coated particles are then washed once in 400 µL 70% ethanol and resuspended in 40 µL of anhydrous ethanol. The DNA/particle suspension can be sonicated three times for one second each. Five microliters of the DNA-coated gold particles are then loaded on each macro carrier disk.

Approximately 300-400 mg of a two-week-old suspension culture is placed in an empty 60x15 mm petri dish and the residual liquid removed from the tissue with a pipette. For each transformation experiment, approximately 5-10 plates of tissue are normally bombarded. Membrane rupture pressure is set at 1100 psi and the chamber is evacuated to
5 a vacuum of 28 inches mercury. The tissue is placed approximately 3.5 inches away from the retaining screen and bombarded three times. Following bombardment, the tissue can be divided in half and placed back into liquid and cultured as described above.

Five to seven days post bombardment, the liquid media may be exchanged with fresh media, and eleven to twelve days post bombardment with fresh media containing
10 50 mg/mL hygromycin. This selective media can be refreshed weekly. Seven to eight weeks post bombardment, green, transformed tissue may be observed growing from untransformed, necrotic embryogenic clusters. Isolated green tissue is removed and inoculated into individual flasks to generate new, clonally propagated, transformed embryogenic suspension cultures. Each new line may be treated as an independent
15 transformation event. These suspensions can then be subcultured and maintained as clusters of immature embryos or regenerated into whole plants by maturation and germination of individual somatic embryos.

Example 7

20 This example describes expression of a transgene in microbial cells.

The cDNAs encoding the instant polypeptides can be inserted into the T7 *E. coli* expression vector pBT430. This vector is a derivative of pET-3a (Rosenberg *et al.* (1987) *Gene* 56:125-135) which employs the bacteriophage T7 RNA polymerase/T7-promoter system. Plasmid pBT430 was constructed by first destroying the EcoR I and Hind III sites
25 in pET-3a at their original positions. An oligonucleotide adaptor containing EcoR I and Hind III sites was inserted at the BamH I site of pET-3a. This created pET-3aM with additional unique cloning sites for insertion of genes into the expression vector. Then, the Nde I site at the position of translation initiation was converted to an Nco I site using oligonucleotide-directed mutagenesis. The DNA sequence of pET-3aM in this region,
30 5'-CATATGG, was converted to 5'-CCCATGG in pBT430.

Plasmid DNA containing a cDNA may be appropriately digested to release a nucleic acid fragment encoding the protein. This fragment may then be purified on a 1% NuSieve GTG low melting agarose gel (FMC). Buffer and agarose contain 10 µg/ml ethidium bromide for visualization of the DNA fragment. The fragment can then be purified from

the agarose gel by digestion with GELase (Epicentre Technologies) according to the manufacturer's instructions, ethanol precipitated, dried and resuspended in 20 μ L of water. Appropriate oligonucleotide adapters may be ligated to the fragment using T4 DNA ligase (New England Biolabs, Beverly, MA). The fragment containing the ligated adapters can be purified from the excess adapters using low melting agarose as described above. The vector pBT430 is digested, dephosphorylated with alkaline phosphatase (NEB) and deproteinized with phenol/chloroform as described above. The prepared vector pBT430 and fragment can then be ligated at 16°C for 15 hours followed by transformation into DH5 electrocompetent cells (GIBCO BRL). Transformants can be selected on agar plates containing LB media and 100 μ g/mL ampicillin. Transformants containing the gene encoding the instant polypeptides are then screened for the correct orientation with respect to the T7 promoter by restriction enzyme analysis.

For high level expression, a plasmid clone with the cDNA insert in the correct orientation relative to the T7 promoter can be transformed into *E. coli* strain BL21(DE3) (Studier et al. (1986) *J. Mol. Biol.* 189:113-130). Cultures are grown in LB medium containing ampicillin (100 mg/L) at 25°C. At an optical density at 600 nm of approximately 1, IPTG (isopropylthio- β -galactoside, the inducer) can be added to a final concentration of 0.4 mM and incubation can be continued for 3 h at 25°. Cells are then harvested by centrifugation and re-suspended in 50 μ L of 50 mM Tris-HCl at pH 8.0 containing 0.1 mM DTT and 0.2 mM phenyl methylsulfonyl fluoride. A small amount of 1 mm glass beads can be added and the mixture sonicated 3 times for about 5 seconds each time with a microprobe sonicator. The mixture is centrifuged and the protein concentration of the supernatant determined. One microgram of protein from the soluble fraction of the culture can be separated by SDS-polyacrylamide gel electrophoresis. Gels can be observed for protein bands migrating at the expected molecular weight.

The above examples are provided to illustrate the invention but not to limit its scope. Other variants of the invention will be readily apparent to one of ordinary skill in the art and are encompassed by the appended claims. All publications, patents, patent applications, and computer programs cited herein are hereby incorporated by reference.

Deposits

Plasmids containing polynucleotide sequences of the invention were deposited on March 21, 2000, with the American Type Culture Collection (ATCC), 10801 University

Boulevard, Manassas, Virginia USA, 20110-2209, and assigned Accession Nos. PTA-1539, PTA-1540, and PTA-1541. These deposits will be maintained under the terms of the Budapest Treaty on the International Recognition of the Deposit of Microorganisms for the Purposes of Patent Procedure. In addition, during the pendency of this patent application,

5 access to the deposited cultures will be available to the Commissioner of Patents and Trademarks and to persons determined by the Commissioner to be entitled thereto under 37 C.F.R. §114 and 35 U.S.C. §122.

These deposits were made merely as a convenience for those of skill in the art and are not an admission that a deposit is required under 35 U.S.C. §112. All restrictions

10 imposed by the depositor on the availability to the public of the deposited material will be irrevocably removed upon granting of a patent. However, it should be understood that the availability of a deposit does not constitute a license to practice the subject invention in derogation of patent rights granted by government action.

WHAT IS CLAIMED IS:

1. An isolated nucleic acid comprising a member selected from the group consisting of:
 - (a) a polynucleotide having at least 80% sequence identity, as determined by the
5 GAP algorithm under default parameters, to a polynucleotide of SEQ ID NO: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, 57, 61, or 65;
 - (b) a polynucleotide encoding a polypeptide of SEQ ID NO: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, 58, 62, or 66;
 - (c) a polynucleotide amplified from a *Zea mays* nucleic acid library using primers
10 which selectively hybridize, under stringent hybridization conditions, to loci within a polynucleotide of SEQ ID NO: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, 57, 61, or 65;
 - (d) a polynucleotide which selectively hybridizes, under stringent hybridization conditions and a wash in 0.1X SSC at 60°C, to a polynucleotide of SEQ ID
15 NO: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, 57, 61, or 65;
 - (e) a polynucleotide of SEQ ID NO: 1, 5, 9, 13, 17, 21, 25, 29, 33, 37, 41, 45, 49, 53, 57, 61, or 65;
 - (f) a polynucleotide which is complementary to a polynucleotide of (a), (b), (c), (d), or (e); and
 - 20 (g) a polynucleotide comprising at least 25 contiguous nucleotides from a polynucleotide of (a), (b), (c), (d), (e), or (f).
2. A recombinant expression cassette, comprising a member of claim 1 operably
25 linked, in sense or anti-sense orientation, to a promoter.
3. A host cell comprising the recombinant expression cassette of claim 2.
4. A transgenic plant comprising a recombinant expression cassette of claim 2.
- 30 5. The transgenic plant of claim 4, wherein said plant is a monocot.
6. The transgenic plant of claim 4, wherein said plant is a dicot.

7. The transgenic plant of claim 4, wherein said plant is selected from the group consisting of: maize, soybean, sunflower, sorghum, canola, wheat, alfalfa, cotton, rice, barley, millet, peanut, and cocoa.

5 8. A transgenic seed from the transgenic plant of claim 4.

9. A method of modulating the level of signal transduction gene expression in a plant cell, comprising:

- 10 (a) introducing into a plant cell a recombinant expression cassette comprising a polynucleotide of claim 1 operably linked to a promoter;
- (b) culturing the plant cell under plant cell growing conditions; and
- (c) inducing expression of said polynucleotide for a time sufficient to modulate the level of signal transduction gene expression in said plant cell.

15 10. The method of claim 9, wherein the plant cell is from maize, wheat, rice, or soybean.

11. A method of modulating the level of signal transduction gene expression in a plant, comprising:

- 20 (a) introducing into a plant cell a recombinant expression cassette comprising a polynucleotide of claim 1 operably linked to a promoter;
- (b) culturing the plant cell under plant cell growing conditions;
- (c) regenerating a plant from said plant cell; and
- 25 (d) inducing expression of said polynucleotide for a time sufficient to modulate the level of signal transduction gene expression in said plant.

12. The method of claim 11, wherein the plant is maize, wheat, rice, or soybean.

13. An isolated protein comprising a member selected from the group consisting
30 of:

- (a) a polypeptide of at least 20 contiguous amino acids from a polypeptide of SEQ ID NO: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, 58, 62, or 66;
- (b) a polypeptide of SEQ ID NO: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, 58, 62, or 66;

- (c) a polypeptide having at least 80% sequence identity to, and having at least one epitope in common with, a polypeptide of SEQ ID NO: 2, 6, 10, 14, 18, 22, 26, 30, 34, 38, 42, 46, 50, 54, 58, 62, or 66, wherein said sequence identity is determined by the GAP algorithm under default parameters; and,
- 5 (d) at least one polypeptide encoded by a member of claim 1.

SEQUENCE LISTING

<110> Pioneer Hi-Bred International, Inc.

<120> Signal Transduction Genes and Methods of Use

<130> 1083-PCT

<150> US 60/134,292

<151> 1999-05-14

<150> US 60/142,996

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acctccttgt gattcacctg acaagtttgc aggtgggtcag gagcggtctc ctctctctag      180
tccactgaga agtccagttt taaaatcaag aaacccaagt gcacctccat cacca atg      238
                                         Met
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cat cca aag ttg ttc ccg gag aac cat gtt tct cgc cct gat ggc aat      286
His Pro Lys Leu Phe Pro Glu Asn His Val Ser Arg Pro Asp Gly Asn
      5              10              15

gga agt gct agt ttt cac cca tta ccg ctc cct cct gct tct gta agc      334
Gly Ser Ala Ser Phe His Pro Leu Pro Leu Pro Pro Ala Ser Val Ser
      20              25              30

cca aag cag aca aat gct agc cac cag tta gtt cca aaa gct gag atg      382
Pro Lys Gln Thr Asn Ala Ser His Gln Leu Val Pro Lys Ala Glu Met
      35              40              45

cct tcg gtg gct ggt caa tgg cag aaa gga aaa ctc ttg ggt agt ggc      430
Pro Ser Val Ala Gly Gln Trp Gln Lys Gly Lys Leu Leu Gly Ser Gly
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acc ttt gga tgc gtg tat gag gcc acc aat agg cac act gga gct ctt      478
Thr Phe Gly Cys Val Tyr Glu Ala Thr Asn Arg His Thr Gly Ala Leu
      70              75              80

tgt gcc atg aaa gag gtg aat ata att cca gat gat gct aaa tca gtc      526
Cys Ala Met Lys Glu Val Asn Ile Ile Pro Asp Asp Ala Lys Ser Val
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gag tct ctg aag caa ttg gaa cag gaa att aag ttt ctt agt caa ttt      574

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Glu Ser Leu Lys Gln Leu Glu Gln Glu Ile Lys Phe Leu Ser Gln Phe	
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Lys His Glu Asn Ile Val Gln Tyr Tyr Gly Ser Glu Thr Ile Glu Asp	
115 120 125	
aga ttc tac ata tac ctg gag tat gtt cat cct ggt tca atc cat aag	670
Arg Phe Tyr Ile Tyr Leu Glu Tyr Val His Pro Gly Ser Ile His Lys	
130 135 140 145	
tat gtt cat caa cac tgt gga tgc ctg aca gaa gca gtc atc cgt aac	718
Tyr Val His Gln His Cys Gly Ser Leu Thr Glu Ala Val Ile Arg Asn	
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ttc aca cgt cat att ctc aag ggt ctt gct ttt tta cat agc caa aag	766
Phe Thr Arg His Ile Leu Lys Gly Leu Ala Phe Leu His Ser Gln Lys	
165 170 175	
att atg cat agg gat atc aaa ggg gca aat ttg tta gtt gat att aac	814
Ile Met His Arg Asp Ile Lys Gly Ala Asn Leu Leu Val Asp Ile Asn	
180 185 190	
ggt gta gtg aaa ttg gct gac ttc gga atg gca aag cat ttg agt act	862
Gly Val Val Lys Leu Ala Asp Phe Gly Met Ala Lys His Leu Ser Thr	
195 200 205	
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Ala Ala Pro Asn Leu Ser Leu Lys Gly Thr Pro Tyr Trp Met Ala Pro	
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Glu Val Val Arg Ala Thr Leu Asp Lys Ser Ala Gly Tyr Asp Leu Ala	
230 235 240	
gtt gat att tgg agc ctt ggc tgc aca atc att gag atg ttt aca gga	1006
Val Asp Ile Trp Ser Leu Gly Cys Thr Ile Ile Glu Met Phe Thr Gly	
245 250 255	
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Lys Pro Pro Trp Ser Gly Leu Glu Gly Pro Ala Ala Met Phe Lys Val	
260 265 270	
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Leu Arg Thr Asp Pro Pro Ile Pro Asp Asn Leu Ser Pro Glu Gly Lys	
275 280 285	
gat ttt ctg aga tgc tgc ttc aag aga aac cct act gag cga cca aca	1150
Asp Phe Leu Arg Cys Cys Phe Lys Arg Asn Pro Thr Glu Arg Pro Thr	
290 295 300 305	
gca aac aag ttg ctc gaa cac cca ttt atc caa act ttg aac cac tac	1198
Ala Asn Lys Leu Glu His Pro Phe Ile Gln Thr Leu Asn His Tyr	
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Ser Pro His Ser Val Val His Ser Phe Ser Gly Ile Lys Ser Pro Asp	
325 330 335	
act gtg cac agc agt gca aga gat aaa gta ccc tgg aaa agt gat tca	1294
Thr Val His Ser Ser Ala Arg Asp Lys Val Pro Trp Lys Ser Asp Ser	
340 345 350	

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tca gag tca ttg gct tat cgg ttg atg aca ccc cta ccc aac ttg gga 1390
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acc aat agt ttg tca cct cca cct ttg agt tcg gcc tcc agt tct ggc 1438
 Thr Asn Ser Leu Ser Pro Pro Pro Leu Ser Ser Ala Ser Ser Ser Gly
 390 395 400

tct gca gca cat aca ccg aac agt atg cat ttt tca gtc cca tac cat 1486
 Ser Ala Ala His Thr Pro Asn Ser Met His Phe Ser Val Pro Tyr His
 405 410 415

cag cct agt cct ttg cca aag cct aac ggg aag gaa gca ata aat ttg 1534
 Gln Pro Ser Pro Leu Pro Lys Pro Asn Gly Lys Glu Ala Ile Asn Leu
 420 425 430

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 Val Ser His
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 35 40 45
 Met Pro Ser Val Ala Gly Gln Trp Gln Lys Gly Lys Leu Leu Gly Ser
 50 55 60
 Gly Thr Phe Gly Cys Val Tyr Glu Ala Thr Asn Arg His Thr Gly Ala
 65 70 75 80
 Leu Cys Ala Met Lys Glu Val Asn Ile Ile Pro Asp Asp Ala Lys Ser
 85 90 95
 Val Glu Ser Leu Lys Gln Leu Glu Gln Glu Ile Lys Phe Leu Ser Gln
 100 105 110
 Phe Lys His Glu Asn Ile Val Gln Tyr Tyr Gly Ser Glu Thr Ile Glu
 115 120 125
 Asp Arg Phe Tyr Ile Tyr Leu Glu Tyr Val His Pro Gly Ser Ile His
 130 135 140
 Lys Tyr Val His Gln His Cys Gly Ser Leu Thr Glu Ala Val Ile Arg
 145 150 155 160
 Asn Phe Thr Arg His Ile Leu Lys Gly Leu Ala Phe Leu His Ser Gln
 165 170 175

- 4 -

Lys Ile Met His Arg Asp Ile Lys Gly Ala Asn Leu Leu Val Asp Ile
 180 185 190
 Asn Gly Val Val Lys Leu Ala Asp Phe Gly Met Ala Lys His Leu Ser
 195 200 205
 Thr Ala Ala Pro Asn Leu Ser Leu Lys Gly Thr Pro Tyr Trp Met Ala
 210 215 220
 Pro Glu Val Val Arg Ala Thr Leu Asp Lys Ser Ala Gly Tyr Asp Leu
 225 230 235 240
 Ala Val Asp Ile Trp Ser Leu Gly Cys Thr Ile Ile Glu Met Phe Thr
 245 250 255
 Gly Lys Pro Pro Trp Ser Gly Leu Glu Gly Pro Ala Ala Met Phe Lys
 260 265 270
 Val Leu Arg Thr Asp Pro Pro Ile Pro Asp Asn Leu Ser Pro Glu Gly
 275 280 285
 Lys Asp Phe Leu Arg Cys Cys Phe Lys Arg Asn Pro Thr Glu Arg Pro
 290 295 300
 Thr Ala Asn Lys Leu Leu Glu His Pro Phe Ile Gln Thr Leu Asn His
 305 310 315 320
 Tyr Ser Pro His Ser Val Val His Ser Phe Ser Gly Ile Lys Ser Pro
 325 330 335
 Asp Thr Val His Ser Ser Ala Arg Asp Lys Val Pro Trp Lys Ser Asp
 340 345 350
 Ser Tyr Met Arg Gly Lys His Ala Asn Gly Glu Thr Ser Ser Ala Arg
 355 360 365
 Ser Ser Glu Ser Leu Ala Tyr Arg Leu Met Thr Pro Leu Pro Asn Leu
 370 375 380
 Gly Thr Asn Ser Leu Ser Pro Pro Pro Leu Ser Ser Ala Ser Ser Ser
 385 390 395 400
 Gly Ser Ala Ala His Thr Pro Asn Ser Met His Phe Ser Val Pro Tyr
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 His Gln Pro Ser Pro Leu Pro Lys Pro Asn Gly Lys Glu Ala Ile Asn
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 tgtcatgtgt gccgtgggaa ttgctccatt cgattggtaa gtgctcgtct ctgaccatgc 180
 agcctctcat ccggattgga ttggttccgg ctgttgcttg ctccaggaag aacactggac 240
 atg gac ttc ttc acg gaa tat ggg gag gcg agt cag tat caa atc caa 288
 Met Asp Phe Phe Thr Glu Tyr Gly Glu Ala Ser Gln Tyr Gln Ile Gln
 1 5 10 15

gag gtc att ggt aag gga agt tat gga gta gtt gct gct gca att gat 336
 Glu Val Ile Gly Lys Gly Ser Tyr Gly Val Val Ala Ala Ala Ile Asp
 20 25 30

agt cac act ggg gag cgg gtt gcg atc aag aag ata aat gat att ttt 384
 Ser His Thr Gly Glu Arg Val Ala Ile Lys Lys Ile Asn Asp Ile Phe
 35 40 45

ggt aat gtc tcg gat gct gct cgc att ttg cgg gaa atc aag ctt ctt 432
 Gly Asn Val Ser Asp Ala Ala Arg Ile Leu Arg Glu Ile Lys Leu Leu
 50 55 60

cgt ctg ctt cgt cac cca aac ata gtc cag att aag cac att atg ttg 480
 Arg Leu Leu Arg His Pro Asn Ile Val Gln Ile Lys His Ile Met Leu
 65 70 75 80

ccc cct acc cga agg gag ttc aaa gat atc tat gtt gtt ttt gag ctg 528
 Pro Pro Thr Arg Arg Glu Phe Lys Asp Ile Tyr Val Val Phe Glu Leu
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 Met Glu Ser Asp Leu His Gln Val Ile Lys Ala Asn Asp Asn Leu Thr
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cca gag cat cac cgc ttt ttc ctg tat caa ctt att cgt gct ctg aag 624
 Pro Glu His His Arg Phe Phe Leu Tyr Gln Leu Ile Arg Ala Leu Lys
 115 120 125

tac atg cat tca gcc cat gta ttt cat cgt gat tta aag ccc agg aac 672
 Tyr Met His Ser Ala His Val Phe His Arg Asp Leu Lys Pro Arg Asn
 130 135 140

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 Ile Leu Ala Asn Ser Asp Ser Lys Leu Lys Ile Cys Asp Phe Gly Leu
 145 150 155 160

gca cgt gca tca ttt aac gac tct ctt tca gct atc tat tgg aca gat 768
 Ala Arg Ala Ser Phe Asn Asp Ser Leu Ser Ala Ile Tyr Trp Thr Asp
 165 170 175

tat gtc gca aca agg tgg tac cgg gct cct gaa tta tgt ggc tcc ttt 816
 Tyr Val Ala Thr Arg Trp Tyr Arg Ala Pro Glu Leu Cys Gly Ser Phe
 180 185 190

ttc tcc agt tac acc cct gca att gat att tgg agt ata ggg tgc att 864
 Phe Ser Ser Tyr Thr Pro Ala Ile Asp Ile Trp Ser Ile Gly Cys Ile
 195 200 205

ttt gct gaa gtt ctg acc gga acg cca ttg ttt cct ggg agg aat gtt 912
 Phe Ala Glu Val Leu Thr Gly Thr Pro Leu Phe Pro Gly Arg Asn Val
 210 215 220

gtg cac caa tta gac tta ata aca gat ctg ctt gga aca cca tcg ttt 960
 Val His Gln Leu Asp Leu Ile Thr Asp Leu Leu Gly Thr Pro Ser Phe
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Arg Ser Leu Ser Gln Ile His Ser Asp Lys Ala Arg Glu Tyr Leu Leu	
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Gly Met Pro Arg Lys Arg Pro Ile Pro Phe Ser His Lys Phe Arg Asn	
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gcc gat cct ctg gct ctc cgg ttg cta gag cgt tta ctt gca ttt gac	1104
Ala Asp Pro Leu Ala Leu Arg Leu Leu Glu Arg Leu Leu Ala Phe Asp	
275 280 285	
ccg aaa gat cga cct act gct gaa gag gct tta gct gat cca tac ttc	1152
Pro Lys Asp Arg Pro Thr Ala Glu Glu Ala Leu Ala Asp Pro Tyr Phe	
290 295 300	
agt ggc ctt tct aaa ttg gaa ctt gag cct tca gca caa cca att ttg	1200
Ser Gly Leu Ser Lys Leu Glu Leu Glu Pro Ser Ala Gln Pro Ile Leu	
305 310 315 320	
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Lys Val Asp Phe Glu Phe Glu Gly Arg Lys Leu Thr Lys Ala Gly Val	
325 330 335	
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Arg Glu Met Ile Tyr Arg Glu Ile Leu Glu Tyr His Pro Gln Met Leu	
340 345 350	
cag gag tac atc gaa ggt gga gat cag att cac ttc cta tac cca agt	1344
Gln Glu Tyr Ile Glu Gly Gly Asp Gln Ile His Phe Leu Tyr Pro Ser	
355 360 365	
ggg gtt gat cgc ttc cag cga cag ttt gct cat ctc gag gag aac tac	1392
Gly Val Asp Arg Phe Gln Arg Gln Phe Ala His Leu Glu Glu Asn Tyr	
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Arg Arg Gly Val Ala Ser Thr Pro Pro Arg Arg Gln Pro Thr Ser Leu	
385 390 395 400	
ccg agg gag cga gtc tgc tca tca gaa gat ggc cac agt cag gac tct	1488
Pro Arg Glu Arg Val Cys Ser Ser Glu Asp Gly His Ser Gln Asp Ser	
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Asp Ser Glu Glu Gln Arg Ala Ala Ser Tyr Val Ala Arg Thr Thr Ile	
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agc cct cca aga tca caa gag gag ggc ggc aag ctc cag cct gcc tac	1584
Ser Pro Pro Arg Ser Gln Glu Glu Gly Gly Lys Leu Gln Pro Ala Tyr	
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cag agc gcc gcc gcc gcc gca gac tcc tgc gcc aag agc tat ctg aag	1632
Gln Ser Ala Ala Ala Ala Asp Ser Cys Ala Lys Ser Tyr Leu Lys	
450 455 460	
ggt gcc gct agt atc agt agt gct tcc agg cgt acc atc aag ggg gat	1680
Gly Ala Ala Ser Ile Ser Ser Ala Ser Arg Arg Thr Ile Lys Gly Asp	
465 470 475 480	
aat ggc cga aag gag aag gga tct ccg agg ctg tgaaagaggt ggcgcaggct	1733

Asn Gly Arg Lys Glu Lys Gly Ser Pro Arg Leu
485 490

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tggaggaatt agatttgtgt actgttaagt tttttttttt ggaattcaac tgctagattc 1973
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35 40 45
Gly Asn Val Ser Asp Ala Ala Arg Ile Leu Arg Glu Ile Lys Leu Leu
50 55 60
Arg Leu Leu Arg His Pro Asn Ile Val Gln Ile Lys His Ile Met Leu
65 70 75 80
Pro Pro Thr Arg Arg Glu Phe Lys Asp Ile Tyr Val Val Phe Glu Leu
85 90 95
Met Glu Ser Asp Leu His Gln Val Ile Lys Ala Asn Asp Asn Leu Thr
100 105 110
Pro Glu His His Arg Phe Phe Leu Tyr Gln Leu Ile Arg Ala Leu Lys
115 120 125
Tyr Met His Ser Ala His Val Phe His Arg Asp Leu Lys Pro Arg Asn
130 135 140
Ile Leu Ala Asn Ser Asp Ser Lys Leu Lys Ile Cys Asp Phe Gly Leu
145 150 155 160
Ala Arg Ala Ser Phe Asn Asp Ser Leu Ser Ala Ile Tyr Trp Thr Asp
165 170 175
Tyr Val Ala Thr Arg Trp Tyr Arg Ala Pro Glu Leu Cys Gly Ser Phe
180 185 190
Phe Ser Ser Tyr Thr Pro Ala Ile Asp Ile Trp Ser Ile Gly Cys Ile
195 200 205
Phe Ala Glu Val Leu Thr Gly Thr Pro Leu Phe Pro Gly Arg Asn Val
210 215 220
Val His Gln Leu Asp Leu Ile Thr Asp Leu Leu Gly Thr Pro Ser Phe
225 230 235 240
Arg Ser Leu Ser Gln Ile His Ser Asp Lys Ala Arg Glu Tyr Leu Leu
245 250 255
Gly Met Pro Arg Lys Arg Pro Ile Pro Phe Ser His Lys Phe Arg Asn
260 265 270
Ala Asp Pro Leu Ala Leu Arg Leu Leu Glu Arg Leu Leu Ala Phe Asp
275 280 285
Pro Lys Asp Arg Pro Thr Ala Glu Glu Ala Leu Ala Asp Pro Tyr Phe
290 295 300
Ser Gly Leu Ser Lys Leu Glu Leu Glu Pro Ser Ala Gln Pro Ile Leu
305 310 315 320
Lys Val Asp Phe Glu Phe Glu Gly Arg Lys Leu Thr Lys Ala Gly Val
325 330 335
Arg Glu Met Ile Tyr Arg Glu Ile Leu Glu Tyr His Pro Gln Met Leu
340 345 350
Gln Glu Tyr Ile Glu Gly Gly Asp Gln Ile His Phe Leu Tyr Pro Ser
355 360 365

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Gly Val Asp Arg Phe Gln Arg Gln Phe Ala His Leu Glu Glu Asn Tyr
 370 375 380
 Arg Arg Gly Val Ala Ser Thr Pro Pro Arg Arg Gln Pro Thr Ser Leu
 385 390 395 400
 Pro Arg Glu Arg Val Cys Ser Ser Glu Asp Gly His Ser Gln Asp Ser
 405 410 415
 Asp Ser Glu Glu Gln Arg Ala Ala Ser Tyr Val Ala Arg Thr Thr Ile
 420 425 430
 Ser Pro Pro Arg Ser Gln Glu Glu Gly Gly Lys Leu Gln Pro Ala Tyr
 435 440 445
 Gln Ser Ala Ala Ala Ala Asp Ser Cys Ala Lys Ser Tyr Leu Lys
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 Gly Ala Ala Ser Ile Ser Ser Ala Ser Arg Arg Thr Ile Lys Gly Asp
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 Asn Gly Arg Lys Glu Lys Gly Ser Pro Arg Leu
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48

gtg agg cag cgt act tgg ggg aag tgg gtt gct gaa ata aga gag cca
 Val Arg Gln Arg Thr Trp Gly Lys Trp Val Ala Glu Ile Arg Glu Pro
 20 25 30

96

aat cgt gtc gac aga ctc tgg ctg ggt acc ttc cca acc gcg gag gat
 Asn Arg Val Asp Arg Leu Trp Leu Gly Thr Phe Pro Thr Ala Glu Asp
 35 40 45

144

gca gct agg gcc tat gat gag gca gcc aga gcg atg tat gga gac ttg
 Ala Ala Arg Ala Tyr Asp Glu Ala Ala Arg Ala Met Tyr Gly Asp Leu
 50 55 60

192

gca cgg act aac ttc ccc gga cag gat gca aca acc tct gcc caa gct
 Ala Arg Thr Asn Phe Pro Gly Gln Asp Ala Thr Thr Ser Ala Gln Ala
 65 70 75 80

240

gct cta gca tcg acc tct gcc cag gct gat cca aca gct gtc gaa gct 288
 Ala Leu Ala Ser Thr Ser Ala Gln Ala Asp Pro Thr Ala Val Glu Ala
 85 90 95

ctt cag act ggc acg tca tgc gag tcg aca acg aca tca aat tac tcg 336
 Leu Gln Thr Gly Thr Ser Cys Glu Ser Thr Thr Ser Asn Tyr Ser
 100 105 110

gac atc gca tcc acc tca cac aag cct gag cct gaa gcc tct gac atc 384
 Asp Ile Ala Ser Thr Ser His Lys Pro Glu Pro Glu Ala Ser Asp Ile
 115 120 125

tcg agc tcc cta aag gca aaa tgt cca gct gga tca tgt ggt atc caa 432
 Ser Ser Ser Leu Lys Ala Lys Cys Pro Ala Gly Ser Cys Gly Ile Gln
 130 135 140

gac ggt aca ccc agt gta gct gac aag gag gtc ttt ggg ccg ttg gag 480
 Asp Gly Thr Pro Ser Val Ala Asp Lys Glu Val Phe Gly Pro Leu Glu
 145 150 155 160

cct atc aca aat ctt cca gat ggt ggt gat ggt ttt gat atc ggt gag 528
 Pro Ile Thr Asn Leu Pro Asp Gly Gly Asp Gly Phe Asp Ile Gly Glu
 165 170 175

atg ctg agg atg atg gaa agc gat cca cat aat gca ggt gga gct gac 576
 Met Leu Arg Met Met Glu Ser Asp Pro His Asn Ala Gly Gly Ala Asp
 180 185 190

gct ggc atg ggg cag ccc tgg tat ctt gat gag ctg gat tcg agt gtc 624
 Ala Gly Met Gly Gln Pro Trp Tyr Leu Asp Glu Leu Asp Ser Ser Val
 195 200 205

ttg gag agc atg ctc cag cca gag cca gag cca gag cca gag cca ttc 672
 Leu Glu Ser Met Leu Gln Pro Glu Pro Glu Pro Glu Pro Glu Pro Phe
 210 215 220

ctg atg tct gaa gaa ccg gac atg ttt ctt gct ggc ttc gaa agc gct 720
 Leu Met Ser Glu Glu Pro Asp Met Phe Leu Ala Gly Phe Glu Ser Ala
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<212> PRT

<213> Zea mays

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Asn Arg Val Asp Arg Leu Trp Leu Gly Thr Phe Pro Thr Ala Glu Asp
 35 40 45

Ala Ala Arg Ala Tyr Asp Glu Ala Ala Arg Ala Met Tyr Gly Asp Leu
 50 55 60

Ala Arg Thr Asn Phe Pro Gly Gln Asp Ala Thr Thr Ser Ala Gln Ala
 65 70 75 80

- 11 -

Ser	Val	Asn	Arg	Glu	Thr	Asn	Glu	Lys	Val	Ala	Ile	Lys	Lys	Ile	Asn		
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Asn	Val	Phe	Asp	Asn	Arg	Val	Asp	Ala	Leu	Arg	Thr	Leu	Arg	Glu	Leu		
65					70				75						80		
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Lys	Leu	Leu	Arg	His	Leu	Arg	His	Glu	Asn	Val	Ile	Ala	Leu	Lys	Asp		
				85					90					95			
ata	atg	atg	cct	gcg	cat	aga	agg	agc	ttc	aag	gac	gtt	tac	ttg	gtt	336	
Ile	Met	Met	Pro	Ala	His	Arg	Arg	Ser	Phe	Lys	Asp	Val	Tyr	Leu	Val		
			100					105					110				
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Tyr	Glu	Leu	Met	Asp	Thr	Asp	Leu	His	Gln	Ile	Ile	Lys	Ser	Ser	Gln		
		115					120					125					
cca	cta	tcc	aat	gac	cac	tgc	cag	tat	ttc	ctt	ttt	cag	ctg	ctc	cga	432	
Pro	Leu	Ser	Asn	Asp	His	Cys	Gln	Tyr	Phe	Leu	Phe	Gln	Leu	Leu	Arg		
		130				135					140						
ggc	ctg	aag	tac	ctc	cat	tca	gcc	ggg	ata	ctc	cac	aga	gac	cta	aag	480	
Gly	Leu	Lys	Tyr	Leu	His	Ser	Ala	Gly	Ile	Leu	His	Arg	Asp	Leu	Lys		
145					150					155					160		
cca	ggg	aac	ctc	ctg	gtc	aac	gca	aac	tgt	gac	ctg	aag	ata	tgc	gac	528	
Pro	Gly	Asn	Leu	Leu	Val	Asn	Ala	Asn	Cys	Asp	Leu	Lys	Ile	Cys	Asp		
			165						170					175			
ttc	ggg	ctc	gcc	cgc	acg	aac	aac	acc	aag	ggc	cag	ttc	atg	acg	gag	576	
Phe	Gly	Leu	Ala	Arg	Thr	Asn	Asn	Thr	Lys	Gly	Gln	Phe	Met	Thr	Glu		
			180					185					190				
tac	gtg	gtg	acc	cgc	tgg	tac	agg	gca	ccc	gag	ctg	ctg	ctc	tgc	tgc	624	
Tyr	Val	Val	Thr	Arg	Trp	Tyr	Arg	Ala	Pro	Glu	Leu	Leu	Leu	Cys	Cys		
		195					200					205					
gac	aac	tac	ggc	acg	tcc	att	gac	gtc	tgg	tct	gtg	ggg	tgc	ata	ttc	672	
Asp	Asn	Tyr	Gly	Thr	Ser	Ile	Asp	Val	Trp	Ser	Val	Gly	Cys	Ile	Phe		
		210				215					220						
gcg	gag	ctg	ctt	ggc	cgc	aag	ccg	atc	ttc	cca	gga	acc	gag	tgc	ctg	720	
Ala	Glu	Leu	Leu	Gly	Arg	Lys	Pro	Ile	Phe	Pro	Gly	Thr	Glu	Cys	Leu		
225					230					235					240		
aac	cag	ctc	aag	ctc	atc	gtc	aac	gtc	ctc	ggc	acc	atg	ggc	gag	gcc	768	
Asn	Gln	Leu	Lys	Leu	Ile	Val	Asn	Val	Leu	Gly	Thr	Met	Gly	Glu	Ala		
				245					250					255			
gac	ctc	gcg	ttc	atc	gac	aac	ccg	aag	gcc	cgc	aag	tac	atc	aag	tcc	816	
Asp	Leu	Ala	Phe	Ile	Asp	Asn	Pro	Lys	Ala	Arg	Lys	Tyr	Ile	Lys	Ser		
			260					265					270				
ctt	ccg	tac	gcc	ccg	ggc	gcc	ccc	ttc	acc	ggc	atg	tac	cct	cag	gcg	864	
Leu	Pro	Tyr	Ala	Pro	Gly	Ala	Pro	Phe	Thr	Gly	Met	Tyr	Pro	Gln	Ala		
		275					280					285					
cac	cct	ctc	gcc	atc	gac	ctg	ctg	cag	aag	atg	ctc	gtg	ttc	gac	ccg	912	
His	Pro	Leu	Ala	Ile	Asp	Leu	Leu	Gln	Lys	Met	Leu	Val	Phe	Asp	Pro		
		290				295					300						

tcc aag agg atc agc gtc acc gag gcg ctg gag cac ccg tac atg tct 960
 Ser Lys Arg Ile Ser Val Thr Glu Ala Leu Glu His Pro Tyr Met Ser
 305 310 315 320

ccg ctc tat gac ccg agc gcg aac cct ccc gcg cag gtg ccg atc gac 1008
 Pro Leu Tyr Asp Pro Ser Ala Asn Pro Pro Ala Gln Val Pro Ile Asp
 325 330 335

ctc gac atc gac gag aac ctc ggc gtc gac atg atc agg gag atg atg 1056
 Leu Asp Ile Asp Glu Asn Leu Gly Val Asp Met Ile Arg Glu Met Met
 340 345 350

tgg cag gag atg atc cac tac cac ccc gag gtc ctc acg aga atc agc 1104
 Trp Gln Glu Met Ile His Tyr His Pro Glu Val Leu Thr Arg Ile Ser
 355 360 365

atg tga 1110
 Met

<210> 14
 <211> 369
 <212> PRT
 <213> Zea mays

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 Val Pro Ile Lys Pro Ile Gly Arg Gly Ala Tyr Gly Val Val Cys Ser
 35 40 45
 Ser Val Asn Arg Glu Thr Asn Glu Lys Val Ala Ile Lys Lys Ile Asn
 50 55 60
 Asn Val Phe Asp Asn Arg Val Asp Ala Leu Arg Thr Leu Arg Glu Leu
 65 70 75 80
 Lys Leu Leu Arg His Leu Arg His Glu Asn Val Ile Ala Leu Lys Asp
 85 90 95
 Ile Met Met Pro Ala His Arg Arg Ser Phe Lys Asp Val Tyr Leu Val
 100 105 110
 Tyr Glu Leu Met Asp Thr Asp Leu His Gln Ile Ile Lys Ser Ser Gln
 115 120 125
 Pro Leu Ser Asn Asp His Cys Gln Tyr Phe Leu Phe Gln Leu Leu Arg
 130 135 140
 Gly Leu Lys Tyr Leu His Ser Ala Gly Ile Leu His Arg Asp Leu Lys
 145 150 155 160
 Pro Gly Asn Leu Leu Val Asn Ala Asn Cys Asp Leu Lys Ile Cys Asp
 165 170 175
 Phe Gly Leu Ala Arg Thr Asn Asn Thr Lys Gly Gln Phe Met Thr Glu
 180 185 190
 Tyr Val Val Thr Arg Trp Tyr Arg Ala Pro Glu Leu Leu Cys Cys
 195 200 205
 Asp Asn Tyr Gly Thr Ser Ile Asp Val Trp Ser Val Gly Cys Ile Phe
 210 215 220
 Ala Glu Leu Leu Gly Arg Lys Pro Ile Phe Pro Gly Thr Glu Cys Leu
 225 230 235 240
 Asn Gln Leu Lys Leu Ile Val Asn Val Leu Gly Thr Met Gly Glu Ala
 245 250 255
 Asp Leu Ala Phe Ile Asp Asn Pro Lys Ala Arg Lys Tyr Ile Lys Ser
 260 265 270

Leu Pro Tyr Ala Pro Gly Ala Pro Phe Thr Gly Met Tyr Pro Gln Ala
 275 280 285
 His Pro Leu Ala Ile Asp Leu Leu Gln Lys Met Leu Val Phe Asp Pro
 290 295 300
 Ser Lys Arg Ile Ser Val Thr Glu Ala Leu Glu His Pro Tyr Met Ser
 305 310 315 320
 Pro Leu Tyr Asp Pro Ser Ala Asn Pro Pro Ala Gln Val Pro Ile Asp
 325 330 335
 Leu Asp Ile Asp Glu Asn Leu Gly Val Asp Met Ile Arg Glu Met Met
 340 345 350
 Trp Gln Glu Met Ile His Tyr His Pro Glu Val Leu Thr Arg Ile Ser
 355 360 365
 Met

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 <212> DNA
 <213> Zea mays

<400> 15
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<210> 16
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<400> 16
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<210> 17
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 acg cgc ggc ggc cgc tac gtg ctg tac aac gtg tac gga aac ctc ttc 96
 Thr His Gly Gly Arg Tyr Val Leu Tyr Asn Val Tyr Gly Asn Leu Phe
 20 25 30
 gag gtc tcc tcc aag tac gcc cca ccc atc cgc ccc atc ggt cgc ggc 144
 Glu Val Ser Ser Lys Tyr Ala Pro Pro Ile Arg Pro Ile Gly Arg Gly
 35 40 45
 gcc tac ggc att gtc tgc gcg gct gtc aac tcg cag tca ggg gag gag 192
 Ala Tyr Gly Ile Val Cys Ala Ala Val Asn Ser Gln Ser Gly Glu Glu
 50 55 60
 gtt gcg atc aag aag gtt ggc aat gcg ttc gac aac cac atc gac gcc 240
 Val Ala Ile Lys Lys Val Gly Asn Ala Phe Asp Asn His Ile Asp Ala
 65 70 75 80
 aag cgg acg ctc agg gaa atc aag ctg ctg cgc cac atg gac cat gag 288

Lys Arg Thr Leu Arg Glu Ile Lys Leu Leu Arg His Met Asp His Glu	
85 90 95	
aac atc ctt gcc tta aag gat gta att cgg ccc cca act aga gag aac	336
Asn Ile Leu Ala Leu Lys Asp Val Ile Arg Pro Pro Thr Arg Glu Asn	
100 105 110	
ttt aat gac gtg tac att gtt act gag tta atg gat aca gat ctc cat	384
Phe Asn Asp Val Tyr Ile Val Thr Glu Leu Met Asp Thr Asp Leu His	
115 120 125	
cag atc gta cgc tca aat cag cca ttg act gat gat cat tgc cag tac	432
Gln Ile Val Arg Ser Asn Gln Pro Leu Thr Asp Asp His Cys Gln Tyr	
130 135 140	
ttc ttg tat cag ttg tta cga ggg cta aaa tat gtg cac tca gca aat	480
Phe Leu Tyr Gln Leu Leu Arg Gly Leu Lys Tyr Val His Ser Ala Asn	
145 150 155 160	
ata ttg cac cgc gat ctg aag ccg agc aat ttg ttc cta aat gca aat	528
Ile Leu His Arg Asp Leu Lys Pro Ser Asn Leu Phe Leu Asn Ala Asn	
165 170 175	
tgt gac ctc aag att gca gac ttt ggg ctt gca agg acc act tca gag	576
Cys Asp Leu Lys Ile Ala Asp Phe Gly Leu Ala Arg Thr Thr Ser Glu	
180 185 190	
aca gat ctc atg aca gag tat gtg gtc act cgt tgg tac cgg gca cca	624
Thr Asp Leu Met Thr Glu Tyr Val Val Thr Arg Trp Tyr Arg Ala Pro	
195 200 205	
gag ctg ctg ttg aac tgt tca cag tat act gct gcc att gat gtc tgg	672
Glu Leu Leu Leu Asn Cys Ser Gln Tyr Thr Ala Ala Ile Asp Val Trp	
210 215 220	
tca gtt gga tgc ata cta ggt gaa atc gtt act cgt caa ccc ctg ttt	720
Ser Val Gly Cys Ile Leu Gly Glu Ile Val Thr Arg Gln Pro Leu Phe	
225 230 235 240	
cct gga cgg gat tac atc cag caa tta aaa ttg atc act gag ctc ata	768
Pro Gly Arg Asp Tyr Ile Gln Gln Leu Lys Leu Ile Thr Glu Leu Ile	
245 250 255	
ggc tct cca gat gat gca agc ctg gga ttt ctt cga agt gat aat gca	816
Gly Ser Pro Asp Asp Ala Ser Leu Gly Phe Leu Arg Ser Asp Asn Ala	
260 265 270	
aaa aga tac atg aaa caa cta cca cag ttt cca aga cag gac ttc cgc	864
Lys Arg Tyr Met Lys Gln Leu Pro Gln Phe Pro Arg Gln Asp Phe Arg	
275 280 285	
ctg cgt ttc cgc aac atg tct cct ggc gca gtc gat ttg ttg gaa agg	912
Leu Arg Phe Arg Asn Met Ser Pro Gly Ala Val Asp Leu Leu Glu Arg	
290 295 300	
atg ctt gtg ttt gat cca agc aga cgg att aca gtt gat gag gct ctg	960
Met Leu Val Phe Asp Pro Ser Arg Arg Ile Thr Val Asp Glu Ala Leu	
305 310 315 320	
cat cat cca tac ttg gct tca ctt cat gag atc aat gat gaa cct acc	1008
His His Pro Tyr Leu Ala Ser Leu His Glu Ile Asn Asp Glu Pro Thr	
325 330 335	

tgc cct gca cct ttc agc ttt gat ttt gag caa cca tcc ttt aca gaa 1056
 Cys Pro Ala Pro Phe Ser Phe Asp Phe Glu Gln Pro Ser Phe Thr Glu
 340 345 350
 gcg cat ata aaa gaa ctc atc tgg agg gaa tct tta gca ttt aac cca 1104
 Ala His Ile Lys Glu Leu Ile Trp Arg Glu Ser Leu Ala Phe Asn Pro
 355 360 365
 gag cct ccc tac taa 1119
 Glu Pro Pro Tyr
 370

<210> 18
 <211> 372
 <212> PRT
 <213> Zea mays

<400> 18
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 Thr His Gly Gly Arg Tyr Val Leu Tyr Asn Val Tyr Gly Asn Leu Phe
 20 25 30
 Glu Val Ser Ser Lys Tyr Ala Pro Pro Ile Arg Pro Ile Gly Arg Gly
 35 40 45
 Ala Tyr Gly Ile Val Cys Ala Ala Val Asn Ser Gln Ser Gly Glu Glu
 50 55 60
 Val Ala Ile Lys Lys Val Gly Asn Ala Phe Asp Asn His Ile Asp Ala
 65 70 75 80
 Lys Arg Thr Leu Arg Glu Ile Lys Leu Leu Arg His Met Asp His Glu
 85 90 95
 Asn Ile Leu Ala Leu Lys Asp Val Ile Arg Pro Pro Thr Arg Glu Asn
 100 105 110
 Phe Asn Asp Val Tyr Ile Val Thr Glu Leu Met Asp Thr Asp Leu His
 115 120 125
 Gln Ile Val Arg Ser Asn Gln Pro Leu Thr Asp Asp His Cys Gln Tyr
 130 135 140
 Phe Leu Tyr Gln Leu Leu Arg Gly Leu Lys Tyr Val His Ser Ala Asn
 145 150 155 160
 Ile Leu His Arg Asp Leu Lys Pro Ser Asn Leu Phe Leu Asn Ala Asn
 165 170 175
 Cys Asp Leu Lys Ile Ala Asp Phe Gly Leu Ala Arg Thr Thr Ser Glu
 180 185 190
 Thr Asp Leu Met Thr Glu Tyr Val Val Thr Arg Trp Tyr Arg Ala Pro
 195 200 205
 Glu Leu Leu Leu Asn Cys Ser Gln Tyr Thr Ala Ala Ile Asp Val Trp
 210 215 220
 Ser Val Gly Cys Ile Leu Gly Glu Ile Val Thr Arg Gln Pro Leu Phe
 225 230 235 240
 Pro Gly Arg Asp Tyr Ile Gln Gln Leu Lys Leu Ile Thr Glu Leu Ile
 245 250 255
 Gly Ser Pro Asp Asp Ala Ser Leu Gly Phe Leu Arg Ser Asp Asn Ala
 260 265 270
 Lys Arg Tyr Met Lys Gln Leu Pro Gln Phe Pro Arg Gln Asp Phe Arg
 275 280 285
 Leu Arg Phe Arg Asn Met Ser Pro Gly Ala Val Asp Leu Leu Glu Arg
 290 295 300
 Met Leu Val Phe Asp Pro Ser Arg Arg Ile Thr Val Asp Glu Ala Leu
 305 310 315 320
 His His Pro Tyr Leu Ala Ser Leu His Glu Ile Asn Asp Glu Pro Thr
 325 330 335

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<211> 20
<212> DNA
<213> Zea mays
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<400> 19
atggattcct ccggcggcgg 20

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<211> 20
<212> DNA
<213> Zea mays
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<400> 20	
ttagtaggga ggctctgggt	20

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<210> 21
<211> 1113
<212> DNA
<213> Zea mays
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 Met Ala Met Met Val Asp Pro Pro Asn Gly Ile Gly Asn Gln Gly Lys
 1 5 10 15

cat tac tac tct atg tgg cag acc tta ttt gag ata gac acc aaa tat 96
His Tyr Tyr Ser Met Trp Gln Thr Leu Phe Glu Ile Asp Thr Lys Tyr
20 25 30

gtg ccg atc aag ccc att ggt cga gga gct tat ggg ata gtt tgt tca 144
Val Pro Ile Lys Pro Ile Gly Arg Gly Ala Tyr Gly Ile Val Cys Ser
35 40 45

tcc att ant cgt gaa aca aat gag aaa gta gca ata aag aag ata cac 192
Ser Ile Asn Arg Glu Thr Asn Glu Lys Val Ala Ile Lys Lys Ile His
50 55 60

aac gtt ttc gac aac cgt gtg gat gca cta cgg acc ttg cgg gag ctg 240
Asn Val Phe Asp Asn Arg Val Asp Ala Leu Arg Thr Leu Arg Glu Leu
65 70 75 80

aaa ctc ctt cgc cat ctc cgg cat gag aat gtc att gct ttg aag gat 288
Lys Leu Leu Arg His Leu Arg His Glu Asn Val Ile Ala Leu Lys Asp
85 90 95

ata atg atg cca ata cac agg aga agc ttt aag gat gtg tac ttg gta 336
Ile Met Met Pro Ile His Arg Arg Ser Phe Lys Asp Val Tyr Leu Val
100 105 110

tac gaa ctc atg gat act gat ttg cac cag ata atc aaa tca cct cag 384

Tyr	Glu	Leu	Met	Asp	Thr	Asp	Leu	His	Gln	Ile	Ile	Lys	Ser	Pro	Gln		
		115					120					125					
ggc	ctt	tcc	aat	gac	cac	tgc	cag	tat	ttt	ctt	ttt	cag	ttg	ctc	cga	432	
Gly	Leu	Ser	Asn	Asp	His	Cys	Gln	Tyr	Phe	Leu	Phe	Gln	Leu	Leu	Arg		
		130				135					140						
gga	ctc	aaa	tat	ctc	cat	tca	gca	gaa	ata	ctc	cac	aga	gac	cta	aaa	480	
Gly	Leu	Lys	Tyr	Leu	His	Ser	Ala	Glu	Ile	Leu	His	Arg	Asp	Leu	Lys		
		145			150					155					160		
cct	gga	aac	ctg	ctg	gtg	aat	gca	aat	tgt	gat	ctg	aag	ata	tgt	gat	528	
Pro	Gly	Asn	Leu	Leu	Val	Asn	Ala	Asn	Cys	Asp	Leu	Lys	Ile	Cys	Asp		
			165					170						175			
ttt	ggt	ctc	gca	cgt	aca	aac	agt	agc	aaa	ggc	cag	ttc	atg	act	gaa	576	
Phe	Gly	Leu	Ala	Arg	Thr	Asn	Ser	Ser	Lys	Gly	Gln	Phe	Met	Thr	Glu		
			180					185					190				
tac	gtc	gtc	acc	cgc	tgg	tac	aga	gct	cct	gag	ctg	ctc	ctc	tgc	tgc	624	
Tyr	Val	Val	Thr	Arg	Trp	Tyr	Arg	Ala	Pro	Glu	Leu	Leu	Leu	Cys	Cys		
			195				200					205					
gac	aac	tac	ggc	aca	tcc	ata	gac	gtc	tgg	tct	gtt	ggg	tgc	atc	ttt	672	
Asp	Asn	Tyr	Gly	Thr	Ser	Ile	Asp	Val	Trp	Ser	Val	Gly	Cys	Ile	Phe		
		210				215					220						
gct	gag	ctc	ctt	ggc	cgc	aag	cca	ata	ttt	cca	gga	act	gaa	tgc	ctg	720	
Ala	Glu	Leu	Leu	Gly	Arg	Lys	Pro	Ile	Phe	Pro	Gly	Thr	Glu	Cys	Leu		
				230						235					240		
aat	caa	ctc	aag	ctc	ata	gtg	aac	gtc	ctc	ggc	aca	atg	agt	gag	gct	768	
Asn	Gln	Leu	Lys	Leu	Ile	Val	Asn	Val	Leu	Gly	Thr	Met	Ser	Glu	Ala		
				245					250					255			
gac	cta	gag	ttc	atc	gac	aac	cca	aag	gct	cgg	aga	tac	atc	aag	tcc	816	
Asp	Leu	Glu	Phe	Ile	Asp	Asn	Pro	Lys	Ala	Arg	Arg	Tyr	Ile	Lys	Ser		
			260					265					270				
ctt	ccc	tat	acc	cct	ggt	gtt	cct	ctc	gta	agt	atg	tac	cca	cat	gcg	864	
Leu	Pro	Tyr	Thr	Pro	Gly	Val	Pro	Leu	Val	Ser	Met	Tyr	Pro	His	Ala		
		275					280					285					
cac	cct	ctt	gcc	att	gat	ctg	ttg	cag	aag	atg	ctc	atc	ttc	gac	ccc	912	
His	Pro	Leu	Ala	Ile	Asp	Leu	Leu	Gln	Lys	Met	Leu	Ile	Phe	Asp	Pro		
		290				295					300						
acc	aaa	agg	atc	agt	gtc	acc	gag	gct	ctc	gag	cac	cct	tac	atg	tcc	960	
Thr	Lys	Arg	Ile	Ser	Val	Thr	Glu	Ala	Leu	Glu	His	Pro	Tyr	Met	Ser		
		305			310				315						320		
cct	ctg	tat	gac	cca	agc	gca	aat	ccc	cca	gcc	caa	gtg	ccc	atc	gat	1008	
Pro	Leu	Tyr	Asp	Pro	Ser	Ala	Asn	Pro	Pro	Ala	Gln	Val	Pro	Ile	Asp		
				325				330					335				
ctg	gac	ata	gac	gaa	aac	atc	agc	tca	gag	atg	atc	cgg	gaa	atg	atg	1056	
Leu	Asp	Ile	Asp	Glu	Asn	Ile	Ser	Ser	Glu	Met	Ile	Arg	Glu	Met	Met		
			340					345					350				
tgg	cag	gag	atg	ctt	cac	tac	cac	cct	gaa	gtt	gcc	aca	gca	ata	agc	1104	
Trp	Gln	Glu	Met	Leu	His	Tyr	His	Pro	Glu	Val	Ala	Thr	Ala	Ile	Ser		
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atg tca tga
Met Ser
370

1113

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<211> 370
<212> PRT
<213> Zea mays

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Val Pro Ile Lys Pro Ile Gly Arg Gly Ala Tyr Gly Ile Val Cys Ser
35 40 45
Ser Ile Asn Arg Glu Thr Asn Glu Lys Val Ala Ile Lys Lys Ile His
50 55 60
Asn Val Phe Asp Asn Arg Val Asp Ala Leu Arg Thr Leu Arg Glu Leu
65 70 75 80
Lys Leu Leu Arg His Leu Arg His Glu Asn Val Ile Ala Leu Lys Asp
85 90 95
Ile Met Met Pro Ile His Arg Arg Ser Phe Lys Asp Val Tyr Leu Val
100 105 110
Tyr Glu Leu Met Asp Thr Asp Leu His Gln Ile Ile Lys Ser Pro Gln
115 120 125
Gly Leu Ser Asn Asp His Cys Gln Tyr Phe Leu Phe Gln Leu Leu Arg
130 135 140
Gly Leu Lys Tyr Leu His Ser Ala Glu Ile Leu His Arg Asp Leu Lys
145 150 155 160
Pro Gly Asn Leu Leu Val Asn Ala Asn Cys Asp Leu Lys Ile Cys Asp
165 170 175
Phe Gly Leu Ala Arg Thr Asn Ser Ser Lys Gly Gln Phe Met Thr Glu
180 185 190
Tyr Val Val Thr Arg Trp Tyr Arg Ala Pro Glu Leu Leu Leu Cys Cys
195 200 205
Asp Asn Tyr Gly Thr Ser Ile Asp Val Trp Ser Val Gly Cys Ile Phe
210 215 220
Ala Glu Leu Leu Gly Arg Lys Pro Ile Phe Pro Gly Thr Glu Cys Leu
225 230 235 240
Asn Gln Leu Lys Leu Ile Val Asn Val Leu Gly Thr Met Ser Glu Ala
245 250 255
Asp Leu Glu Phe Ile Asp Asn Pro Lys Ala Arg Arg Tyr Ile Lys Ser
260 265 270
Leu Pro Tyr Thr Pro Gly Val Pro Leu Val Ser Met Tyr Pro His Ala
275 280 285
His Pro Leu Ala Ile Asp Leu Leu Gln Lys Met Leu Ile Phe Asp Pro
290 295 300
Thr Lys Arg Ile Ser Val Thr Glu Ala Leu Glu His Pro Tyr Met Ser
305 310 315 320
Pro Leu Tyr Asp Pro Ser Ala Asn Pro Pro Ala Gln Val Pro Ile Asp
325 330 335
Leu Asp Ile Asp Glu Asn Ile Ser Ser Glu Met Ile Arg Glu Met Met
340 345 350
Trp Gln Glu Met Leu His Tyr His Pro Glu Val Ala Thr Ala Ile Ser
355 360 365
Met Ser
370

<210> 23

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<211> 20
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<213> Zea mays

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atggcgatga tgggtggatcc 20

<210> 24
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<212> DNA
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<400> 24
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<210> 25
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<212> DNA
<213> Zea mays

<220>
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<222> (1)...(1122)

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Met Ser Gly Gly Gly Val Asp Gly Ala Pro Val Ala Glu Phe Arg Gln
1 5 10 15

acg gtg acg cac ggc ggc cgg ttc ctg cag tac aac atc ttc ggc aac 96
Thr Val Thr His Gly Gly Arg Phe Leu Gln Tyr Asn Ile Phe Gly Asn
20 25 30

ctg ttc gag atc acg cac aag tac cag ccc ccc atc atg ccc atc ggc 144
Leu Phe Glu Ile Thr His Lys Tyr Gln Pro Pro Ile Met Pro Ile Gly
35 40 45

cgc ggc gcc tac ggg atc gtc tgc tgc gtg atg aac tcc gag acg aag 192
Arg Gly Ala Tyr Gly Ile Val Cys Ser Val Met Asn Ser Glu Thr Lys
50 55 60

gag atg gtg gcc atc aag aag atc gcc aac gcc ttc gac aac cac atg 240
Glu Met Val Ala Ile Lys Lys Ile Ala Asn Ala Phe Asp Asn His Met
65 70 75 80

gac gcc aag cgc acg ctc cgg gag atc aag ctg ctg agg cac ctc gac 288
Asp Ala Lys Arg Thr Leu Arg Glu Ile Lys Leu Leu Arg His Leu Asp
85 90 95

cac gag aac atc atc ggc atc agg gac gtg atc ccg ccg ccc gtc ccg 336
His Glu Asn Ile Ile Gly Ile Arg Asp Val Ile Pro Pro Pro Val Pro
100 105 110

caa gcg ttc aac gac gtg tac atc ggg acg gag ctg atg gac acg gac 384
Gln Ala Phe Asn Asp Val Tyr Ile Gly Thr Glu Leu Met Asp Thr Asp
115 120 125

ctg cac cac atc atc cgg tcc aac cag gag ctc tcc gag gag cac tcc 432
Leu His His Ile Ile Arg Ser Asn Gln Glu Leu Ser Glu Glu His Ser
130 135 140

cag tac ttc atg tac cag atc ctc cgc ggg ctc aag tac atc cac tcc 480

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- 20 -

Gln Tyr Phe Met Tyr Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser
 145 150 155 160

gcc aac gtg atc cac cgc gac ctc aag ccc agc aac ctg ctg gtg aac 528
 Ala Asn Val Ile His Arg Asp Leu Lys Pro Ser Asn Leu Leu Val Asn
 165 170 175

gcc aac tgc gac ctc aag atc tgc gac ttc ggg ctg gcg cgc ccg tcc 576
 Ala Asn Cys Asp Leu Lys Ile Cys Asp Phe Gly Leu Ala Arg Pro Ser
 180 185 190

tcc gag agc gac atg atg acg gag tac gtg gtc acg cgc tgg tac cgc 624
 Ser Glu Ser Asp Met Met Thr Glu Tyr Val Val Thr Arg Trp Tyr Arg
 195 200 205

gcg ccc gag ctg ctg ctc aac tcc acc gac tac tcg gcg gcc atc gac 672
 Ala Pro Glu Leu Leu Leu Asn Ser Thr Asp Tyr Ser Ala Ala Ile Asp
 210 215 220

gtc tgg tcc gtc ggc tgc atc ttc atg gag ctc atc aac cgc cag ccg 720
 Val Trp Ser Val Gly Cys Ile Phe Met Glu Leu Ile Asn Arg Gln Pro
 225 230 235 240

ctc ttc ccc gga cgc gac cac atg cac cag atg cgc ctc ata acc gag 768
 Leu Phe Pro Gly Arg Asp His Met His Gln Met Arg Leu Ile Thr Glu
 245 250 255

gtg atc ggg acg ccc acg gac gat gag ctc ggg ttc atc cgg aac gag 816
 Val Ile Gly Thr Pro Thr Asp Asp Glu Leu Gly Phe Ile Arg Asn Glu
 260 265 270

gac gcg cgg aag tac atg cgc cac ctc ccg cag ttc ccg cgc cgg ccg 864
 Asp Ala Arg Lys Tyr Met Arg His Leu Pro Gln Phe Pro Arg Arg Pro
 275 280 285

ttc gtc agc ctg ttc ccg cgg atg cag ccc gtc gcg ctg gac ctc atc 912
 Phe Val Ser Leu Phe Pro Arg Met Gln Pro Val Ala Leu Asp Leu Ile
 290 295 300

gag cgg atg ctc acc ttc aac ccg ctg cag agg atc aca gtg gaa gag 960
 Glu Arg Met Leu Thr Phe Asn Pro Leu Gln Arg Ile Thr Val Glu Glu
 305 310 315 320

gcg ctg gag cac ccg tac ctg gaa cgg cta cac gac gtc gcc gac gag 1008
 Ala Leu Glu His Pro Tyr Leu Glu Arg Leu His Asp Val Ala Asp Glu
 325 330 335

ccc atc tgc acg gac ccg ttc tcg ttc gac ttc gag cag cag gct ctg 1056
 Pro Ile Cys Thr Asp Pro Phe Ser Phe Asp Phe Glu Gln Gln Ala Leu
 340 345 350

acg gaa gac caa atg aag cag ctg ata ttc aac gag gcc atg gaa ctc 1104
 Thr Glu Asp Gln Met Lys Gln Leu Ile Phe Asn Glu Ala Met Glu Leu
 355 360 365

aac ccc aac ttc cga tac tag 1125
 Asn Pro Asn Phe Arg Tyr
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<210> 26

<211> 374

- 21 -

<212> PPT

<213> Zea mays

<400> 26

Met Ser Gly Gly Gly Val Asp Gly Ala Pro Val Ala Glu Phe Arg Gln
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 20 25 30
 Leu Phe Glu Ile Thr His Lys Tyr Gln Pro Pro Ile Met Pro Ile Gly
 35 40 45
 Arg Gly Ala Tyr Gly Ile Val Cys Ser Val Met Asn Ser Glu Thr Lys
 50 55 60
 Glu Met Val Ala Ile Lys Lys Ile Ala Asn Ala Phe Asp Asn His Met
 65 70 75 80
 Asp Ala Lys Arg Thr Leu Arg Glu Ile Lys Leu Leu Arg His Leu Asp
 85 90 95
 His Glu Asn Ile Ile Gly Ile Arg Asp Val Ile Pro Pro Pro Val Pro
 100 105 110
 Gln Ala Phe Asn Asp Val Tyr Ile Gly Thr Glu Leu Met Asp Thr Asp
 115 120 125
 Leu His His Ile Ile Arg Ser Asn Gln Glu Leu Ser Glu Glu His Ser
 130 135 140
 Gln Tyr Phe Met Tyr Gln Ile Leu Arg Gly Leu Lys Tyr Ile His Ser
 145 150 155 160
 Ala Asn Val Ile His Arg Asp Leu Lys Pro Ser Asn Leu Leu Val Asn
 165 170 175
 Ala Asn Cys Asp Leu Lys Ile Cys Asp Phe Gly Leu Ala Arg Pro Ser
 180 185 190
 Ser Glu Ser Asp Met Met Thr Glu Tyr Val Val Thr Arg Trp Tyr Arg
 195 200 205
 Ala Pro Glu Leu Leu Leu Asn Ser Thr Asp Tyr Ser Ala Ala Ile Asp
 210 215 220
 Val Trp Ser Val Gly Cys Ile Phe Met Glu Leu Ile Asn Arg Gln Pro
 225 230 235 240
 Leu Phe Pro Gly Arg Asp His Met His Gln Met Arg Leu Ile Thr Glu
 245 250 255
 Val Ile Gly Thr Pro Thr Asp Asp Glu Leu Gly Phe Ile Arg Asn Glu
 260 265 270
 Asp Ala Arg Lys Tyr Met Arg His Leu Pro Gln Phe Pro Arg Arg Pro
 275 280 285
 Phe Val Ser Leu Phe Pro Arg Met Gln Pro Val Ala Leu Asp Leu Ile
 290 295 300
 Glu Arg Met Leu Thr Phe Asn Pro Leu Gln Arg Ile Thr Val Glu Glu
 305 310 315 320
 Ala Leu Glu His Pro Tyr Leu Glu Arg Leu His Asp Val Ala Asp Glu
 325 330 335
 Pro Ile Cys Thr Asp Pro Phe Ser Phe Asp Phe Glu Gln Gln Ala Leu
 340 345 350
 Thr Glu Asp Gln Met Lys Gln Leu Ile Phe Asn Glu Ala Met Glu Leu
 355 360 365
 Asn Pro Asn Phe Arg Tyr
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<210> 27

<211> 20

<212> DNA

<213> Zea mays

<400> 27

atgagcggag gaggcgtgga

20

<210> 28

<211> 20
 <212> DNA
 <213> Zea mays

<400> 28
 ctagtatcgg aagttggggt

20

<210> 29
 <211> 1197
 <212> DNA
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<220>
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 Met Asp Gly Gly Gly Gln Pro Pro Asp Thr Glu Met Ser Glu Ala Gly
 1 5 10 15

gcg ggc ggc gga ggg cag ccg ccg cag cag cca ctg ccg ccg gtg ggc 96
 Ala Gly Gly Gly Gly Gln Pro Pro Gln Gln Pro Leu Pro Pro Val Gly
 20 25 30

ggg ggc gtg atg ttg gac aac atc cag gcg acg ctc agc cac ggc ggc 144
 Gly Gly Val Met Leu Asp Asn Ile Gln Ala Thr Leu Ser His Gly Gly
 35 40 45

cgc ttc atc cag tac aac atc ttc ggc aac gtg ttc gag gtc acc gcc 192
 Arg Phe Ile Gln Tyr Asn Ile Phe Gly Asn Val Phe Glu Val Thr Ala
 50 55 60

aag tac aag ccc ccc gtc ctc ccc atc ggc aag ggc gcc tac ggc atc 240
 Lys Tyr Lys Pro Pro Val Leu Pro Ile Gly Lys Gly Ala Tyr Gly Ile
 65 70 75 80

gtc tgc tcg gcg ctc aac tcc gag acg gcg gag cag gtg gcc atc aag 288
 Val Cys Ser Ala Leu Asn Ser Glu Thr Ala Glu Gln Val Ala Ile Lys
 85 90 95

aag atc gcc aac gcc ttc gac aac aag atc gat gcc aag cgc acg ctc 336
 Lys Ile Ala Asn Ala Phe Asp Asn Lys Ile Asp Ala Lys Arg Thr Leu
 100 105 110

cgc gag atc aag ctg ctc cgc cac atg gac cac gag aat att gtt gca 384
 Arg Glu Ile Lys Leu Leu Arg His Met Asp His Glu Asn Ile Val Ala
 115 120 125

ata agg gga atc ata cct cct gcg cag agg gct gca ttc aat gat gtg 432
 Ile Arg Gly Ile Ile Pro Pro Ala Gln Arg Ala Ala Phe Asn Asp Val
 130 135 140

tat att gca tat gaa ttg atg gat act gat ctg cat caa att att cgt 480
 Tyr Ile Ala Tyr Glu Leu Met Asp Thr Asp Leu His Gln Ile Ile Arg
 145 150 155 160

tca aat caa gct ttg tca gag gag cac tgt cag tat ttt ctt tat caa 528
 Ser Asn Gln Ala Leu Ser Glu Glu His Cys Gln Tyr Phe Leu Tyr Gln
 165 170 175

att ctt cgt ggc ttg aag tat ata cat tca gca aat gtt ctt cac cgt 576

- 23 -

Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu His Arg	
180 185 190	
gac ttg aag cct agc aat ctt ctt ttg aat gca aac tgt gac ctc aag	624
Asp Leu Lys Pro Ser Asn Leu Leu Leu Asn Ala Asn Cys Asp Leu Lys	
195 200 205	
ata tgt gat ttt ggg ctt gct cgc acc acc tca gaa act gat ttt atg	672
Ile Cys Asp Phe Gly Leu Ala Arg Thr Thr Ser Glu Thr Asp Phe Met	
210 215 220	
act gaa tat gtt gtc aca aga tgg tat aga gca cca gag ctt ttg ttg	720
Thr Glu Tyr Val Val Thr Arg Trp Tyr Arg Ala Pro Glu Leu Leu Leu	
225 230 235 240	
aac tcc tca gaa tat act gct gcc atc gat gtg tgg tct gtg ggc tgt	768
Asn Ser Ser Glu Tyr Thr Ala Ala Ile Asp Val Trp Ser Val Gly Cys	
245 250 255	
ata ttt atg gaa ctg atg gac cgc aaa ccc ttg ttt cct gga aga gac	816
Ile Phe Met Glu Leu Met Asp Arg Lys Pro Leu Phe Pro Gly Arg Asp	
260 265 270	
cat gtc cat cag cta cgt ctt cta atg gag ctc att ggc acg cca aat	864
His Val His Gln Leu Arg Leu Leu Met Glu Leu Ile Gly Thr Pro Asn	
275 280 285	
gag ggt gat ctt gat ttt gta aat gaa aat gca aga aga tat atc cgc	912
Glu Gly Asp Leu Asp Phe Val Asn Glu Asn Ala Arg Arg Tyr Ile Arg	
290 295 300	
caa ctt cct cgt cac cct aga cag tcc tta cct gaa aaa ttt cca cat	960
Gln Leu Pro Arg His Pro Arg Gln Ser Leu Pro Glu Lys Phe Pro His	
305 310 315 320	
gta caa ccc tta gca att gac ctg gtg gaa aag atg ctg act ttt gat	1008
Val Gln Pro Leu Ala Ile Asp Leu Val Glu Lys Met Leu Thr Phe Asp	
325 330 335	
cct aga cag aga ata act gtt gaa ggc gca ctt gca cac cct tac ttg	1056
Pro Arg Gln Arg Ile Thr Val Glu Gly Ala Leu Ala His Pro Tyr Leu	
340 345 350	
gca tca ctt cat gac ata agt gat gag cca ggc tgc tca atg ccc ttc	1104
Ala Ser Leu His Asp Ile Ser Asp Glu Pro Gly Cys Ser Met Pro Phe	
355 360 365	
agc ttt gac ttc gag cag cat gca ttg tct gaa gaa caa atg aag gat	1152
Ser Phe Asp Phe Glu Gln His Ala Leu Ser Glu Glu Gln Met Lys Asp	
370 375 380	
ctt atc tac caa gag gct ctt gca ttc aac cct gat tat cag	1194
Leu Ile Tyr Gln Glu Ala Leu Ala Phe Asn Pro Asp Tyr Gln	
385 390 395	
tag	1197
<210> 30	
<211> 398	
<212> PRT	
<213> Zea mays	

- 24 -

<400> 30
 Met Asp Gly Gly Gly Gln Pro Pro Asp Thr Glu Met Ser Glu Ala Gly
 1 5 10 15
 Ala Gly Gly Gly Gly Gln Pro Pro Gln Gln Pro Leu Pro Pro Val Gly
 20 25 30
 Gly Gly Val Met Leu Asp Asn Ile Gln Ala Thr Leu Ser His Gly Gly
 35 40 45
 Arg Phe Ile Gln Tyr Asn Ile Phe Gly Asn Val Phe Glu Val Thr Ala
 50 55 60
 Lys Tyr Lys Pro Pro Val Leu Pro Ile Gly Lys Gly Ala Tyr Gly Ile
 65 70 75 80
 Val Cys Ser Ala Leu Asn Ser Glu Thr Ala Glu Gln Val Ala Ile Lys
 85 90 95
 Lys Ile Ala Asn Ala Phe Asp Asn Lys Ile Asp Ala Lys Arg Thr Leu
 100 105 110
 Arg Glu Ile Lys Leu Leu Arg His Met Asp His Glu Asn Ile Val Ala
 115 120 125
 Ile Arg Gly Ile Ile Pro Pro Ala Gln Arg Ala Ala Phe Asn Asp Val
 130 135 140
 Tyr Ile Ala Tyr Glu Leu Met Asp Thr Asp Leu His Gln Ile Ile Arg
 145 150 155 160
 Ser Asn Gln Ala Leu Ser Glu Glu His Cys Gln Tyr Phe Leu Tyr Gln
 165 170 175
 Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu His Arg
 180 185 190
 Asp Leu Lys Pro Ser Asn Leu Leu Leu Asn Ala Asn Cys Asp Leu Lys
 195 200 205
 Ile Cys Asp Phe Gly Leu Ala Arg Thr Thr Ser Glu Thr Asp Phe Met
 210 215 220
 Thr Glu Tyr Val Val Thr Arg Trp Tyr Arg Ala Pro Glu Leu Leu Leu
 225 230 235 240
 Asn Ser Ser Glu Tyr Thr Ala Ala Ile Asp Val Trp Ser Val Gly Cys
 245 250 255
 Ile Phe Met Glu Leu Met Asp Arg Lys Pro Leu Phe Pro Gly Arg Asp
 260 265 270
 His Val His Gln Leu Arg Leu Leu Met Glu Leu Ile Gly Thr Pro Asn
 275 280 285
 Glu Gly Asp Leu Asp Phe Val Asn Glu Asn Ala Arg Arg Tyr Ile Arg
 290 295 300
 Gln Leu Pro Arg His Pro Arg Gln Ser Leu Pro Glu Lys Phe Pro His
 305 310 315 320
 Val Gln Pro Leu Ala Ile Asp Leu Val Glu Lys Met Leu Thr Phe Asp
 325 330 335
 Pro Arg Gln Arg Ile Thr Val Glu Gly Ala Leu Ala His Pro Tyr Leu
 340 345 350
 Ala Ser Leu His Asp Ile Ser Asp Glu Pro Gly Cys Ser Met Pro Phe
 355 360 365
 Ser Phe Asp Phe Glu Gln His Ala Leu Ser Glu Glu Gln Met Lys Asp
 370 375 380
 Leu Ile Tyr Gln Glu Ala Leu Ala Phe Asn Pro Asp Tyr Gln
 385 390 395

<210> 31

<211> 20

<212> DNA

<213> Zea mays

<400> 31

atggacggcg gggggcagcc

<210> 32

<211> 20

20

<212> DNA
<213> Zea mays

<400> 32
ctactgataa tcagggttga 20

<210> 33
<211> 1197
<212> DNA
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<220>
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1 5 10 15

ttg ggc ggc ggg ggg caa ccg ccg ccg ccg ccg caa cag ccg gcg ggc 96
Leu Gly Gly Gly Gly Gln Pro Pro Pro Pro Pro Gln Gln Pro Ala Gly
20 25 30

ggg gcc ggg atg atg gag aac atc cac gcg acg ctc agc cac ggt ggc 144
Gly Ala Gly Met Met Glu Asn Ile His Ala Thr Leu Ser His Gly Gly
35 40 45

cgc ttc atc cag tac aac atc ttc ggc aac gtg ttc gag gtc acc tcc 192
Arg Phe Ile Gln Tyr Asn Ile Phe Gly Asn Val Phe Glu Val Thr Ser
50 55 60

aag tac aag ccc ccc atc ctc ccc atc ggc aag ggc gcc tac ggc atc 240
Lys Tyr Lys Pro Pro Ile Leu Pro Ile Gly Lys Gly Ala Tyr Gly Ile
65 70 75 80

gtc tgc tcg gcg ctc aac tcc gag acg gca gag cag gtg gcc atc aag 288
Val Cys Ser Ala Leu Asn Ser Glu Thr Ala Glu Gln Val Ala Ile Lys
85 90 95

aag atc gcc aac gcc ttc gac aac aag atc gat gcc aag cgc acg ctc 336
Lys Ile Ala Asn Ala Phe Asp Asn Lys Ile Asp Ala Lys Arg Thr Leu
100 105 110

cgc gag atc aag ctg ctc cgc cac atg gac cac gag aat att gtt gca 384
Arg Glu Ile Lys Leu Leu Arg His Met Asp His Glu Asn Ile Val Ala
115 120 125

ata aga gat atc ata cct cct cca ttg agg gag gca ttc aat gat gtg 432
Ile Arg Asp Ile Ile Pro Pro Pro Leu Arg Glu Ala Phe Asn Asp Val
130 135 140

tat att gcc tat gaa ttg atg gat act gat ctg cat caa att att cgt 480
Tyr Ile Ala Tyr Glu Leu Met Asp Thr Asp Leu His Gln Ile Ile Arg
145 150 155 160

tca aat caa gct ttg tca gag gag cac tgt cag tat ttt ctt tat caa 528
Ser Asn Gln Ala Leu Ser Glu Glu His Cys Gln Tyr Phe Leu Tyr Gln
165 170 175

att ctt cgt ggc ttg aag tat ata cat tca gca aat gtc ctt cac cgt 576
Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu His Arg

180	185	190	
gac ttg aag cct agc aat ctt ctt ttg aat gca aac tgt gac ctc aag			624
Asp Leu Lys Pro Ser Asn Leu Leu Leu Asn Ala Asn Cys Asp Leu Lys			
195	200	205	
ata tgt gat ttt ggg ctt gct cgc acc acc tca gaa act gat ttt atg			672
Ile Cys Asp Phe Gly Leu Ala Arg Thr Thr Ser Glu Thr Asp Phe Met			
210	215	220	
act gaa tat gtt gtc aca aga tgg tat aga gca cca gag ctt tta ttg			720
Thr Glu Tyr Val Val Thr Arg Trp Tyr Arg Ala Pro Glu Leu Leu			
225	230	235	240
aac tcc tct gaa tat act gct gcc att gat gtg tgg tct gtg ggc tgt			768
Asn Ser Ser Glu Tyr Thr Ala Ala Ile Asp Val Trp Ser Val Gly Cys			
245	250	255	
ata ttt atg gaa ctg atg gac cga aaa ccc ttg ttt cct gga aga gat			816
Ile Phe Met Glu Leu Met Asp Arg Lys Pro Leu Phe Pro Gly Arg Asp			
260	265	270	
cat gtc cat cag cta cgt cta cta atg gag ctc att gga aca ccg aat			864
His Val His Gln Leu Arg Leu Leu Met Glu Leu Ile Gly Thr Pro Asn			
275	280	285	
gag gct gat ctt gat ttt gta aat gaa aat gca aga aga tat atc cgc			912
Glu Ala Asp Leu Asp Phe Val Asn Glu Asn Ala Arg Arg Tyr Ile Arg			
290	295	300	
caa ctt ccc tgt cat gct aga cag tcc ttc cct gaa aaa ttt cca cat			960
Gln Leu Pro Cys His Ala Arg Gln Ser Phe Pro Glu Lys Phe Pro His			
305	310	315	320
gta caa cct tta gca att gac cta gtg gaa aag atg cta act ttt gat			1008
Val Gln Pro Leu Ala Ile Asp Leu Val Glu Lys Met Leu Thr Phe Asp			
325	330	335	
cct aga cag aga ata act gtt gaa ggc gca ctt gca cac cct tac ttg			1056
Pro Arg Gln Arg Ile Thr Val Glu Gly Ala Leu Ala His Pro Tyr Leu			
340	345	350	
gca tca ctt cat gac ata agt gat gag cca gtc tgc tca atg ccc ttc			1104
Ala Ser Leu His Asp Ile Ser Asp Glu Pro Val Cys Ser Met Pro Phe			
355	360	365	
agc ttc gac ttc gag cag cat gca tta tct gaa gaa cag atg aag gat			1152
Ser Phe Asp Phe Glu Gln His Ala Leu Ser Glu Glu Gln Met Lys Asp			
370	375	380	
ctg atc tac caa gag gct ctt gca ttc aac cca gat tac cag			1194
Leu Ile Tyr Gln Glu Ala Leu Ala Phe Asn Pro Asp Tyr Gln			
385	390	395	
tag			1197

<210> 34
 <211> 398
 <212> PRT
 <213> Zea mays

<400> 34

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Met Asp Gly Gly Gly Gln Pro Pro Asp Thr Glu Met Thr Asp Ala Gly
 1 5 10 15
 Leu Gly Gly Gly Gly Gln Pro Pro Pro Pro Gln Gln Pro Ala Gly
 20 25 30
 Gly Ala Gly Met Met Glu Asn Ile His Ala Thr Leu Ser His Gly Gly
 35 40 45
 Arg Phe Ile Gln Tyr Asn Ile Phe Gly Asn Val Phe Glu Val Thr Ser
 50 55 60
 Lys Tyr Lys Pro Pro Ile Leu Pro Ile Gly Lys Gly Ala Tyr Gly Ile
 65 70 75 80
 Val Cys Ser Ala Leu Asn Ser Glu Thr Ala Glu Gln Val Ala Ile Lys
 85 90 95
 Lys Ile Ala Asn Ala Phe Asp Asn Lys Ile Asp Ala Lys Arg Thr Leu
 100 105 110
 Arg Glu Ile Lys Leu Leu Arg His Met Asp His Glu Asn Ile Val Ala
 115 120 125
 Ile Arg Asp Ile Ile Pro Pro Pro Leu Arg Glu Ala Phe Asn Asp Val
 130 135 140
 Tyr Ile Ala Tyr Glu Leu Met Asp Thr Asp Leu His Gln Ile Ile Arg
 145 150 155 160
 Ser Asn Gln Ala Leu Ser Glu Glu His Cys Gln Tyr Phe Leu Tyr Gln
 165 170 175
 Ile Leu Arg Gly Leu Lys Tyr Ile His Ser Ala Asn Val Leu His Arg
 180 185 190
 Asp Leu Lys Pro Ser Asn Leu Leu Leu Asn Ala Asn Cys Asp Leu Lys
 195 200 205
 Ile Cys Asp Phe Gly Leu Ala Arg Thr Thr Ser Glu Thr Asp Phe Met
 210 215 220
 Thr Glu Tyr Val Val Thr Arg Trp Tyr Arg Ala Pro Glu Leu Leu Leu
 225 230 235 240
 Asn Ser Ser Glu Tyr Thr Ala Ala Ile Asp Val Trp Ser Val Gly Cys
 245 250 255
 Ile Phe Met Glu Leu Met Asp Arg Lys Pro Leu Phe Pro Gly Arg Asp
 260 265 270
 His Val His Gln Leu Arg Leu Leu Met Glu Leu Ile Gly Thr Pro Asn
 275 280 285
 Glu Ala Asp Leu Asp Phe Val Asn Glu Asn Ala Arg Arg Tyr Ile Arg
 290 295 300
 Gln Leu Pro Cys His Ala Arg Gln Ser Phe Pro Glu Lys Phe Pro His
 305 310 315 320
 Val Gln Pro Leu Ala Ile Asp Leu Val Glu Lys Met Leu Thr Phe Asp
 325 330 335
 Pro Arg Gln Arg Ile Thr Val Glu Gly Ala Leu Ala His Pro Tyr Leu
 340 345 350
 Ala Ser Leu His Asp Ile Ser Asp Glu Pro Val Cys Ser Met Pro Phe
 355 360 365
 Ser Phe Asp Phe Glu Gln His Ala Leu Ser Glu Glu Gln Met Lys Asp
 370 375 380
 Leu Ile Tyr Gln Glu Ala Leu Ala Phe Asn Pro Asp Tyr Gln
 385 390 395

<210> 35
 <211> 20
 <212> DNA
 <213> Zea mays

<400> 35
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20

<210> 36
 <211> 20
 <212> DNA

<213> Zea mays

<400> 36

ctactggtaa tctgggttga

20

<210> 37

<211> 1053

<212> DNA

<213> Zea mays

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<400> 37

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Met	Ala	Thr	Pro	Arg	Lys	Pro	Ile	Lys	Leu	Thr	Leu	Pro	Ser	His	Glu	
1				5					10					15		

acc	acc	atc	ggc	aag	ttc	ctg	acg	cac	agc	ggg	acg	ttc	acg	gac	ggg	96
Thr	Thr	Ile	Gly	Lys	Phe	Leu	Thr	His	Ser	Gly	Thr	Phe	Thr	Asp	Gly	
			20					25					30			

gat	ctg	cgc	gtc	aac	aag	gac	ggc	ctc	cgc	atc	gtc	tcg	cgg	agg	gag	144
Asp	Leu	Arg	Val	Asn	Lys	Asp	Gly	Leu	Arg	Ile	Val	Ser	Arg	Arg	Glu	
		35					40					45				

gga	ggc	gag	gct	cct	cct	ata	gag	ccg	ttg	gat	agt	caa	ctg	agc	tta	192
Gly	Gly	Glu	Ala	Pro	Pro	Ile	Glu	Pro	Leu	Asp	Ser	Gln	Leu	Ser	Leu	
	50					55				60						

gat	gat	cta	gac	gtt	ata	aaa	gtg	atc	ggg	aaa	ggg	agc	agc	gga	aat	240
Asp	Asp	Leu	Asp	Val	Ile	Lys	Val	Ile	Gly	Lys	Gly	Ser	Ser	Gly	Asn	
	65				70				75					80		

gtg	caa	ttg	gtc	cgc	cac	aaa	ttt	act	ggc	cag	ttt	ttt	gct	ctg	aag	288
Val	Gln	Leu	Val	Arg	His	Lys	Phe	Thr	Gly	Gln	Phe	Phe	Ala	Leu	Lys	
			85						90					95		

gtt	att	caa	cta	aat	att	gat	gag	agt	ata	cgc	aaa	cag	att	gcc	aag	336
Val	Ile	Gln	Leu	Asn	Ile	Asp	Glu	Ser	Ile	Arg	Lys	Gln	Ile	Ala	Lys	
			100					105					110			

gag	ttg	aag	ata	aac	tta	tca	aca	cag	tgc	caa	tat	gtt	gtt	gtg	ttc	384
Glu	Leu	Lys	Ile	Asn	Leu	Ser	Thr	Gln	Cys	Gln	Tyr	Val	Val	Val	Phe	
		115					120					125				

tat	cag	tgt	ttc	tat	ttc	aat	ggg	gcc	att	tct	att	gtt	ttg	gaa	tac	432
Tyr	Gln	Cys	Phe	Tyr	Phe	Asn	Gly	Ala	Ile	Ser	Ile	Val	Leu	Glu	Tyr	
	130					135					140					

atg	gat	ggg	ggc	tcc	ctt	gca	gat	ttc	ctg	aag	act	gtt	aaa	acc	att	480
Met	Asp	Gly	Gly	Ser	Leu	Ala	Asp	Phe	Leu	Lys	Thr	Val	Lys	Thr	Ile	
	145				150				155					160		

cca	gag	gcc	tac	ctc	gct	gct	atc	tgt	acg	cag	atg	cta	aaa	gga	ctg	528
Pro	Glu	Ala	Tyr	Leu	Ala	Ala	Ile	Cys	Thr	Gln	Met	Leu	Lys	Gly	Leu	
			165					170						175		

atc	tat	ttg	cat	aac	gag	aag	cgc	gtt	ata	cac	cga	gat	ctg	aaa	cca	576
Ile	Tyr	Leu	His	Asn	Glu	Lys	Arg	Val	Ile	His	Arg	Asp	Leu	Lys	Pro	
			180				185						190			

tca aat ata ttg ata aat cat agg ggt gaa gta aaa ata tca gat ttt 624
 Ser Asn Ile Leu Ile Asn His Arg Gly Glu Val Lys Ile Ser Asp Phe
 195 200 205

ggt gtg agt gcc att ata tct agt tcc tct tct caa cga gat aca ttt 672
 Gly Val Ser Ala Ile Ile Ser Ser Ser Ser Ser Gln Arg Asp Thr Phe
 210 215 220

att ggc aca cgc aac tac atg gcg cca gaa aga atc gat gga aag aaa 720
 Ile Gly Thr Arg Asn Tyr Met Ala Pro Glu Arg Ile Asp Gly Lys Lys
 225 230 235 240

cat ggt tct atg agt gat atc tgg agt ttg gga cta gtg ata ctg gaa 768
 His Gly Ser Met Ser Asp Ile Trp Ser Leu Gly Leu Val Ile Leu Glu
 245 250 255

tgt gca acc ggc atc ttt cca ttt cct cct tgt gaa agc ttc tac gaa 816
 Cys Ala Thr Gly Ile Phe Pro Phe Pro Pro Cys Glu Ser Phe Tyr Glu
 260 265 270

ctt ctc gtg gct gtt gtt gat caa ccg cca cct tct gcg ccg ccg gat 864
 Leu Leu Val Ala Val Val Asp Gln Pro Pro Pro Ser Ala Pro Pro Asp
 275 280 285

cag ttt tca cca gaa ttc tgt ggg ttc att tct gca tgt ctc cag aag 912
 Gln Phe Ser Pro Glu Phe Cys Gly Phe Ile Ser Ala Cys Leu Gln Lys
 290 295 300

gat gct aat gac agg tca tca gcc caa gcc tta ttg gac cat ccg ttc 960
 Asp Ala Asn Asp Arg Ser Ser Ala Gln Ala Leu Leu Asp His Pro Phe
 305 310 315 320

ctg agc atg tat gat gac ctg cat gta gat ctt gct tct tac ttc acg 1008
 Leu Ser Met Tyr Asp Asp Leu His Val Asp Leu Ala Ser Tyr Phe Thr
 325 330 335

aca gca gga tct cct ctc gcc acc ttc aat tcc agg caa ctc 1050
 Thr Ala Gly Ser Pro Leu Ala Thr Phe Asn Ser Arg Gln Leu
 340 345 350

taa 1053

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 <213> Zea mays

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 1 5 10 15
 Thr Thr Ile Gly Lys Phe Leu Thr His Ser Gly Thr Phe Thr Asp Gly
 20 25 30
 Asp Leu Arg Val Asn Lys Asp Gly Leu Arg Ile Val Ser Arg Arg Glu
 35 40 45
 Gly Gly Glu Ala Pro Pro Ile Glu Pro Leu Asp Ser Gln Leu Ser Leu
 50 55 60
 Asp Asp Leu Asp Val Ile Lys Val Ile Gly Lys Gly Ser Ser Gly Asn
 65 70 75 80
 Val Gln Leu Val Arg His Lys Phe Thr Gly Gln Phe Phe Ala Leu Lys
 85 90 95
 Val Ile Gln Leu Asn Ile Asp Glu Ser Ile Arg Lys Gln Ile Ala Lys

- 30 -

				100					105					110			
Glu	Leu	Lys	Ile	Asn	Leu	Ser	Thr	Gln	Cys	Gln	Tyr	Val	Val	Val	Phe		
				115				120				125					
Tyr	Gln	Cys	Phe	Tyr	Phe	Asn	Gly	Ala	Ile	Ser	Ile	Val	Leu	Glu	Tyr		
				130			135				140						
Met	Asp	Gly	Gly	Ser	Leu	Ala	Asp	Phe	Leu	Lys	Thr	Val	Lys	Thr	Ile		
145					150					155					160		
Pro	Glu	Ala	Tyr	Leu	Ala	Ala	Ile	Cys	Thr	Gln	Met	Leu	Lys	Gly	Leu		
				165					170					175			
Ile	Tyr	Leu	His	Asn	Glu	Lys	Arg	Val	Ile	His	Arg	Asp	Leu	Lys	Pro		
				180				185					190				
Ser	Asn	Ile	Leu	Ile	Asn	His	Arg	Gly	Glu	Val	Lys	Ile	Ser	Asp	Phe		
				195			200					205					
Gly	Val	Ser	Ala	Ile	Ile	Ser	Ser	Ser	Ser	Ser	Gln	Arg	Asp	Thr	Phe		
						215					220						
Ile	Gly	Thr	Arg	Asn	Tyr	Met	Ala	Pro	Glu	Arg	Ile	Asp	Gly	Lys	Lys		
225					230					235					240		
His	Gly	Ser	Met	Ser	Asp	Ile	Trp	Ser	Leu	Gly	Leu	Val	Ile	Leu	Glu		
				245					250					255			
Cys	Ala	Thr	Gly	Ile	Phe	Pro	Phe	Pro	Pro	Cys	Glu	Ser	Phe	Tyr	Glu		
				260				265					270				
Leu	Leu	Val	Ala	Val	Val	Asp	Gln	Pro	Pro	Pro	Ser	Ala	Pro	Pro	Asp		
				275			280					285					
Gln	Phe	Ser	Pro	Glu	Phe	Cys	Gly	Phe	Ile	Ser	Ala	Cys	Leu	Gln	Lys		
				290			295				300						
Asp	Ala	Asn	Asp	Arg	Ser	Ser	Ala	Gln	Ala	Leu	Leu	Asp	His	Pro	Phe		
305					310					315					320		
Leu	Ser	Met	Tyr	Asp	Asp	Leu	His	Val	Asp	Leu	Ala	Ser	Tyr	Phe	Thr		
				325					330					335			
Thr	Ala	Gly	Ser	Pro	Leu	Ala	Thr	Phe	Asn	Ser	Arg	Gln	Leu				
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<213> Zea mays
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20

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<212> DNA
<213> Zea mays
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20

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<222> (1) ... (843)
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Met Ala Gln Glu Leu His Glu Thr Ser Ser Cys Ser Ala Thr Thr Thr
1 5 10 15

48

tcg tcg tgc acc aca tcc tgc tgc tcg tcc act gtc aca gac tcg tcc	96
Ser Ser Cys Thr Thr Ser Cys Cys Ser Ser Thr Val Thr Asp Ser Ser	
20 25 30	
tct tcg ccc ccg tca ccg gcg gcg gcc aat gcc gcg ccc gcg aca cgg	144
Ser Ser Pro Pro Ser Pro Ala Ala Ala Asn Ala Ala Pro Ala Thr Arg	
35 40 45	
aag cgg cag gcg ttg gag gcc gag gcc gag gcc gag gcg ggc ggt gag	192
Lys Arg Gln Ala Leu Glu Ala Glu Ala Glu Ala Glu Ala Gly Gly Glu	
50 55 60	
gag gag gag gag gag gag gga agc tgt gct ggt aat aaa gcg gcg ccg	240
Glu Glu Glu Glu Glu Glu Gly Ser Cys Ala Gly Asn Lys Ala Ala Pro	
65 70 75 80	
gcc aag aag cga ccg cgg ggc agc gag ggg aaa cac ccg acg ttc cgc	288
Ala Lys Lys Arg Pro Arg Gly Ser Glu Gly Lys His Pro Thr Phe Arg	
85 90 95	
ggc gtg cgg atg cgg gcg tgg ggc aag tgg gtg tcg gag atc cgc gag	336
Gly Val Arg Met Arg Ala Trp Gly Lys Trp Val Ser Glu Ile Arg Glu	
100 105 110	
ccg cgc aag aag tcg cgc ata tgg ctc ggc acg ttc ccc acc gcc gag	384
Pro Arg Lys Lys Ser Arg Ile Trp Leu Gly Thr Phe Pro Thr Ala Glu	
115 120 125	
atg gcc gcg cgc gcc cac gac gtc gcg gcg ctc gcc atc aaa ggc cgc	432
Met Ala Ala Arg Ala His Asp Val Ala Ala Leu Ala Ile Lys Gly Arg	
130 135 140	
gcc gcg cac ctc aac ttc ccg gac ttc gcc ggc gcg ctc ccg cgc gcc	480
Ala Ala His Leu Asn Phe Pro Asp Phe Ala Gly Ala Leu Pro Arg Ala	
145 150 155 160	
gcg tcc gcg gcg ccc aag gac gtc cag gca gcc gcc gca ttg gcc gct	528
Ala Ser Ala Ala Pro Lys Asp Val Gln Ala Ala Ala Ala Leu Ala Ala	
165 170 175	
gcg ttc acg tcg ccg tca tcg gag ccc ggc gcc ggc gcg cac gag gag	576
Ala Phe Thr Ser Pro Ser Ser Glu Pro Gly Ala Gly Ala His Glu Glu	
180 185 190	
ccc gct gcc aag gac ggc gcc gcg ccc gag gag gca gcc gcc gac gca	624
Pro Ala Ala Lys Asp Gly Ala Ala Pro Glu Glu Ala Ala Asp Ala	
195 200 205	
cag gca cca gta cca gta gca cta cca ccg ccg gcg gcc tct cgg cca	672
Gln Ala Pro Val Pro Val Ala Leu Pro Pro Pro Ala Ala Ser Arg Pro	
210 215 220	
ggg acg ccg tcg agc ggc gtg gag gac gag cgg cag ctg ttc gac ctg	720
Gly Thr Pro Ser Ser Gly Val Glu Asp Glu Arg Gln Leu Phe Asp Leu	
225 230 235 240	
ccg gac ctg ctc ctc gac atc ccg gac ggg ttc ggg cgc ttc ccg ccg	768
Pro Asp Leu Leu Asp Ile Arg Asp Gly Phe Gly Arg Phe Pro Pro	
245 250 255	
atg tgg gcc ccg ctc act gac gtg gag gag gtg gtc aat gcg gag ctg	816
Met Trp Ala Pro Leu Thr Asp Val Glu Glu Val Val Asn Ala Glu Leu	

- 32 -

260 265 270

cgc ctc gag gag ccg ctg ctt tgg gag tag 846

Arg Leu Glu Glu Pro Leu Leu Trp Glu

275 280

<210> 42

<211> 281

<212> PRT

<213> Zea mays

<400> 42

Met Ala Gln Glu Leu His Glu Thr Ser Ser Cys Ser Ala Thr Thr Thr

1 5 10 15

Ser Ser Cys Thr Thr Ser Cys Cys Ser Ser Thr Val Thr Asp Ser Ser

20 25 30

Ser Ser Pro Pro Ser Pro Ala Ala Ala Asn Ala Ala Pro Ala Thr Arg

35 40 45

Lys Arg Gln Ala Leu Glu Ala Glu Ala Glu Ala Gly Gly Glu

50 55 60

Glu Glu Glu Glu Glu Glu Gly Ser Cys Ala Gly Asn Lys Ala Ala Pro

65 70 75 80

Ala Lys Lys Arg Pro Arg Gly Ser Glu Gly Lys His Pro Thr Phe Arg

85 90 95

Gly Val Arg Met Arg Ala Trp Gly Lys Trp Val Ser Glu Ile Arg Glu

100 105 110

Pro Arg Lys Lys Ser Arg Ile Trp Leu Gly Thr Phe Pro Thr Ala Glu

115 120 125

Met Ala Ala Arg Ala His Asp Val Ala Ala Leu Ala Ile Lys Gly Arg

130 135 140

Ala Ala His Leu Asn Phe Pro Asp Phe Ala Gly Ala Leu Pro Arg Ala

145 150 155 160

Ala Ser Ala Ala Pro Lys Asp Val Gln Ala Ala Ala Leu Ala Ala

165 170 175

Ala Phe Thr Ser Pro Ser Ser Glu Pro Gly Ala Gly Ala His Glu Glu

180 185 190

Pro Ala Ala Lys Asp Gly Ala Ala Pro Glu Glu Ala Ala Ala Asp Ala

195 200 205

Gln Ala Pro Val Pro Val Ala Leu Pro Pro Pro Ala Ala Ser Arg Pro

210 215 220

Gly Thr Pro Ser Ser Gly Val Glu Asp Glu Arg Gln Leu Phe Asp Leu

225 230 235 240

Pro Asp Leu Leu Leu Asp Ile Arg Asp Gly Phe Gly Arg Phe Pro Pro

245 250 255

Met Trp Ala Pro Leu Thr Asp Val Glu Glu Val Val Asn Ala Glu Leu

260 265 270

Arg Leu Glu Glu Pro Leu Leu Trp Glu

275 280

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<211> 20

<212> DNA

<213> Zea mays

<400> 43

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20

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 <210> 45
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 gcgggcgggc gggcagccaa tgccgtcggg cgcgcagggc taggagc atg ggc gga 116
 Met Gly Gly
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 ggg ggc gcg atc gtg cac ggc ttc cgc cgc tgg ttc cac cgc cgg agc 164
 Gly Gly Ala Ile Val His Gly Phe Arg Arg Trp Phe His Arg Arg Ser
 5 10 15

 gaa ggc tcc acc tcc aac tcc aac cag tcc tcc ctc gcc ggc gag ggg 212
 Glu Gly Ser Thr Ser Asn Ser Asn Gln Ser Ser Leu Ala Gly Glu Gly 35
 20 25 30

 gaa gcg gac gac ggc tca tcc gat ctc gag gtc atc gag gac ccg gat 260
 Glu Ala Asp Asp Gly Ser Ser Asp Leu Glu Val Ile Glu Asp Pro Asp 50
 40 45

 ctc ctc ggc ctc cgc gcc atc cgc gtg ccc aag cgc aag atg ccg ctg 308
 Leu Leu Gly Leu Arg Ala Ile Arg Val Pro Lys Arg Lys Met Pro Leu 65
 55 60

 ccc gtc gag agc cac agg aag aac tca gta gaa atg gag ttc ttc aca 356
 Pro Val Glu Ser His Arg Lys Asn Ser Val Glu Met Glu Phe Phe Thr 80
 70 75

 gag tac gga gag gca agc cag tac caa atc caa gaa gtc att ggt aaa 404
 Glu Tyr Gly Glu Ala Ser Gln Tyr Gln Ile Gln Glu Val Ile Gly Lys 95
 85 90

 ggg agc tac gga gta gtt gct gca gct gtg gac acc cgc acc ggt gag 452
 Gly Ser Tyr Gly Val Val Ala Ala Ala Val Asp Thr Arg Thr Gly Glu 115
 100 105 110

 cgg gtt gct atc aag aag atc aat gat gtg ttt gag cac gtc tcc gat 500
 Arg Val Ala Ile Lys Lys Ile Asn Asp Val Phe Glu His Val Ser Asp 130
 120 125

 gcc aca cgc ata ctg cgg gag atc aag ctc ctc cgg ttg ctt cgt cac 548
 Ala Thr Arg Ile Leu Arg Glu Ile Lys Leu Leu Arg Leu Arg His 145
 135 140

 cca gac ata gtc gag att aag cac atc atg ctc cct cct tct cgg agg 596
 Pro Asp Ile Val Glu Ile Lys His Ile Met Leu Pro Pro Ser Arg Arg 160
 150 155

 gag ttc caa gac atc tat gtt gtt ttt gag ctc atg gag tcc gat ctc 644
 Glu Phe Gln Asp Ile Tyr Val Val Phe Glu Leu Met Glu Ser Asp Leu 175
 165 170

cat caa gtg atc agg gcg aat gat gac ctc acc cct gaa cat cac cag	692
His Gln Val Ile Arg Ala Asn Asp Asp Leu Thr Pro Glu His His Gln	
180 185 190 195	
ttt ttc ctg tac caa ctt ctt cga gct ctc aag tac ata cat gcc gct	740
Phe Phe Leu Tyr Gln Leu Leu Arg Ala Leu Lys Tyr Ile His Ala Ala	
200 205 210	
aat gta ttc cat cgt gat cta aag ccc aag aat ata ctg gct aac tcg	788
Asn Val Phe His Arg Asp Leu Lys Pro Lys Asn Ile Leu Ala Asn Ser	
215 220 225	
gac tgc aaa ctg aaa ata tgt gac ttt gga ctt gcc cgt gca tca ttc	836
Asp Cys Lys Leu Lys Ile Cys Asp Phe Gly Leu Ala Arg Ala Ser Phe	
230 235 240	
aat gat gcc cct tca gct ata ttt tgg aca gac tat gtg gca aca agg	884
Asn Asp Ala Pro Ser Ala Ile Phe Trp Thr Asp Tyr Val Ala Thr Arg	
245 250 255	
tgg tac cgg gca cct gaa cta tgt ggt tcc ttt ttc tcc aaa tac act	932
Trp Tyr Arg Ala Pro Glu Leu Cys Gly Ser Phe Phe Ser Lys Tyr Thr	
260 265 270 275	
cct gca att gat att tgg agc att ggg tgc ata ttt gct gaa ctt ctc	980
Pro Ala Ile Asp Ile Trp Ser Ile Gly Cys Ile Phe Ala Glu Leu Leu	
280 285 290	
act gga agg cca cta ttt cct ggg aag aat gtt gta cac caa tta gat	1028
Thr Gly Arg Pro Leu Phe Pro Gly Lys Asn Val Val His Gln Leu Asp	
295 300 305	
att ata aca gat ctt ctt ggg act cca tca tct gag acc tta tcc cgg	1076
Ile Ile Thr Asp Leu Leu Gly Thr Pro Ser Ser Glu Thr Leu Ser Arg	
310 315 320	
att cga aat gag aag gcc agg aga tat ttg agc tgc atg cgg aaa aaa	1124
Ile Arg Asn Glu Lys Ala Arg Arg Tyr Leu Ser Cys Met Arg Lys Lys	
325 330 335	
tat ccc gtg ccc ttt act cat aaa ttt cga cat gct gat cct ttg gct	1172
Tyr Pro Val Pro Phe Thr His Lys Phe Arg His Ala Asp Pro Leu Ala	
340 345 350 355	
ctt cgt ctt cta gag cgt ttg ctt gca ttt gat cct aaa gac cgg cct	1220
Leu Arg Leu Leu Glu Arg Leu Leu Ala Phe Asp Pro Lys Asp Arg Pro	
360 365 370	
agt gca gaa gag gct cta gct gat cca tac ttt gca tcc ctt gct aat	1268
Ser Ala Glu Glu Ala Leu Ala Asp Pro Tyr Phe Ala Ser Leu Ala Asn	
375 380 385	
gtg gaa cgt gag cca tca aga cac ccg att tct aaa ctt gag ttt gaa	1316
Val Glu Arg Glu Pro Ser Arg His Pro Ile Ser Lys Leu Glu Phe Glu	
390 395 400	
ttt gaa aga aag aag ctg gcc aaa gat gat gtt aga gaa ctg atc tat	1364
Phe Glu Arg Lys Lys Leu Ala Lys Asp Asp Val Arg Glu Leu Ile Tyr	
405 410 415	
cga gag att ttg gag tat cat cca cag atg ctg gag gag tac atg aaa	1412

Arg Glu Ile Leu Glu Tyr His Pro Gln Met Leu Glu Glu Tyr Met Lys
 420 425 430 435
 ggc gga gaa cag att agc ttc ctc tat cca agc gga gtc gac cgc ttc 1460
 Gly Gly Glu Gln Ile Ser Phe Leu Tyr Pro Ser Gly Val Asp Arg Phe
 440 445 450
 aaa ctg cag ttc gct cat ctt gag gag cat tac agc aaa gga gaa aga 1508
 Lys Leu Gln Phe Ala His Leu Glu Glu His Tyr Ser Lys Gly Glu Arg
 455 460 465
 ggt tcc cca ctg caa aga aag cat gct tcc tta cca agg gaa aga gtg 1556
 Gly Ser Pro Leu Gln Arg Lys His Ala Ser Leu Pro Arg Glu Arg Val
 470 475 480
 gtt gta tcc aag gat ggc aat act gaa caa caa att aat gac cag gag 1604
 Val Val Ser Lys Asp Gly Asn Thr Glu Gln Gln Ile Asn Asp Gln Glu
 485 490 495
 aga agc gca gat tct gtt gcc cgc act acc gtg agc cct cca agg tca 1652
 Arg Ser Ala Asp Ser Val Ala Arg Thr Thr Val Ser Pro Pro Arg Ser
 500 505 510 515
 gag gat gct gac atg gat gat gtg aaa ccc aca agc tta agc tct cgg 1700
 Glu Asp Ala Asp Met Asp Asp Val Lys Pro Thr Ser Leu Ser Ser Arg
 520 525 530
 agc tac ctg aag agt gca agc ata agc gct tcc aag tgc gtc gtt gtc 1748
 Ser Tyr Leu Lys Ser Ala Ser Ile Ser Ala Ser Lys Cys Val Val Val
 535 540 545
 aga aat aaa cac cca gag cca gag gat gat gga ata cct gag gag atg 1796
 Arg Asn Lys His Pro Glu Pro Glu Asp Asp Gly Ile Pro Glu Glu Met
 550 555 560
 gaa ggc acc gtc gat gga ttg tct gaa aag gtc tcc agg atg cac tcc 1844
 Glu Gly Thr Val Asp Gly Leu Ser Glu Lys Val Ser Arg Met His Ser
 565 570 575
 taaggcagga ctgtccatgg cgacagcaat ctgcacacaa gatttctgtc tgggtgactg 1904
 taatgccgac gacacactgt ttggatcttg tgtacaccca cagctagttc aactaagatg 1964
 tagcactagt tcgtccatgg aggaagcaga atggacgatg aagtgtggtc agcctgtcga 2024
 ccatgaggtg ttttcttcca atggagtcta gggatgttt gagttctaca acggcttgtg 2084
 ccgtacgaag tgtcagcagt gtacgaggag gaggatateg ttgtcagtag acattttgcc 2144
 tgtctagaaa attatgagta ctttctccaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa 2204
 aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaaaaaaaa aaaggcgccg cgc 2257

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 <211> 579
 <212> PRT
 <213> Zea mays

<400> 46
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 20 25 30
 Gly Glu Gly Glu Ala Asp Asp Gly Ser Ser Asp Leu Glu Val Ile Glu
 35 40 45
 Asp Pro Asp Leu Leu Gly Leu Arg Ala Ile Arg Val Pro Lys Arg Lys
 50 55 60
 Met Pro Leu Pro Val Glu Ser His Arg Lys Asn Ser Val Glu Met Glu

65					70					75				80	
Phe	Phe	Thr	Glu	Tyr	Gly	Glu	Ala	Ser	Gln	Tyr	Gln	Ile	Gln	Glu	Val
				85					90					95	
Ile	Gly	Lys	Gly	Ser	Tyr	Gly	Val	Val	Ala	Ala	Ala	Val	Asp	Thr	Arg
			100					105					110		
Thr	Gly	Glu	Arg	Val	Ala	Ile	Lys	Lys	Ile	Asn	Asp	Val	Phe	Glu	His
			115				120					125			
Val	Ser	Asp	Ala	Thr	Arg	Ile	Leu	Arg	Glu	Ile	Lys	Leu	Leu	Arg	Leu
			130				135				140				
Leu	Arg	His	Pro	Asp	Ile	Val	Glu	Ile	Lys	His	Ile	Met	Leu	Pro	Pro
			145			150				155				160	
Ser	Arg	Arg	Glu	Phe	Gln	Asp	Ile	Tyr	Val	Val	Phe	Glu	Leu	Met	Glu
				165				170						175	
Ser	Asp	Leu	His	Gln	Val	Ile	Arg	Ala	Asn	Asp	Asp	Leu	Thr	Pro	Glu
			180					185					190		
His	His	Gln	Phe	Phe	Leu	Tyr	Gln	Leu	Leu	Arg	Ala	Leu	Lys	Tyr	Ile
			195				200					205			
His	Ala	Ala	Asn	Val	Phe	His	Arg	Asp	Leu	Lys	Pro	Lys	Asn	Ile	Leu
			210				215				220				
Ala	Asn	Ser	Asp	Cys	Lys	Leu	Lys	Ile	Cys	Asp	Phe	Gly	Leu	Ala	Arg
			225			230				235				240	
Ala	Ser	Phe	Asn	Asp	Ala	Pro	Ser	Ala	Ile	Phe	Trp	Thr	Asp	Tyr	Val
			245					250					255		
Ala	Thr	Arg	Trp	Tyr	Arg	Ala	Pro	Glu	Leu	Cys	Gly	Ser	Phe	Phe	Ser
			260					265					270		
Lys	Tyr	Thr	Pro	Ala	Ile	Asp	Ile	Trp	Ser	Ile	Gly	Cys	Ile	Phe	Ala
			275			280						285			
Glu	Leu	Leu	Thr	Gly	Arg	Pro	Leu	Phe	Pro	Gly	Lys	Asn	Val	Val	His
			290			295					300				
Gln	Leu	Asp	Ile	Ile	Thr	Asp	Leu	Leu	Gly	Thr	Pro	Ser	Ser	Glu	Thr
			305			310				315				320	
Leu	Ser	Arg	Ile	Arg	Asn	Glu	Lys	Ala	Arg	Arg	Tyr	Leu	Ser	Cys	Met
				325				330					335		
Arg	Lys	Lys	Tyr	Pro	Val	Pro	Phe	Thr	His	Lys	Phe	Arg	His	Ala	Asp
			340					345					350		
Pro	Leu	Ala	Leu	Arg	Leu	Leu	Glu	Arg	Leu	Leu	Ala	Phe	Asp	Pro	Lys
			355				360					365			
Asp	Arg	Pro	Ser	Ala	Glu	Glu	Ala	Leu	Ala	Asp	Pro	Tyr	Phe	Ala	Ser
			370			375					380				
Leu	Ala	Asn	Val	Glu	Arg	Glu	Pro	Ser	Arg	His	Pro	Ile	Ser	Lys	Leu
			385			390				395				400	
Glu	Phe	Glu	Phe	Glu	Arg	Lys	Lys	Leu	Ala	Lys	Asp	Asp	Val	Arg	Glu
				405				410					415		
Leu	Ile	Tyr	Arg	Glu	Ile	Leu	Glu	Tyr	His	Pro	Gln	Met	Leu	Glu	Glu
			420					425					430		
Tyr	Met	Lys	Gly	Gly	Glu	Gln	Ile	Ser	Phe	Leu	Tyr	Pro	Ser	Gly	Val
			435			440						445			
Asp	Arg	Phe	Lys	Leu	Gln	Phe	Ala	His	Leu	Glu	Glu	His	Tyr	Ser	Lys
			450			455					460				
Gly	Glu	Arg	Gly	Ser	Pro	Leu	Gln	Arg	Lys	His	Ala	Ser	Leu	Pro	Arg
			465			470				475				480	
Glu	Arg	Val	Val	Val	Ser	Lys	Asp	Gly	Asn	Thr	Glu	Gln	Gln	Ile	Asn
				485				490					495		
Asp	Gln	Glu	Arg	Ser	Ala	Asp	Ser	Val	Ala	Arg	Thr	Thr	Val	Ser	Pro
			500					505					510		
Pro	Arg	Ser	Glu	Asp	Ala	Asp	Met	Asp	Asp	Val	Lys	Pro	Thr	Ser	Leu
			515				520					525			
Ser	Ser	Arg	Ser	Tyr	Leu	Lys	Ser	Ala	Ser	Ile	Ser	Ala	Ser	Lys	Cys
			530			535					540				
Val	Val	Val	Arg	Asn	Lys	His	Pro	Glu	Pro	Glu	Asp	Asp	Gly	Ile	Pro
			545			550				555				560	
Glu	Glu	Met	Glu	Gly	Thr	Val	Asp	Gly	Leu	Ser	Glu	Lys	Val	Ser	Arg

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                    565                    570                    575
Met His Ser

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<213> Zea mays

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<210> 48
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<400> 48
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<210> 49
<211> 938
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                    1                    5

cac acc tcc tcc agc ctt tct tcc tct aca acc cca att ctt ccc tgc    104
His Thr Ser Ser Ser Leu Ser Ser Ser Thr Thr Pro Ile Leu Pro Cys
                    10                    15                    20

agc tct gct gct gcc ttg ttt cga tgc gtc atg gcg gcg gtg gcc aca    152
Ser Ser Ala Ala Ala Leu Phe Arg Ser Val Met Ala Ala Val Ala Thr
                    25                    30                    35

gag act cgg ttc cat gtc ctg gcg gtg gac gac agc ctc ccg gac agg    200
Glu Thr Pro Phe His Val Leu Ala Val Asp Asp Ser Leu Pro Asp Arg
                    40                    45                    50

aag ctc atc gag agg ctc ctc aag acc tct tcc ttc caa gtg acc act    248
Lys Leu Ile Glu Arg Leu Leu Lys Thr Ser Ser Phe Gln Val Thr Thr
                    55                    60                    65                    70

gtc gac tcc ggg agc aag gcg ctg cag ttc ctg ggc ctc cat gac cag    296
Val Asp Ser Gly Ser Lys Ala Leu Gln Phe Leu Gly Leu His Asp Gln
                    75                    80                    85

gac agc acg gtt cct cct gtc cac acg cac cag ctg gat gtg gct gcc    344
Asp Ser Thr Val Pro Pro Val His Thr His Gln Leu Asp Val Ala Ala
                    90                    95                    100

aat cag gat gtg gct gtg aac ctg atc atc aca gac tac tgc atg cct    392
Asn Gln Asp Val Ala Val Asn Leu Ile Ile Thr Asp Tyr Cys Met Pro
                    105                    110                    115

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ggc atg aca gga tat gac ctg ctc aag aag atc aag gag tcg tcg tct 440
 Gly Met Thr Gly Tyr Asp Leu Leu Lys Lys Ile Lys Glu Ser Ser Ser
 120 125 130

ctc aga gat atc ccg gtg gtg atc atg tcc tct gag aac att cct tca 488
 Leu Arg Asp Ile Pro Val Val Ile Met Ser Ser Glu Asn Ile Pro Ser
 135 140 145 150

agg atc aat agg tgc ctg gag gaa gga gct gac gag ttc ttc cta aaa 536
 Arg Ile Asn Arg Cys Leu Glu Glu Gly Ala Asp Glu Phe Phe Leu Lys
 155 160 165

cct gtg cgg cta tca gac atg aac aag ctc aag ccc cac ata ctg aaa 584
 Pro Val Arg Leu Ser Asp Met Asn Lys Leu Lys Pro His Ile Leu Lys
 170 175 180

agc aga tgc aac cag gaa cag cac cag caa agt gac agt cac agt ggc 632
 Ser Arg Cys Asn Gln Glu Gln His Gln Gln Ser Asp Ser His Ser Gly
 185 190 195

gaa cgc agg aac ccc aca atc agc agc agc gat agc ata aac aac cgc 680
 Glu Arg Arg Asn Pro Thr Ile Ser Ser Ser Asp Ser Ile Asn Asn Arg
 200 205 210

aag aga aag ggt gca ggc acc gaa gaa atc ttg ccc cag ctg gca aac 728
 Lys Arg Lys Gly Ala Gly Thr Glu Glu Ile Leu Pro Gln Leu Ala Asn
 215 220 225 230

aga tca agg cac agt taactgagaa ctgactagta cagctaaaac cttttctttt 783
 Arg Ser Arg His Ser
 235

tcacttttact ttggttact gatttgtaac gtagatgtta gccctggatt tgcaaaacgg 843
 aacggaatct gtaactgat actgcttgag tcgaatcgat cgaagtgatt ctgcaccaga 903
 tctccctaaa aaaaaaaaaa aaaaaaaaaac tcgag 938

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 <213> Zea mays

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 Met Ala Ala Val Ala Thr Glu Thr Pro Phe His Val Leu Ala Val Asp
 35 40 45
 Asp Ser Leu Pro Asp Arg Lys Leu Ile Glu Arg Leu Leu Lys Thr Ser
 50 55 60
 Ser Phe Gln Val Thr Thr Val Asp Ser Gly Ser Lys Ala Leu Gln Phe
 65 70 75 80
 Leu Gly Leu His Asp Gln Asp Ser Thr Val Pro Pro Val His Thr His
 85 90 95
 Gln Leu Asp Val Ala Ala Asn Gln Asp Val Ala Val Asn Leu Ile Ile
 100 105 110
 Thr Asp Tyr Cys Met Pro Gly Met Thr Gly Tyr Asp Leu Leu Lys Lys
 115 120 125
 Ile Lys Glu Ser Ser Ser Leu Arg Asp Ile Pro Val Val Ile Met Ser
 130 135 140
 Ser Glu Asn Ile Pro Ser Arg Ile Asn Arg Cys Leu Glu Glu Gly Ala
 145 150 155 160

Asp Glu Phe Phe Leu Lys Pro Val Arg Leu Ser Asp Met Asn Lys Leu
 165 170 175
 Lys Pro His Ile Leu Lys Ser Arg Cys Asn Gln Glu Gln His Gln Gln
 180 185 190
 Ser Asp Ser His Ser Gly Glu Arg Arg Asn Pro Thr Ile Ser Ser Ser
 195 200 205
 Asp Ser Ile Asn Asn Arg Lys Arg Lys Gly Ala Gly Thr Glu Glu Ile
 210 215 220
 Leu Pro Gln Leu Ala Asn Arg Ser Arg His Ser
 225 230 235

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<400> 52
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<210> 53
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 ctacctagcc aggccaagga gcccaacgag ggggaacgct gcggtaggcg cgcgctgctc 120
 gtcccacccc acgcccgcct gacgggtgcc gatgccgagt cgcgcttcca tgcctctcgcg 180
 gtggacgaca gcctcgtcga caggaaactc atcgag atg ctg ctc aag acc tcg 234
 Met Leu Leu Lys Thr Ser
 1 5

tcc tac caa gtg acc acg gtg gat tcc ggg agc aag gcg ctg gag ctg 282
 Ser Tyr Gln Val Thr Val Asp Ser Gly Ser Lys Ala Leu Glu Leu
 10 15 20

ctg ggg ctg agg gac gcg tcg tcg ccg tct ccg tcc tcg cct gac cac 330
 Leu Gly Leu Arg Asp Ala Ser Ser Pro Ser Pro Ser Pro Asp His
 25 30 35

cag gag atc gac gtg aat ctc atc atc act gac tac tgc atg cca ggc 378
 Gln Glu Ile Asp Val Asn Leu Ile Ile Thr Asp Tyr Cys Met Pro Gly
 40 45 50

atg aca gga tac gat ctg ctc aag aga atg aag ggg tcc tct tcg ctc 426
 Met Thr Gly Tyr Asp Leu Leu Lys Arg Met Lys Gly Ser Ser Ser Leu
 55 60 65 70

aag gac att cct gtg gtg atc atg tcg tct gag aat gtg cct gcc cgg 474
 Lys Asp Ile Pro Val Val Ile Met Ser Ser Glu Asn Val Pro Ala Arg

75	80	85	
atc agc agg tgc ttg caa gac ggc	gcg gag gag ttc ttc ctg aag ccc		522
Ile Ser Arg Cys Leu Gln Asp Gly	Ala Glu Glu Phe Phe Leu Lys Pro		
90	95	100	
gtg aag ctg gcc gac atg aag aag ctc	aag tgc cac ctg ctg aaa cgg		570
Val Lys Leu Ala Asp Met Lys Lys	Leu Lys Ser His Leu Leu Lys Arg		
105	110	115	
aag cag ccc aag gag gcg cag gcg	cag gcg cag gcg cag gcg cag cag		618
Lys Gln Pro Lys Glu Ala Gln Ala	Gln Ala Gln Ala Gln Ala Gln Gln		
120	125	130	
gga cag gcg gtg gag ctg gag cct	gag cag cag ctg gac ccg cgc acg		666
Gly Gln Ala Val Glu Leu Glu Pro	Glu Gln Gln Leu Asp Pro Arg Thr		
135	140	145	150
cag ccg gcg cac gac gcg gag gaa	acc gcg gca gag ccg ccg ccg gcc		714
Gln Pro Ala His Asp Ala Glu Glu	Thr Ala Ala Glu Pro Pro Pro Ala		
155	160	165	
gca tcc aac gga acc acc gat ggc	ggc aac aag agg aag gcg gca gcc		762
Ala Ser Asn Gly Thr Thr Asp Gly	Gly Asn Lys Arg Lys Ala Ala Ala		
170	175	180	
atg gag gag gag ggg atg ctg gcc	gtg atg acg gtg gcg gcg ccg gag		810
Met Glu Glu Glu Gly Met Leu Ala	Val Met Thr Val Ala Ala Pro Glu		
185	190	195	
agc agc acc aag ccg agg ctg tcc	acc acc acc agc agc ctg gcg gtg		858
Ser Ser Thr Lys Pro Arg Leu Ser	Thr Thr Ser Ser Leu Ala Val		
200	205	210	
gaa acc tgagctgaga atcggacggc gaccggatca catattacta cctacgtaca			914
Glu Thr			
215			
ttaatccacc attgcgggca gtcgagaggc aaccaaccaa ccaagaacca accgacccaa			974
cccaggccac cgccgagaaa gagctcgctc tccctttctt tcgctgggtc ttttggttct			1034
cggagctaataaatggaatg aaggtgtgag ctccctgggga tggagggggcg gttacgggct			1094
ctaagtgaag aacaattacg acgtacctta cctagcggct gctgcttcgg ccggcaaat			1154
ttgttggtgct cgtgggtcatg gacacccttg tgctcgcttag gaggggggtcg ggggtgaacg			1214
gtgaaaaggga agctgtagat agaaaaggca ggcgtctcat gctcttggtg tctagtctaa			1274
tttttaattt tactcggctt attaaaaaaa aaaaaaaagg gcggccgc			1322

<210> 54

<211> 216

<212> PRT

<213> Zea mays

<400> 54

Met	Leu	Leu	Lys	Thr	Ser	Ser	Tyr	Gln	Val	Thr	Thr	Val	Asp	Ser	Gly
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Ser	Lys	Ala	Leu	Glu	Leu	Leu	Gly	Leu	Arg	Asp	Ala	Ser	Ser	Pro	Ser
			20				25					30			
Pro	Ser	Ser	Pro	Asp	His	Gln	Glu	Ile	Asp	Val	Asn	Leu	Ile	Ile	Thr
			35				40				45				
Asp	Tyr	Cys	Met	Pro	Gly	Met	Thr	Gly	Tyr	Asp	Leu	Leu	Lys	Arg	Met
	50					55				60					
Lys	Gly	Ser	Ser	Ser	Leu	Lys	Asp	Ile	Pro	Val	Val	Ile	Met	Ser	Ser
65					70				75					80	

Glu	Asn	Val	Pro	Ala	Arg	Ile	Ser	Arg	Cys	Leu	Gln	Asp	Gly	Ala	Glu
				85					90					95	
Glu	Phe	Phe	Leu	Lys	Pro	Val	Lys	Leu	Ala	Asp	Met	Lys	Lys	Leu	Lys
			100					105					110		
Ser	His	Leu	Leu	Lys	Arg	Lys	Gln	Pro	Lys	Glu	Ala	Gln	Ala	Gln	Ala
		115					120					125			
Gln	Ala	Gln	Ala	Gln	Gln	Gly	Gln	Ala	Val	Glu	Leu	Glu	Pro	Glu	Gln
		130				135					140				
Gln	Leu	Asp	Pro	Arg	Thr	Gln	Pro	Ala	His	Asp	Ala	Glu	Glu	Thr	Ala
145					150					155					160
Ala	Glu	Pro	Pro	Pro	Ala	Ala	Ser	Asn	Gly	Thr	Thr	Asp	Gly	Gly	Asn
			165						170					175	
Lys	Arg	Lys	Ala	Ala	Ala	Met	Glu	Glu	Glu	Gly	Met	Leu	Ala	Val	Met
			180					185					190		
Thr	Val	Ala	Ala	Pro	Glu	Ser	Ser	Thr	Lys	Pro	Arg	Leu	Ser	Thr	Thr
		195					200					205			
Thr	Ser	Ser	Leu	Ala	Val	Glu	Thr								
		210				215									

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<210> 55
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<212> DNA
<213> Zea mays
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<400> 55
atgctgctca agacctcgtc 20

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<210> 56
<211> 20
<212> DNA
<213> Zea mays
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<400> 56
tcagqtttcc accgccaggc 20

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<210> 57
<211> 1635
<212> DNA
<213> Zea mays
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<220>  
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Met Asp Phe Phe Thr Glu Tyr Gly Glu Gly Ser Arg Tyr Lys Ile Glu
1 5 10 15

gag gtc ata ggg aaa ggc agt tac ggt gtg gtt tgc tct gcc ttg gac 96
Glu Val Ile Gly Lys Gly Ser Tyr Gly Val Val Cys Ser Ala Leu Asp
20 25 30

act cat act ggt gaa aag gtc gcc ata aag aag ata aat gac atc ttt 144
Thr His Thr Gly Glu Lys Val Ala Ile Lys Lys Ile Asn Asp Ile Phe
35 40 45

gag cat gtg tct gat gcg aca agg ata ctt cgg gaa atc aag ttg ctt 192
Glu His Val Ser Asp Ala Thr Arg Ile Leu Arg Glu Ile Lys Leu Leu
50 55 60

aga ctc ctg cgg cat ccg gat atc gtg qaa ata aaq cat att cta ctt 240

Arg	Leu	Leu	Arg	His	Pro	Asp	Ile	Val	Glu	Ile	Lys	His	Ile	Leu	Leu	
65					70					75					80	
cct	cca	tca	agg	agg	gaa	ttc	agg	gat	att	tac	gtt	gtg	ttt	gaa	ctt	288
Pro	Pro	Ser	Arg	Arg	Glu	Phe	Arg	Asp	Ile	Tyr	Val	Val	Phe	Glu	Leu	
				85					90					95		
atg	gag	tct	gat	ttg	cac	caa	gtt	ata	aag	gct	aat	gat	gac	ttg	acc	336
Met	Glu	Ser	Asp	Leu	His	Gln	Val	Ile	Lys	Ala	Asn	Asp	Asp	Leu	Thr	
			100					105					110			
cca	gag	cat	tat	cag	ttc	ttc	ttg	tat	cag	ttg	ctt	cga	ggg	ttg	aaa	384
Pro	Glu	His	Tyr	Gln	Phe	Phe	Leu	Tyr	Gln	Leu	Leu	Arg	Gly	Leu	Lys	
		115					120					125				
tac	ata	cac	aca	gca	aat	gta	ttc	cat	cga	gat	ctc	aaa	cca	aaa	aat	432
Tyr	Ile	His	Thr	Ala	Asn	Val	Phe	His	Arg	Asp	Leu	Lys	Pro	Lys	Asn	
	130					135					140					
att	ctg	gca	aat	gct	gat	tgt	aag	ctc	aaa	ata	tgt	gat	ttt	ggc	cta	480
Ile	Leu	Ala	Asn	Ala	Asp	Cys	Lys	Leu	Lys	Ile	Cys	Asp	Phe	Gly	Leu	
145					150				155					160		
gca	aga	gta	gct	ttc	agt	gat	act	cca	acc	gcg	atc	ttt	tgg	aca	gat	528
Ala	Arg	Val	Ala	Phe	Ser	Asp	Thr	Pro	Thr	Ala	Ile	Phe	Trp	Thr	Asp	
				165				170					175			
tat	gtt	gca	act	agg	tgg	tac	cga	gca	ccg	gag	ctg	tgt	gga	tca	ttt	576
Tyr	Val	Ala	Thr	Arg	Trp	Tyr	Arg	Ala	Pro	Glu	Leu	Cys	Gly	Ser	Phe	
			180				185						190			
ttc	tca	aag	tat	aca	cca	gcg	ata	gat	ata	tgg	agt	att	ggg	tgt	ata	624
Phe	Ser	Lys	Tyr	Thr	Pro	Ala	Ile	Asp	Ile	Trp	Ser	Ile	Gly	Cys	Ile	
		195				200						205				
ttt	gcg	gaa	cta	tta	act	gga	aaa	ccg	ctt	ttc	ccg	ggg	aaa	aat	gtg	672
Phe	Ala	Glu	Leu	Leu	Thr	Gly	Lys	Pro	Leu	Phe	Pro	Gly	Lys	Asn	Val	
	210					215					220					
gta	cat	caa	ctt	gat	ata	att	aca	gat	ctc	atg	gga	aca	cct	tct	cca	720
Val	His	Gln	Leu	Asp	Ile	Ile	Thr	Asp	Leu	Met	Gly	Thr	Pro	Ser	Pro	
225					230				235						240	
gaa	gca	ata	tcg	agg	att	cgt	aat	gag	aag	gcc	agg	cgc	tac	ttg	agt	768
Glu	Ala	Ile	Ser	Arg	Ile	Arg	Asn	Glu	Lys	Ala	Arg	Arg	Tyr	Leu	Ser	
				245				250						255		
aat	atg	agg	cgg	aaa	aag	ccc	ata	cct	ttt	aca	cag	aag	ttc	cca	aat	816
Asn	Met	Arg	Arg	Lys	Lys	Pro	Ile	Pro	Phe	Thr	Gln	Lys	Phe	Pro	Asn	
			260					265					270			
gca	gat	cca	ctt	gca	tta	ggg	ttg	tta	gag	cga	atg	cta	gct	ttt	gag	864
Ala	Asp	Pro	Leu	Ala	Leu	Gly	Leu	Leu	Glu	Arg	Met	Leu	Ala	Phe	Glu	
		275				280						285				
cca	aaa	gac	agg	cca	agt	gct	gaa	gag	gct	ctt	gct	cat	cct	tat	ttt	912
Pro	Lys	Asp	Arg	Pro	Ser	Ala	Glu	Glu	Ala	Leu	Ala	His	Pro	Tyr	Phe	
	290					295				300						
aag	aat	ata	gcc	aat	gtg	gat	aga	gag	cct	tct	gca	caa	gct	gtc	aca	960
Lys	Asn	Ile	Ala	Asn	Val	Asp	Arg	Glu	Pro	Ser	Ala	Gln	Ala	Val	Thr	
305					310					315					320	

aag ctc gaa ttt gag ttc gag aga cgg aga gtc act aaa gat gac ata	1008
Lys Leu Glu Phe Glu Phe Glu Arg Arg Arg Val Thr Lys Asp Asp Ile	
325 330 335	
agg gaa ctt ata tat aga gaa att ctt gag tat cat cca aag atg ttg	1056
Arg Glu Leu Ile Tyr Arg Glu Ile Leu Glu Tyr His Pro Lys Met Leu	
340 345 350	
aaa gaa ttt att gag gga act gag tca agt ggt ttc atg tac cca agt	1104
Lys Glu Phe Ile Glu Gly Thr Glu Ser Ser Gly Phe Met Tyr Pro Ser	
355 360 365	
gca gta gat aat ttt aaa aag cag ttt gca tac ctt gag gaa cat tat	1152
Ala Val Asp Asn Phe Lys Lys Gln Phe Ala Tyr Leu Glu Glu His Tyr	
370 375 380	
gcc aag gga tct acg gga act cca cct gag agg caa cat aac tca ttg	1200
Ala Lys Gly Ser Thr Gly Thr Pro Pro Glu Arg Gln His Asn Ser Leu	
385 390 395 400	
cca cgg cct tct gtt gtc tat tca gac aac caa tca caa agc acg gcc	1248
Pro Arg Pro Ser Val Val Tyr Ser Asp Asn Gln Ser Gln Ser Thr Ala	
405 410 415	
aat ata aca gag gat ctt tcc aag tgt ata atc aga gaa aat gca cag	1296
Asn Ile Thr Glu Asp Leu Ser Lys Cys Ile Ile Arg Glu Asn Ala Gln	
420 425 430	
aag tca cag caa tat aat gct tca gtt gca aat aaa ttt cca tcc cat	1344
Lys Ser Gln Gln Tyr Asn Ala Ser Val Ala Asn Lys Phe Pro Ser His	
435 440 445	
gtt cct caa ggt gct gct gca agg cct ggt aaa gca gtt ggt tcg gtg	1392
Val Pro Gln Gly Ala Ala Ala Arg Pro Gly Lys Ala Val Gly Ser Val	
450 455 460	
ctg cag tac aac ctc tct caa gca tct gct gct gag caa tat gag cag	1440
Leu Gln Tyr Asn Leu Ser Gln Ala Ser Ala Ala Glu Gln Tyr Glu Gln	
465 470 475 480	
cga agg gtt gct agg cac cca gca gtt gcc cca aac aac atc cct tct	1488
Arg Arg Val Ala Arg His Pro Ala Val Ala Pro Asn Asn Ile Pro Ser	
485 490 495	
gga agc tca tac ccc aga aga aat cag acc tgc aag agc gag aca ggc	1536
Gly Ser Ser Tyr Pro Arg Arg Asn Gln Thr Cys Lys Ser Glu Thr Gly	
500 505 510	
gac act gag agg atg gac gtg aac caa gct ggg cag ccg aag tcc tat	1584
Asp Thr Glu Arg Met Asp Val Asn Gln Ala Gly Gln Pro Lys Ser Tyr	
515 520 525	
gca gca aac aaa cta cct gca act gtt gat ggt cgc ggc ggc cat tgg	1632
Ala Ala Asn Lys Leu Pro Ala Thr Val Asp Gly Arg Gly Gly His Trp	
530 535 540	
tag	1635
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 <211> 544
 <212> PRT
 <213> Zea mays

<400> 58
 Met Asp Phe Phe Thr Glu Tyr Gly Glu Gly Ser Arg Tyr Lys Ile Glu
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 Glu Val Ile Gly Lys Gly Ser Tyr Gly Val Val Cys Ser Ala Leu Asp
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 Thr His Thr Gly Glu Lys Val Ala Ile Lys Lys Ile Asn Asp Ile Phe
 35 40 45
 Glu His Val Ser Asp Ala Thr Arg Ile Leu Arg Glu Ile Lys Leu Leu
 50 55 60
 Arg Leu Leu Arg His Pro Asp Ile Val Glu Ile Lys His Ile Leu Leu
 65 70 75 80
 Pro Pro Ser Arg Arg Glu Phe Arg Asp Ile Tyr Val Val Phe Glu Leu
 85 90 95
 Met Glu Ser Asp Leu His Gln Val Ile Lys Ala Asn Asp Asp Leu Thr
 100 105 110
 Pro Glu His Tyr Gln Phe Phe Leu Tyr Gln Leu Leu Arg Gly Leu Lys
 115 120 125
 Tyr Ile His Thr Ala Asn Val Phe His Arg Asp Leu Lys Pro Lys Asn
 130 135 140
 Ile Leu Ala Asn Ala Asp Cys Lys Leu Lys Ile Cys Asp Phe Gly Leu
 145 150 155 160
 Ala Arg Val Ala Phe Ser Asp Thr Pro Thr Ala Ile Phe Trp Thr Asp
 165 170 175
 Tyr Val Ala Thr Arg Trp Tyr Arg Ala Pro Glu Leu Cys Gly Ser Phe
 180 185 190
 Phe Ser Lys Tyr Thr Pro Ala Ile Asp Ile Trp Ser Ile Gly Cys Ile
 195 200 205
 Phe Ala Glu Leu Leu Thr Gly Lys Pro Leu Phe Pro Gly Lys Asn Val
 210 215 220
 Val His Gln Leu Asp Ile Ile Thr Asp Leu Met Gly Thr Pro Ser Pro
 225 230 235 240
 Glu Ala Ile Ser Arg Ile Arg Asn Glu Lys Ala Arg Arg Tyr Leu Ser
 245 250 255
 Asn Met Arg Arg Lys Lys Pro Ile Pro Phe Thr Gln Lys Phe Pro Asn
 260 265 270
 Ala Asp Pro Leu Ala Leu Gly Leu Leu Glu Arg Met Leu Ala Phe Glu
 275 280 285
 Pro Lys Asp Arg Pro Ser Ala Glu Glu Ala Leu Ala His Pro Tyr Phe
 290 295 300
 Lys Asn Ile Ala Asn Val Asp Arg Glu Pro Ser Ala Gln Ala Val Thr
 305 310 315 320
 Lys Leu Glu Phe Glu Phe Glu Arg Arg Arg Val Thr Lys Asp Asp Ile
 325 330 335
 Arg Glu Leu Ile Tyr Arg Glu Ile Leu Glu Tyr His Pro Lys Met Leu
 340 345 350
 Lys Glu Phe Ile Glu Gly Thr Glu Ser Ser Gly Phe Met Tyr Pro Ser
 355 360 365
 Ala Val Asp Asn Phe Lys Lys Gln Phe Ala Tyr Leu Glu Glu His Tyr
 370 375 380
 Ala Lys Gly Ser Thr Gly Thr Pro Pro Glu Arg Gln His Asn Ser Leu
 385 390 395 400
 Pro Arg Pro Ser Val Val Tyr Ser Asp Asn Gln Ser Gln Ser Thr Ala
 405 410 415
 Asn Ile Thr Glu Asp Leu Ser Lys Cys Ile Ile Arg Glu Asn Ala Gln
 420 425 430
 Lys Ser Gln Gln Tyr Asn Ala Ser Val Ala Asn Lys Phe Pro Ser His
 435 440 445

Val Pro Gln Gly Ala Ala Ala Arg Pro Gly Lys Ala Val Gly Ser Val
 450 455 460
 Leu Gln Tyr Asn Leu Ser Gln Ala Ser Ala Ala Glu Gln Tyr Glu Gln
 465 470 475 480
 Arg Arg Val Ala Arg His Pro Ala Val Ala Pro Asn Asn Ile Pro Ser
 485 490 495
 Gly Ser Ser Tyr Pro Arg Arg Asn Gln Thr Cys Lys Ser Glu Thr Gly
 500 505 510
 Asp Thr Glu Arg Met Asp Val Asn Gln Ala Gly Gln Pro Lys Ser Tyr
 515 520 525
 Ala Ala Asn Lys Leu Pro Ala Thr Val Asp Gly Arg Gly Gly His Trp
 530 535 540

<210> 59
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 <212> DNA
 <213> Zea mays

<400> 59
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<400> 60
 tcaatgcgaa accaaattta 20

<210> 61
 <211> 4014
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gac aac aaa tac atg ctt gga gat gaa ata ggg aag ggg gcg tac ggc 96
 Asp Asn Lys Tyr Met Leu Gly Asp Glu Ile Gly Lys Gly Ala Tyr Gly
 20 25 30

cgc gta tac aag ggg ctt gac ctg gag aat ggc gat ttc gtg gcc atc 144
 Arg Val Tyr Lys Gly Leu Asp Leu Glu Asn Gly Asp Phe Val Ala Ile
 35 40 45

aaa cag gtc tcg ctg gag aac att ccg cag gag gat ctc aac ata ata 192
 Lys Gln Val Ser Leu Glu Asn Ile Pro Gln Glu Asp Leu Asn Ile Ile
 50 55 60

atg caa gag atc gac ctt ttg aaa aat ctt aat cat aaa aat att gtc 240
 Met Gln Glu Ile Asp Leu Leu Lys Asn Leu Asn His Lys Asn Ile Val
 65 70 75 80

aag tat ttg gga tca ttg aag aca aag agc cac ctc cat att att ttg 288
 Lys Tyr Leu Gly Ser Leu Lys Thr Lys Ser His Leu His Ile Ile Leu
 85 90 95

gag tat gtg gag aat ggc tca ctt gct aat att atc aag cca aac aaa	336
Glu Tyr Val Glu Asn Gly Ser Leu Ala Asn Ile Ile Lys Pro Asn Lys	
100 105 110	
ttc gga cct ttt cct gaa tct ttg gtg gct gta tac att gct cag gtg	384
Phe Gly Pro Phe Pro Glu Ser Leu Val Ala Val Tyr Ile Ala Gln Val	
115 120 125	
ttg gaa ggt ctt gtt tat ctg cat gaa caa ggt gtc att cat aga gat	432
Leu Glu Gly Leu Val Tyr Leu His Glu Gln Gly Val Ile His Arg Asp	
130 135 140	
atc aag ggc gca aat ata ctg act acc aaa gag ggc ctt gtc aaa ctt	480
Ile Lys Gly Ala Asn Ile Leu Thr Thr Lys Glu Gly Leu Val Lys Leu	
145 150 155 160	
gct gac ttt gga gtt gcc act aaa ttg act gaa gct gac atc aac act	528
Ala Asp Phe Gly Val Ala Thr Lys Leu Thr Glu Ala Asp Ile Asn Thr	
165 170 175	
cat tct gtg gtc ggc act cca tac tgg atg gcg cct gag gtc atc gaa	576
His Ser Val Val Gly Thr Pro Tyr Trp Met Ala Pro Glu Val Ile Glu	
180 185 190	
atg tct ggt gtt tgt gct gcc tct gat atc tgg agt gtg ggt tgc acc	624
Met Ser Gly Val Cys Ala Ala Ser Asp Ile Trp Ser Val Gly Cys Thr	
195 200 205	
gta att gaa ttg ctg aca tgt gtc ccg cca tat tat gat ctc caa cct	672
Val Ile Glu Leu Leu Thr Cys Val Pro Pro Tyr Tyr Asp Leu Gln Pro	
210 215 220	
atg cct gcc ctg ttc cgt att gtt cag gat gtg cat cca cca ata cca	720
Met Pro Ala Leu Phe Arg Ile Val Gln Asp Val His Pro Pro Ile Pro	
225 230 235 240	
gaa gga ctg tca cct gag att act gat ttt ctc cgg caa tgt ttt caa	768
Glu Gly Leu Ser Pro Glu Ile Thr Asp Phe Leu Arg Gln Cys Phe Gln	
245 250 255	
aag gat gcg atg caa agg cct gat gcg aag aca tta ttg atg cat cca	816
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Asn Asn Ser Gly Phe Cys Asp Thr Pro Gly Asp Thr Arg Ala Thr Ile	
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Cys Gln Lys Leu Leu Ser Ile Phe Ser His Arg Pro Glu Gln Lys Gln	
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Met	Thr	Gly	Lys	Asn	Arg	Ser	Ala	Ala	Ile	Val	Ala	Ala	Phe	Leu	Met		
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 poly A tail but no cDNA.

<400> 69
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36

(12) INTERNATIONAL APPLICATION PUBLISHED UNDER THE PATENT COOPERATION TREATY (PCT)

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23 November 2000 (23.11.2000)

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WO 00/70059 A3

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- (71) Applicant (for all designated States except US): **PIO-NEER HI-BRED INTERNATIONAL, INC.** [US/US]; 800 Capital Square, 400 Locust Street, Des Moines, IA 50309 (US).
- (72) Inventor; and
- (75) Inventor/Applicant (for US only): **HELENTJARIS, Timothy, G.** [US/US]; 2960 NW 73rd Lane, Ankeny, IA 50021 (US).
- (74) Agents: **VARLEY, Karen, K.** et al.; 7100 N.W. 62nd Avenue, Darwin Building, Johnston, IA 50131-1000 (US).
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- For two-letter codes and other abbreviations, refer to the "Guidance Notes on Codes and Abbreviations" appearing at the beginning of each regular issue of the PCT Gazette.



WO 00/70059 A3

(54) Title: SIGNAL TRANSDUCTION GENES AND METHODS OF USE

(57) Abstract: The invention provides isolated signal transduction nucleic acids and their encoded proteins. The present invention provides methods and compositions relating to altering signal transduction gene expression levels in plants. The invention further provides recombinant expression cassettes, host cells, transgenic plants, and antibody compositions.

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 00/11687

A. CLASSIFICATION OF SUBJECT MATTER

IPC 7 C12N15/54 C12N15/29 C12N15/82 C12N9/12 A01H5/00

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 7 C12N A01H

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

WPI Data, BIOSIS, EPO-Internal, EMBL, PAJ

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☒ Further documents are listed in the continuation of box C.☒ Patent family members are listed in annex.

* Special categories of cited documents:

- "A" document defining the general state of the art which is not considered to be of particular relevance
- "E" earlier document but published on or after the international filing date
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- "O" document referring to an oral disclosure, use, exhibition or other means
- "P" document published prior to the international filing date but later than the priority date claimed

- "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
- "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
- "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.
- "&" document member of the same patent family

Date of the actual completion of the international search

18 December 2000

Date of mailing of the international search report

22 12 00

Name and mailing address of the ISA

European Patent Office, P.B. 5818 Patentlaan 2
NL - 2280 HV Rijswijk
Tel. (+31-70) 340-2040, Tx. 31 651 epo nl,
Fax: (+31-70) 340-3016

Authorized officer

Maddox, A

INTERNATIONAL SEARCH REPORT

 Intel Application No
 PCT/US 00/11687

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
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Y	& DATABASE EMBL [Online] ACCESSION NO:C22570, 15 August 1997 (1997-08-15) SASAKI, T., ET AL.: "Rice cDNA, partial sequence (E40447_2A)." abstract ---	2-12
X	DATABASE EMBL [Online] ACCESSION NO:Y14316, 4 August 1997 (1997-08-04) HENRY Y.: "Arabidopsis thaliana mRNA for MAP3K gamma protein kinase" XP002147373 the whole document -& JOUANNIC, S., ET AL.: "Characterisation of novel plant genes encoding MEKK/STE11 and RAF-related protein kinases." GENE, vol. 229, 18 March 1999 (1999-03-18), pages 171-181, XP004161172 the whole document ---	1,13
Y	--- -/--	2-12

INTERNATIONAL SEARCH REPORT

 Inter: 31 Application No
 PCT/US 00/11687

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
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P,X	DATABASE EMBL [Online] ACCESSION NO: AI668456, 17 May 1999 (1999-05-17) WALBOT, V., ET AL.: "mRNA sequence." XP002155765 the whole document ---	1,13
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Form PCT/ISA/210 (continuation of second sheet) (July 1992)

INTERNATIONAL SEARCH REPORT

Intern Application No
PCT/US 00/11687

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
P,X	<p>DATABASE WPI Section Ch, Week 200034 Derwent Publications Ltd., London, GB; Class C06, AN 2000-389819 XP002155767 & JP 2000 116260 A (BIO-ORIENTED TECHNOLOGY RES ADVANCEMENT), 25 April 2000 (2000-04-25) abstract & DATABASE GENESEQ [Online] ACCESSION NO: A53412, 4 October 2000 (2000-10-04) abstract</p>	1-3,13
P,X	<p>--- MENKE FRANK L H ET AL: "A novel jasmonate- and elicitor-responsive element in the periwinkle secondary metabolite biosynthetic gene Str interacts with a jasmonate- and elicitor-inducible AP2-domain transcription factor, ORCA2." EMBO (EUROPEAN MOLECULAR BIOLOGY ORGANIZATION) JOURNAL, vol. 18, no. 16, 16 August 1999 (1999-08-16), pages 4455-4463, XP002155761 ISSN: 0261-4189 figure 4</p>	13
P,X	<p>--- WO 00 08187 A (VERBRUGGEN NATHALIE ;VLAAMS INTERUNIV INST BIOTECH (BE); LEE JEONG) 17 February 2000 (2000-02-17) see SEQ ID NO:33 page 143 -page 146</p>	1-13
P,X	<p>--- WO 00 09724 A (GEN HOSPITAL CORP) 24 February 2000 (2000-02-24) see figs 14 and 15 the whole document</p>	1-13
E	<p>--- WO 00 46383 A (UNIV LEIDEN ;MEMELINK JOHAN (NL); FITS CORNELIA THEODORA ELISABE ()) 10 August 2000 (2000-08-10) see SEQ ID NO:1</p>	1,13
E	<p>--- EP 1 033 405 A (CERES INC) 6 September 2000 (2000-09-06) see SEQ ID NOS:3964 and 3965</p>	1,13
A	<p>--- DATABASE EMBL [Online] ACCESSION NO:AB016801, 9 February 1999 (1999-02-09) KUSANO T.: "Zea mays mRNA for MAP kinase 4, complete cds." XP002147375 the whole document</p> <p>---</p> <p>-/--</p>	1-13

INTERNATIONAL SEARCH REPORT

Inte Application No
PCT/US 00/11687

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	<p>DATABASE EMBL [Online] ACCESSION NO:AB016802, 9 February 1999 (1999-02-09) KUSANO T.: "Zea mays mRNA for MAP kinase 5, complete cds." XP002147376 the whole document</p> <p>---</p>	1-13
A	<p>HARDIN SHANE C ET AL: "Molecular cloning and characterization of maize ZmMEK1, a protein kinase with a catalytic domain homologous to mitogen- and stress-activated protein kinase kinases." PLANTA (BERLIN), vol. 206, no. 4, November 1998 (1998-11), pages 577-584, XP000939378 ISSN: 0032-0935 the whole document & DATABASE EMBL [Online] ACCESSION NO:U83625, 8 January 1998 (1998-01-08)</p> <p>---</p>	1-13
A	<p>DATABASE WPI Section Ch, Week 199720 Derwent Publications Ltd., London, GB; Class B04, AN 1997-220416 XP002079552 & JP 09 065881 A (NORINSUISANSHO NOGYO SEIBUTSU SHIGEN), 11 March 1997 (1997-03-11) abstract -& DATABASE GENESEQ [Online] ACCESSION NO:T60349, 4 June 1997 (1997-06-04) "MAP kinase #1 coding sequence" XP002079541 the whole document -& DATABASE GENESEQ [Online] ACCESSION NO:W15512, 4 June 1997 (1997-06-04) "MAP kinase #1" XP002079540 the whole document</p> <p>---</p>	1-13
A	<p>SEO ET AL: "Tobacco MAP kinase: a possible mediator in wound signal transduction pathways" SCIENCE, AMERICAN ASSOCIATION FOR THE ADVANCEMENT OF SCIENCE, US, vol. 270, 22 December 1995 (1995-12-22), pages 1988-1992, XP002079542 ISSN: 0036-8075 the whole document</p> <p>---</p>	1-13

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INTERNATIONAL SEARCH REPORT

Inter If Application No
PCT/US 00/11687

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT

PCT/US 00/11687

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
A	LIGTERINK ET AL: "Receptor-mediated activation of a MAP kinase in pathogen defense of plants" SCIENCE,US,AMERICAN ASSOCIATION FOR THE ADVANCEMENT OF SCIENCE,, vol. 276, 27 June 1997 (1997-06-27), pages 2054-2057, XP002079548 ISSN: 0036-8075 the whole document -----	1-13

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US 00/11687

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This International Search Report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:

2. ☐ Claims Nos.:
because they relate to parts of the International Application that do not comply with the prescribed requirements to such an extent that no meaningful International Search can be carried out, specifically:

3. ☐ Claims Nos.:
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

see additional sheet

1. ☐ As all required additional search fees were timely paid by the applicant, this International Search Report covers all searchable claims.

2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.

3. ☒ As only some of the required additional search fees were timely paid by the applicant, this International Search Report covers only those claims for which fees were paid, specifically claims Nos.:
1-13 all partially, subject groups 1 and 3.

4. ☐ No required additional search fees were timely paid by the applicant. Consequently, this International Search Report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
- ☒ No protest accompanied the payment of additional search fees.

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

This International Searching Authority found multiple (groups of) inventions in this international application, as follows:

1. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:1 and 2, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:1 and 2 and said related polynucleotides and proteins.

2. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:5 and 6, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:5 and 6 and said related polynucleotides and proteins

3. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:9 and 10, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:9 and 10 and said related polynucleotides and proteins.

4. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:13 and 14, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:13 and 14 and said related polynucleotides and proteins.

5. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:17 and 18, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:17 and 18 and said

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

related polynucleotides and proteins.

6. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:21 and 22, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:21 and 22 and said related polynucleotides and proteins.

7. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:25 and 26, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:25 and 26 and said related polynucleotides and proteins.

8. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:29 and 30, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:29 and 30 and said related polynucleotides and proteins.

9. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:33 and 34, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:33 and 34 and said related polynucleotides and proteins.

10. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:37 and 38, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:37 and 38 and said

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

related polynucleotides and proteins.

11. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:41 and 42, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:41 and 42 and said related polynucleotides and proteins.

12. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:45 and 46, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:45 and 46 and said related polynucleotides and proteins.

13. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:49 and 50, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:49 and 50 and said related polynucleotides and proteins.

14. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:53 and 54, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:53 and 54 and said related polynucleotides and proteins.

15. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:57 and 58, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:57 and 58 and said

INTERNATIONAL SEARCH REPORT

International Application No. PCT/US 00/11687

FURTHER INFORMATION CONTINUED FROM PCT/ISA/ 210

related polynucleotides and proteins.

16. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:61 and 62, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:61 and 62 and said related polynucleotides and proteins.

17. Claims: 1-13 all partially

Maize nucleic acid and corresponding protein as characterised by SEQ ID NOS:65 and 66, polynucleotides and proteins related to said sequences as defined by the claims, recombinant expression cassettes, host cells, transgenic plants, methods for modulation of signal transduction gene expression, based on said SEQ ID NOS:65 and 66 and said related polynucleotides and proteins.

INTERNATIONAL SEARCH REPORT

Information on patent family members

Inter ai Application No

PCT/US 00/11687

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
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STANDARD FORM NO. 64